RESEARCH Open Access

Constitutively active androgen receptor supports the metastatic phenotype of endocrine-resistant hormone receptor-positive breast cancer



Shaymaa Bahnassy¹, Hariprasad Thangavel², Maram Quttina¹, Ashfia Fatima Khan¹, Dhanya Dhanyalayam¹, Joan Ritho³, Samaneh Karami¹, Jing Ren⁴ and Tasneem Bawa-Khalfe^{1*}

Abstract

Background: Hormone receptor positive (HR+) breast cancer (BCa) is the most frequently diagnosed subtype. Acquired and intrinsic resistance to conventional endocrine therapy (ET) commonly occurs and prompts incurable metastatic disease. Hence, ET-resistant (ET-R) HR+ BCa presents a therapeutic challenge. Previous studies show elevated androgen receptor (AR) that supports resistance to ET tamoxifen and correlates with HR+ BCa metastasis. Yet surprisingly, studies with AR-blocker enzalutamide (Enz) in ET-R HR+ BCa present conflicting results. We now report that a constitutively active, unique from canonical Enz-targeted, AR accumulates in endocrine resistant HR+ BCa cells.

Methods: AR protein profiles in acquired and intrinsic ET-R HR + -BCa were defined with cell-free modification tests, in-house in-vivo SUMOylation assays, and PLA imaging. Genomic activity of native AR and modified-AR mimetic was tested with reporter assays and limited transcriptome analysis. Spheroid growth and migration studies were used to evaluate inhibitory actions of Enz and combinatorial therapy.

Results: Sustained higher molecular weight SUMO-modified AR (SUMO-AR) persists in acquired and intrinsic ET-R BCa cell lines. Concurrently, SUMO isoforms and global SUMO-modified proteome also accumulates in the same cell lines. We identified AR as a novel substrate for the SUMO-E3 ligase HSPB1/Hsp27. Independent of ligand, SUMO-AR is resilient to ubiquitin-mediated proteasomal degradation, enriched in the nucleus, readily chromatin-bound, and transcriptionally active. Constitutive SUMO-AR initiates a gene-expression profile that favors epithelial-mesenchymal transition. Enz combined with a SUMO inhibitor attenuates migration and metastatic phenotype of ET-R HR+ BCa.

Conclusion: Targeting both unmodified and SUMO-modified AR prevents the metastatic progression of HR+ BCa with FT-R

Keywords: HR+ breast cancer, Endocrine resistance, Androgen receptor, SUMO, Enzalutamide

¹Center for Nuclear Receptors & Cell Signaling, Department of Biology & Biochemistry, University of Houston, 3517 Cullen Blvd, SERC Bldg, Rm 3010, Houston, TX 77204-5056, USA

Full list of author information is available at the end of the article



© The Author(s). 2020 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

^{*} Correspondence: tbawa-khalfe@uh.edu

Background

Endocrine therapy (ET) remains the first line cancer therapy for early-stage hormone-receptor positive (HR+) breast cancers (BCa). The selective estrogen receptor modulator (SERM), Tamoxifen (Tam) is effective for pre –/peri-menopausal women while post-menopausal women receive anti-estrogen aromatase inhibitors (AI). However, intrinsic or acquired ET-resistance is common and equates to rapid onset of malignant metastatic disease. Hence, HR+ BCa unresponsive to conventional ET poses a new challenge with no targeted therapy available to prevent cancer progression.

The androgen receptor (AR) is emerging as a good therapeutic target for other advanced BCa subtypes [1]. Canonical AR functions with ligand androgen binding, subsequently translocate form the cytosol to the nucleus, associates with androgen response elements (ARE) in gene enhancer and/or promoter regions, and transcriptionally regulates corresponding genes [2]. This liganddriven transcription factor is widely expressed in primary and metastatic breast tumors and specifically, 75% of HR+ BCa tissue samples express AR [1, 3]. Higher AR expression in HR+-patients correlates with reduced clinical response to ET tamoxifen and worse prognosis with greater cancer recurrence [4]. Induction of AR initiates Tam-resistance (TamR) in HR+ BCa cell models and promotes tumor growth in mouse xenografts [5], highlighting a role for AR in ET-R and disease progression [4–6]. However, previous studies using a secondgeneration AR antagonist Enzalutamide (Enz) in ET-R HR+ BCa present conflicting results. Treatment of MCF7-TamR cells with Enz, reduced growth and colony formation in one study [6] but showed no effect in another study [7]. In addition, Enz failed to reduce tumor growth of the HR+ ET-R patient-derived xenograft (HCI-13) [8]. In clinical trials, positive response of HR+ BCa patients to Enz is dependent on prior ET history. While combinatorial Enz and AI therapy improved progression-free survival for women with advanced HR+ BCa without prior ET history, patients previously on ET did not respond to the same Enz-AI combination [9]. Yet, ongoing clinical trials continue evaluating Enz alone or in combination with other endocrine therapies (NCT02676986, NCT02953860 and NCT02955394). This raises the question: how is the AR different in HR+ BCa patients previously treated with ET?

The AR is subject to several post-translational modifications (PTM) including Small Ubiquitin Like Modifier (SUMO)-PTM [10, 11]. Protein SUMOylation is the reversible covalent binding of SUMO to a lysine residue within a specific consensus sequence ψ KXE (ψ is a hydrophobic residue and X is any amino acid) [12]. Upon ligand activation, AR residues K386 and K520 are targets for the 3 SUMO paralogs SUMO-1, – 2, and – 3

[13, 14]. Also, SUMO enzymatic components like deSU-MOylases and SUMO-E3 ligases dictate the level of SUMOylated AR. Previously we demonstrated that elevated deSUMOylase SENP1 prevents accumulation of SUMOylated AR in prostate cancer [15–17]. This is important as high-throughput AR functional studies show that SUMO-PTM regulates the chromatin occupancy and transcriptional activity of ligand-activated AR in a gene dependent manner [18].

We now report that high levels of SUMOylated AR persist independent of AR-ligand in ET-R HR+ BCa cells. In these BCa lines, SUMO-PTM supports constitutive AR genomic activity to promote metastatic transformation. Concurrent targeting of SUMOylated AR improves the inhibitory actions of Enz on the drugresistant HR+ BCa.

Methods

Cell culture and transfections

Sensitive (MCF-7) and resistant (derived MCF-7-TamR or intrinsically resistant GI-101A and GILM2) human breast carcinoma cell lines to ET were cultured in Dulbecco's modified Eagle's medium (DMEM; Gibco) supplemented with 10% FBS and 1% penicillin/streptomycin at 37 °C in a humidified atmosphere containing 5% CO $_2$. MCF-7-TamR cells were generated in our lab by chronically treating MCF-7 with 1 μ M 4-OH-tamoxifen (Sigma-Aldrich) for over 6 months. After developing resistance, the derived cells were used and continuously cultivated in presence of tamoxifen. All transient transfections were performed with Lipofectamine 2000 (Invitrogen) according to manufacturer instructions.

Immunoprecipitation (IP) and Chromatin-Immunoprecipitation (CB/IP) assays

After harvesting and lysing the cells in a buffer containing protease inhibitor (Roche) and *N*-ethylmaleimide (Sigma-Aldrich), proteins were either immunoprecipitated from whole-cells or chromatin fractions using the same protocol described before [19]. Immunoprecipitated proteins were then bound to Protein A-agarose beads (Millipore) and samples were resolved on SDS-PAGE gels and immunoblotted with specific antibodies.

SDS-page and Western blot analysis

Whole-cell or immunoprecipitated protein extracts were denatured, resolved on SDS-PAGE gels and transferred onto PVDF membranes (BioRad) using a wet transfer apparatus. After blocking, blots were probed overnight and the following primary antibodies were used: AR (sc-816), Ubiquitin (sc-8017) and GAPDH (sc-8017) from Santa Cruz Biotechnology, AR441 (MA5–13426) and Hsp27 (MA3–015) from ThermoFisher, SUMO1 (49, 305, Cell Signaling), SUMO2/3 (ab81371, Abcam),

PIAS1 (49,305, Cell Signaling), and β -actin (Sigma-Aldrich). Antigen–antibody complexes were then detected by the chemiluminescence Western Lighting Plus-ECL reagent (PerkinElmer).

Statistical analysis

All results are expressed as mean \pm SEM. Prism 7 (GraphPad Software) was used for data analysis. Student's two-tailed *t-test* or one-way ANOVA followed by Tukey's multiple comparison test was employed to evaluate statistical significance between groups and *p*-values < 0.05 were considered statistically significant.

Remaining methods used in this study are found in Additional file 1: supplemental materials and methods.

Results

SUMO-modified AR persists in ET-R BCa

Elevated RNA levels of AR were previously reported in metastatic TamR BCa tumors [5]. We analyzed AR expression in publicly available survival datasets from the Kaplan-Meier Plotter database [20], and interestingly observed a reversed pattern in distant-metastasis free

survival (DMFS) of total versus Tam-treated HR+ BCa patients (Fig. 1a-b). Specifically, patients with high AR levels exhibit a better prognosis with lower probability to develop metastasis; inversely patients treated with Tam have a worse prognosis with higher susceptibility for metastatic onset (log-rank p = 0.0076, HR = 0.56 versus p = 0.18, HR = 2.21, respectively Fig. 1a-b).

Consistent with a previous report [7], both long-term estrogen-deprived cells (EDR-7) that mimic AI resistant HR+ BCa, and acquired TamR-7 cells, generated with continuous culturing of MCF-7 cells with 1 μM 4-OH-tamoxifen, express higher protein levels of unmodified AR at $\sim 100 \, \text{kDa}$ when compared with endocrine therapy-sensitive (ET-S) parental MCF-7 cells (Fig. 1c). In addition, we observe two slowly migrating higher molecular weight bands (modified AR) elevated in both EDR-7 and TamR-7 along with the unmodified AR. Intrinsic TamR-BCa cell lines, GI-101A and its highly metastatic variant GILM2 [21, 22], also express the modified but not unmodified AR (Fig. 1c). Previous AR SUMOylation studies demonstrate analogous higher molecular weight bands corresponding to the two major

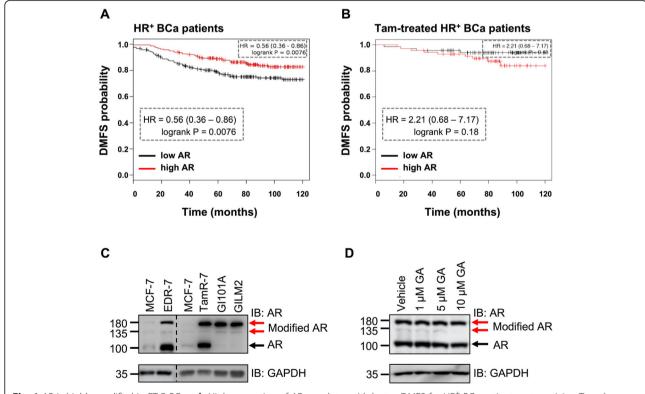


Fig. 1 AR is highly modified in ET-R BCa. **a-b** High expression of AR correlates with better DMFS for HR⁺ BCa patients not receiving Tam therapy. Generated plots from KM Plotter database for BCa (www.kmplot.com) illustrate 10-year DMFS for: (**a**) HR+-BCa and (**b**) Tam-treated HR+-BCa patients. Plots separate high and low gene expressions. Both HR, representing hazard ratio at 95% confidence, and log-rank *P* values are enlarged in the dotted box. **c** Endogenous AR protein is highly modified in long-term estrogen deprived (EDR-7), acquired (TamR-7) and intrinsic TamR-BCa cell lines (GI-101A and GILM2). Blot represents three independent experiments; arrows indicate modified and unmodified AR. **d** Ginkgolic acid (GA) inhibits SUMO-modification of AR in a dose-dependent manner. TamR-7 cells were treated with increasing concentrations of GA and immunoblot represents two independent experiments

SUMO acceptor sites of AR at lysine residues 386 and 520 [13]. To test if the modified AR in TamR cells is SUMOylated, TamR-7 cells were treated with ginkgolic acid (GA) to block the activity of SUMO-E1 and inhibit protein SUMOylation [23]. Increasing GA treatment reduced high molecular weight AR (Fig. 1d).

A hyperSUMO environment exists in TamR-BCa

We postulated that altered expression of SUMO paralogs and/or enzymatic components could support elevated levels of SUMOylated AR in drug resistant BCa cells. In TamR-7 versus TamS cells, transcript levels of all three SUMO isoforms significantly increased (Fig. 2a). In addition, we included components of the SUMO-enzymatic machinery that SUMOylate and/or interact with AR based on published literature and high-throughput screens [24–27]. SUMO-specific activating E1-SAE1/SAE2 dimers, conjugating E2 Ubc9, ligating E3 PIAS1, and deSUMOylase SENP1 enzymes are equivalently transcribed in TamS and TamR-7 cells (Fig. 2a). In contrast, the RNA levels for a canonical AR binding partner with proposed SUMO-E3 activity for other

substrates Hsp27 is upregulated (p < 0.05; Additional file 1: Fig. S1A). To evaluate whether transcript changes correlate with disease progression, publicly available datasets were analyzed with KM plotter. Specifically, survival data shows that high SUMO levels directly correlate with high probability of metastasis in TamR-BCa patients (log-rank p = 0.027; Additional file 1: Fig. S1B). Moreover, Tam treated HR⁺-BCa patients with concurrent elevated levels of AR and SUMO exhibit a greater risk for developing metastasis (log-rank p = 0.024, HR = 4.85 in Tam-treated versus p = 0.35, HR = 1.59 for total HR⁺ patients, respectively Fig. 2c and b). Clearly ET treatment of HR+ BCa supports unique gene expression of the SUMO system.

Next, we determined if this induction of SUMO proteins equate to changes in the SUMO-modified proteome. As compared to noncancerous MCF10-2A cells, SUMO-1 and SUMO2/3 conjugated proteins are greatly enriched in all HR+ BCa lines (Fig. 2d and e). Interestingly, SUMO2/3 conjugates are favored in TamR HR+ BCa; specifically, acquired TamR-7 cells and intrinsic TamR GI-101A and GILM2 cells express greater

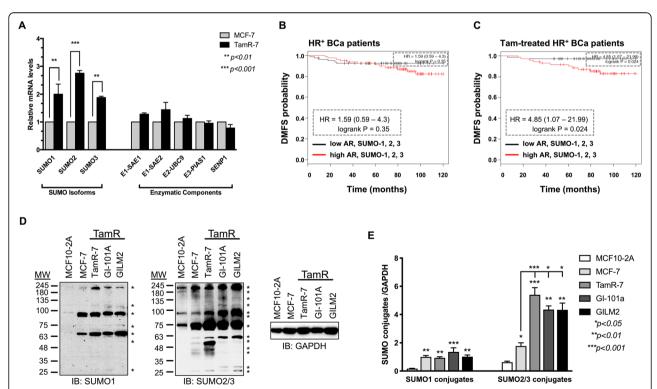


Fig. 2 Global SUMOylation increases in TamR-BCa. **a** SUMO transcripts are significantly higher in TamR-7 BCa cells. Real-time PCR was performed using TamR-7 and parental MCF-7 cells. Graph represents fold-changes ($2^{\Delta\Delta Ct}$ values) in gene expressions of 3 independent experiments and statistical significance in raw **Δ**Ct values using Student's t-test. **b-c** Tam-treated HR+ BCa patients expressing high levels of AR and SUMO isoforms show high probability for developing metastasis. Plots for: **b** HR+-BCa patients; (**c**) Tam-treated HR+ BCa patients illustrate 10-year DMFS. HR and log-rank P values are enlarged in dotted box. **d-e** Western blot analysis show enhanced global SUMO2/3 conjugates in acquired and intrinsic TamR-BCa cell lines; SUMO conjugates are marked by asterisks. **e** Blots from (D) were analyzed by densitometry using ImageJ from at least two independent experiments. Overall SUMO-1 or SUMO-2/3 conjugates were normalized to GAPDH and graphed as mean ± SEM. One-way ANOVA followed by Tukey's multiple comparison test was performed to compare between groups and statistical significance is shown on the graph

SUMO2/3-conjugated proteomes than TamS MCF-7 cells (Fig. 2d-e). Hence, induction of SUMO isoforms supports the elevated SUMOylome in TamR-BCa.

AR is a substrate for the SUMO E3 ligase Hsp27

We evaluated the relationship between AR and SUMO E3 ligases expressed in drug-resistant BCa cells; specifically, PIAS1, an established E3 [25, 28] versus Hsp27, a novel E3 for AR. Interestingly, Hsp27, but not PIAS1, transcript is elevated in TamR cells (Additional file 1: Fig. S1A and Fig. 2a) and this induction correlates with metastasis (log-rank p = 0.01 in Additional file 1: Fig. S1C and p = 0.82 in Fig. S1D). Also, induction of SUMO, analogous to native TamR conditions, enhances the interaction of AR with Hsp27, but not PIAS1 (Additional file 1: Fig. S2A). As SUMOylated AR in TamR-7 cells is chromatin bound (Additional file 1: Fig. S2B), coimmunoprecipitation studies were performed in chromatin fractions. We observe enhanced interaction of AR with Hsp27 in the chromatin fraction of SUMO-induced TamS MCF-7 cells (Fig. 3a) and natural hyperSUMO environments of TamR-BCa cells (Fig. 3b).

Next, we tested if Hsp27 can serve as a SUMO-E3-ligase for AR. As required for ligase activity, recombinant Hsp27 associates with the substrate AR, SUMO-E2 Ubc9, and SUMO2 protein (Fig. 3c). To test this further, we did in vitro SUMOylation assays of AR in presence and absence of Hsp27. Surprisingly, SUMO-2 by itself did not generate high-molecular-weight AR conjugates which were only observed when recombinant Hsp27 was added (lanes 3 versus 4, Fig. 3d). Hence, we show for the first time that Hsp27 potentiates AR SUMOylation via its SUMO E3 ligase activity.

AR SUMOylation drives its chromatin binding

As SUMOylation is known to alter the subcellular localization of conjugated proteins, we performed proximity ligation assays to examine the SUMO-AR distribution. AR-SUMO conjugates (red puncta) is readily detected and shifts from the cytoplasm to the nucleus with the overexpression of SUMO3 in TamS MCF-7 cells (Fig. 4a). A significant 4-fold increase in the total number of AR/SUMO complexes per nuclei occurs under hyperSUMO conditions (p < 0.001; Fig. 4b). In

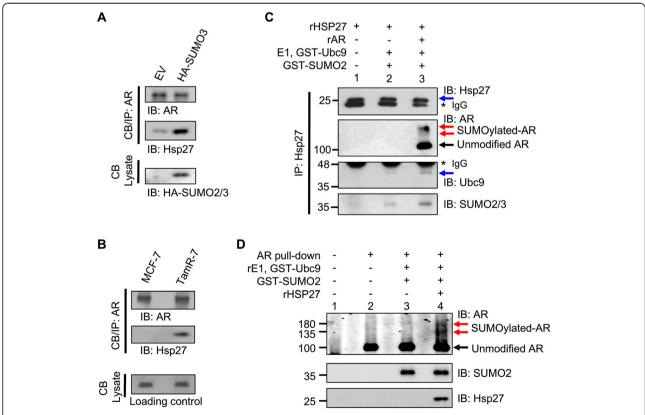


Fig. 3 AR interacts with and is a substrate for the SUMO E3 ligase Hsp27. **a** Ectopic induction of a hyperSUMO environment in MCF-7 cells with HA-SUMO3 increases the interaction between AR and Hsp27. **b** AR-Hsp27 interaction complex is enhanced endogenously in the naturally occurring hyperSUMO environment of TamR-7 cells. **a-b** Interactions with chromatin-bound (CB) AR were analyzed by western blot. **c** Immunoprecipitation of Hsp27 was performed to assess its binding with AR, Ubc9 and SUMO-2/3 in a cell-free system. **d** SUMOylation of AR is enhanced under in vitro conditions when recombinant Hsp27 is added. Details are found in (Additional file 1)

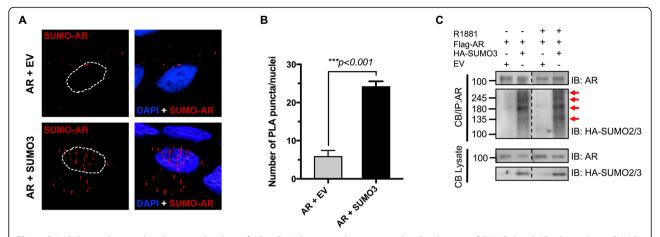


Fig. 4 SUMOylation dictates the chromatin binding of AR. **a** PLA shows predominant nuclear localization of SUMOylated AR when a hyperSUMO environment is introduced. MCF-7 cells were co-transfected with AR + EV or SUMO-3. The DuoLink kit was used according to manufacturer's instructions and total number of AR/SUMO complexes (red puncta) per nuclei were quantified using Image J. Represented images were derived from original images by deconvolution; white-dashed lines highlight the nuclei. **b** Graph represents quantification of nuclear red puncta as mean ± SEM. Student's *t*-test was used to compare groups and ****p* < 0.001 was considered statistically significant. **c** SUMO-3 enhances SUMOylation and recruitment of AR to chromatin even when cells were unstimulated with synthetic androgens (R1881). ET-R-like conditions were introduced in hormone-deprived MCF-7 cells by overexpressing AR with and without SUMO-3. After 24 h, cells were unstimulated with 10 nM R1881. Chromatin-bound AR were immunoprecipitated, resolved by SDS-PAGE and immunoblotted with specific antibodies. SUMO-2/3 conjugates of AR are identified with red arrows

vivo SUMOylation studies show that treatment with synthetic androgen R1881 enriches SUMOylated AR at the chromatin (Fig. 4c). However, and to our surprise, hyperSUMO environment in TamS cells also increase SUMOylated AR at the chromatin independent of androgen (Fig. 4c). We also observe endogenous SUMOylated AR in chromatin fractions in TamR cells (Additional file 1: Fig. S2B). Collectively, these results show that AR SUMOylation promotes chromatin enrichment independent of ligand activation.

Additionally, as inhibition of SUMOylation decreases modified AR protein levels (Fig. 1d), we investigated if and how the SUMO system stabilizes AR. First, cyclohexamide (CHX) pulse experiments were conducted to determine the rate of degradation of endogenous AR in TamR-7 versus TamS parental MCF-7 cells. Clearly, greater AR protein stability contributes to the greater population of AR in TamR-7; specifically, $T_{1/2}\!>\!24\,h$ in TamR versus $T_{1/2}\approx$ 13 h in TamS (Additional file 1: Fig. S3A). Next, to test if SUMO supports AR stability, parental TamS MCF-7 were overexpressed with SUMO and then subjected to CHX studies. SUMO overexpression significantly reduces the rate of AR degradation $(T_{1/2} \approx 24 \text{ h})$ as compared to empty vector ($T_{1/2} \approx 8.2 \text{ h}$). However, SUMO overexpression produced no significant difference in the degradation rate of upper band modified AR (Additional file 1: Fig. S4B), suggesting that SUMOmodified AR is more resilient to degradation. Consistently, analogous SUMO condition also reduces AR ubiquitylation (Additional file 1: Fig. S4C). Hence, hyperSUMO conditions favors AR stability.

AR SUMOylation drives androgen-independent genomic activity

Since SUMO enhanced the chromatin binding of AR (Fig. 4c), we postulated that AR SUMOylation likely impacts AR transcriptional activity. We generated a SUMO-mimetic AR (S-AR) to test if direct AR SUMOylation enhances AR transcriptional activity. Higher molecular weight bands of AR-SUMO conjugates are induced by S-AR but not ARwt (Fig. 5a). As observed previously in other cell lines [29], 10 nM of synthetic androgen R1881 stimulates ARwt transcription of a probasin-reporter vector in parental MCF-7 (Fig. 5b). Treatments with Enz significantly lower the R1881mediated transactivation of ARwt. In absence of exogenous ligand, the S-AR mimetic exhibits substantially greater basal transcriptional activity than ARwt (p < 0.001; Fig. 5b), suggesting that addition of SUMO to AR directs AR transcriptional function. Enzalutamide treatments on the other hand, had no effect on the increased basal transcriptional activity of S-AR, proposing that SUMOylated AR is resilient to AR antagonists. Additional studies with alternate ARE-reporter constructs were conducted to test for basal activity of S-AR. In absence of exogenous ligand, we observe significant increases in the basal S-AR transactivation of PSA-, GREand KLF5-ARE luciferase reporters (Additional file 1: Fig. S4). Consequently, we postulated that S-AR would likely modulate AR-regulated genes including previously identified epithelial-mesenchymal transition genes [30]. In absence of exogenous R1881, S-AR significantly inmesenchymal creased several gene transcripts

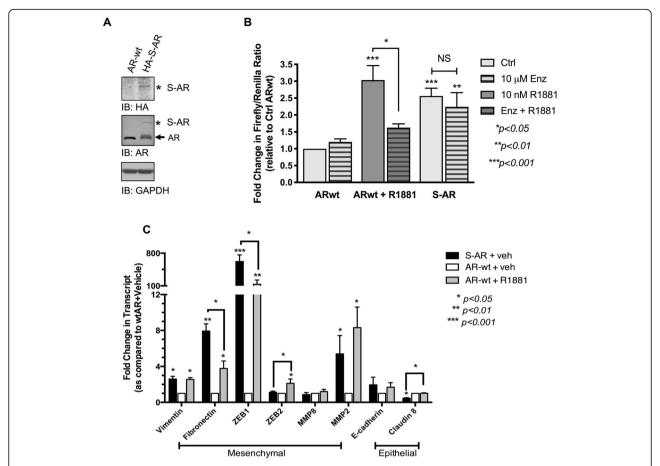


Fig. 5 SUMOylation upregulates AR-dependent transcriptional activity and mesenchymal gene expression in BCa. **a** Characterizing the SUMO-mimetic AR construct by western blot; asterisk represents AR-SUMO conjugates. **b** MCF-7 cells were hormone-deprived for 48 h then transfected with probasin and renilla luciferase together with either ARwt or SUMO-fused AR (S-AR). The next day, cells were either untreated or treated with 10 μM Enz, 10 nM R1881 or a combination of both for additional 24 h then harvested and assayed for luciferase activity. Graph B represents fold changes in firefly/renilla ratios for at least three independent experiments, each performed in triplicates and asterisks denote significant changes using One-way ANOVA followed by Tukey's multiple comparison test. **c** RNA from cells expressing ARwt or S-AR were used for real-time PCR to assess the expression of mesenchymal, epithelial markers and EMT inducers (ZEB1 and ZEB2). Graph represents fold-changes in indicated mRNA ($2^{\Delta\Delta Ct}$ values) for 3 independent experiments; asterisks indicate statistically significant differences in raw ΔCt values when compared to the ARwt + vehicle group using Student's *t*-test

specifically vimentin, fibronectin, transcription factor ZEB1, and MMP2 compared to wild-type AR (Fig. 5d). Activation of ARwt with 10 nM R1881 also enhanced similar mesenchymal gene transcripts. While comparable induction is observed for vimentin and MMP2, S-AR initiates fibronectin and ZEB1 transcription and represses claudin-8 more efficiently than R1881-activated ARwt (Fig. 5d). Collectively, these results highlight that SUMO-PTM supports a ligand-independent hyperactive AR population in TamR-BCa.

SUMO inhibitor potentiates the anti-metastatic activity of AR's antagonist Enzalutamide

We next asked if hyperactive SUMOylated AR supports the migration and metastatic phenotype of TamR-BCa cells. Confluent monolayers of TamR-7 cells were scratched and treated with the SUMO-PTM inhibitor (GA), AR antagonist (Enz), or a combination of GA and Enz (GA + Enz). Concurrent GA + Enz treatment significantly inhibited migration and scratch gap closure of TamR-7 at all time points as compared to control or either drug alone (Fig. 6a-b). We also tested whether suppressed migration of TamR-7 BCa cells by combined GA + Enz therapy is due to reduced proliferation. Percentages of both viable and dead cells of 48-h treated TamR-7 cells were not significantly affected by mono or combined drug treatments when compared to the control group (Additional file 1: Fig. S5A), indicating that GA + Enz inhibits TamR-BCa migration more than growth. Hence, reducing hyperSUMO conditions concurrent with inhibiting AR activation reduces the migration of TamR-7 BCa cells.

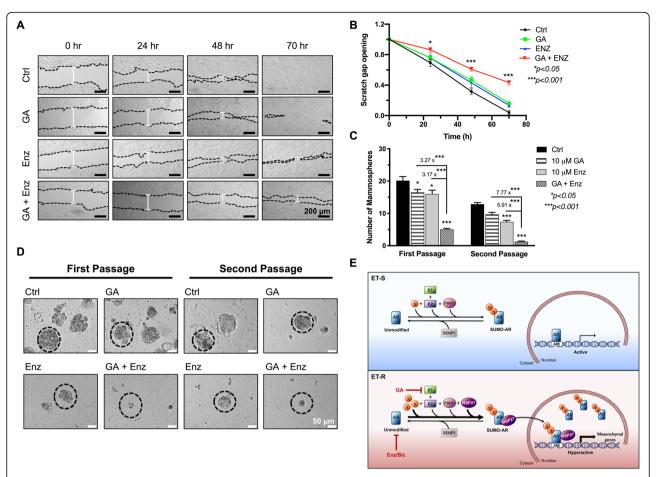


Fig. 6 Co-treatments with enzalutamide and SUMO inhibitor decrease the migration and metastatic phenotype of TamR BCa. **a-b** Images from scratch wound healing assays indicate that GA + Enz decrease the migration of TamR-7 cells. Black dotted lines define the scratch borders and white lines represent gap width. Remaining scratch gap opening in (**b**) for each treatment at each time point was calculated as a fold relative to the corresponding original gap width at 0 h. Graph is a representation of two independent experiments, each performed in duplicates. **c-d** Single-cell suspensions of adherent TamR-7 cells were generated, grown in nonadherent media with indicated treatments and mammosphere images were taken 7 days post treatments. First-generation mammospheres were passaged and dissociated into single cell suspensions and cultured the same way to test for second-generation spheroids. Graph in panel (**c**) represents total mammosphere counts as mean ± SEM from four independent experiments; representative images are shown in (Additional file 1: Fig. S5B). **d** Images illustrate mammosphere size and black-dashed circles highlight size differences between untreated and treated spheroids. Comparisons with control group or between groups, in graphs B and C, were assessed using one-way ANOVA followed by Tukey's multiple comparison test and significance is illustrated on graphs. **e** Schematic representation of AR regulation by SUMO in ET-S and ET-R BCa. Detailed description is provided in text. Encircled S represents SUMO

We and others observe that TamR-derived MCF-7 BCa cells grow efficiently in non-adherent spheroid cultures than parental MCF-7 cells consistent with their more metastatic phenotype [31, 32]. Hence, we assessed this common anchorage-independent growth and metastatic phenotype of TamR-7 cells. Concurrent GA and Enz or bicalutamide (Bic) treatments reduce the number of mammospheres formed from single-cell suspensions of TamR-7 cells (Fig. 6c and Additional file 1: Fig. S5B, S5D). In addition, the self-renewal capacity of TamR-7 cells is significantly blunted with the GA and Enz treatment as seen with fewer mammospheres forming in second passage studies (Additional file 1: Fig. S5B and 6C). Interestingly, combining GA and Enz produce a

synergistic effect on mammosphere counts as compared to control (3.99-fold for 1st generation and 10.47-fold for 2nd generation; Fig. 6c). In addition, treatments with GA + Enz significantly decrease the size of spheroids when compared to control or either drug alone mammospheres (Fig. 6d). Although, 50–100 μm diameter spheroids represent the majority of control and drug alone spheroid population, GA + Enz/Bic treatments produced a population shift with the majority being small-diameter spheroids < 50 μm in first and second passage studies (Additional file 1: Fig. S5C and S5E). Hence, the combination therapy reduces anchorage-independent growth and self-renewal properties of TamR-BCa; both critical for metastasis. Collectively, targeting both

unmodified (active) and SUMO-modified (hyperactive) forms of AR could serve as a better approach for treating tamoxifen-resistant breast tumors.

Discussion

In this study, we report enrichment of SUMO-modified AR with ligand-independent activity in ET-R BCa as supported with the following results. First, while the unmodified AR is present in hormone-sensitive MCF-7 cells, the higher molecular weight SUMO-AR accumulates in cells with acquired (TamR-7) and intrinsic (GI101A and GILM2) resistance to tamoxifen endocrine therapy, and long-term estrogen deprived cells (Fig. 1c). Second, upregulation of SUMO in hormone-sensitive MCF-7 promotes chromatin accumulation of AR analogous to conventional androgen-activated AR (Fig. 4c). Third, also like canonical ligand-activated AR, SUMObound AR initiates transcription of ARE-expressing luciferase reporter (Fig. 5b and Fig. S4). Finally, AR antagonist Enz reduces ET-R BCa cell migration and mammosphere formation (Fig. 6) demonstrating that basal AR activity supports aberrant metastatic properties of the drug-resistant cell line.

Clearly tamoxifen therapy impacts how AR directs disease progression; unlike the general HR+ BCa patient population, high AR in patients receiving tamoxifen favors a worse prognosis (Fig. 1a-b). In the tamoxifentreated patient population, the hazard ratio is further potentiated with concurrent induction of the SUMO isoforms (Fig. 2c). Consistently the global SUMOylome is also upregulated in the TamR cell lines as compared to non-cancerous or hormone-sensitive BCa cells (Fig. 2de). A previous study shows that Myc-dependent BCa require specifically the E1-SAE2 dimer component for tumor growth [33]. While we did not observe an SAE2 elevation in ET-R HR+ BCa cells, clearly a need to potentiate the SUMOylome is a shared event for cancer progression. Consistently, others propose inhibiting global SUMO-PTM with GA or less selective Anacardic acid as promising therapy for the another BCa subtype TNBC [34, 35]. In our studies, HR+ BCa cells resistant to ET are responsive to the inhibitory actions of GA (Fig. 6). Like basal ER negative, ET-R ER α + BCa cells are more mesenchymal than stagnant MCF7, suggesting that accumulation of SUMOylated proteins could drive the metastatic phenotype in multiple BCa subtypes. In the present study, we demonstrate that SUMO-AR supports the transcriptome profile required to initiate this phenotype.

For AR, induction of SUMO is not sufficient for AR SUMOylation. In our cell-free system, we demonstrate that the SUMO-E3 ligase Hsp27 is required for SUMO-

PTM of AR (Fig. 3d). In vivo, high SUMO levels, analogous to endogenous levels in TamR-BCa, support greater binding between AR and Hsp27 than another established AR SUMO E3 ligase PIAS1 [25, 28] (Fig. 3a, b, and S2A). Hsp27 exhibits SUMO-E3 ligase enzymatic activity for limited protein substrates specifically HSF1 [36] and CFTR mutant F508del [37]. Consistent with SUMO E3 ligase activity for these other substrates, Hsp27 concurrently binds SUMO, SUMO-E2 Ubc9, and substrate-AR (Fig. 3c) to drive AR SUMO-conjugation kinetics (Fig. 3d). Unlike PIAS1 [25, 28], Hsp27 supports AR SUMOylation in the absence AR-ligand (Fig. 3). These findings present an additional regulatory control mechanism based on PTM to the canonical substrate-chaperone relationship between AR and Hsp27 established previously [27, 38].

AR agonist DHT stimulates transcription of mesenchymal genes and metastasis of MCF7 cells in xenograft studies [30]. In MCF7 cells, SUMO-bound AR functions analogous to ligand-activated AR. Specifically, SUMO-AR initiates transcription of multiple mesenchymaldriving genes equivalent to or greater than R1881activated AR (Fig. 5d). Yet surprisingly, transcriptional activity of canonical ligand-activated but not SUMObound AR responded to Enz treatments. Present results show that lowering global SUMOylation potentiates the effectiveness of AR antagonists and may have greater impact in inhibiting TamR-BCa migration more than growth (Fig. 6b; Additional file 1: Fig. S5A). Mesenchymal TamR BCa enriched in nonadherent spheroid conditions are more responsive to Enz/GA combinatorial therapy (Fig. 6c). While raising evidence advocates the use of AR antagonists clinically for advanced HR + -BCa [39, 40], Enz in combination with exemestane failed to improve the PFS of HR+ postmenopausal women with prior endocrine therapy but was successful for those who did not receive it [9]. It is intriguing to speculate that these clinical results are due to differences in a mixed AR population (unmodified and SUMOylated AR). If the majority of AR is being highly modified, then Enz will antagonize the ligand activated unmodified AR but not the hyperactive modified AR which functionally does not depend entirely on the ligand.

Conclusion

Collectively and as illustrated in Fig. 6e, SUMOylation of AR in ET-S cells is dynamic and reversible. However, a sustained hyperSUMO environment, observed in ET-R but not in ET-S cells, favors AR-SUMOylation and enhances its interaction with Hsp27. Concomitant rise in the levels of nuclear SUMOylated AR available for chromatin binding results in AR-mediated transcriptional

activation and induction of genes promoting metastasis. Hence, a better approach in targeting various AR populations in ET-R BCa, as the unmodified (active) and SUMO-modified (hyperactive) forms of AR, could be achieved by combining AR-targeted therapies (as Enz or Bic) with SUMOylation inhibitors (as GA).

Our data suggest that dual targeting of AR and SUMO might have clinical and therapeutic relevance in the ET-R HR+ BCa subset of breast cancer. Therefore, increased global SUMOylation and in particular SUMO-modified AR may be a potential predictive biomarker useful to stratify HR+ BCa patients that are most likely to benefit from combining AR antagonists with SUMO inhibitors.

Supplementary information

Supplementary information accompanies this paper at https://doi.org/10. 1186/s12964-020-00649-z.

Additional file 1. Supplemental materials and methods: in vitro SUMOylation; RT-PCR; Luciferase reporter assays; PLA, mammosphere studies; scratch assays; transcriptomic data analysis. **Table S1.** List of primer sequences used for the detection of transcripts. Supplemental figures and figure legends: **Figure S1.** Elevated levels of SUMO isoforms and HSP27 correlate with high probability of metastasis in ET-treated HR+ BCa patients. **Figure S2.** HyperSUMO conditions promotes AR SUMOylation and enhances its interaction with Hsp27. **Figure S3.** SUMO stabilizes AR and reduces its proteasomal degradation. **Figure S4.** SUMO stimulates basal AR transcriptional activity regardless of the AR-luciferase reporter construct. **Figure S5.** Concurrent targeting of SUMO-modified and unmodified AR decreases TamR-7 BCa growth in 3D cultures.

Abbreviations

Al: aromatase inhibitor; AR: androgen receptor; ARE: androgen response elements; BCa: breast cancer; DMFS: distant-metastasis free survival; Enz: enzalutamide; HR+: hormone receptor-positive; GA: ginkgolic acid; Hsp27: heat shock protein 27; HR+: hormone receptor positive; ET-R: endocrine therapy-resistant; ET-S: endocrine therapy-sensitive; PFS: progression-free survival; PIAS1: Protein inhibitor of activated STAT; PTM: posttranslational modifications; PLA: Proximity Ligation Assay; SENP1: SUMO specific peptidase 1; SUMO: Small Ubiquitin Like Modifier; S-AR: SUMO-fused-AR; TamR: tamoxifen resistant; TamS: tamoxifen sensitive; ARwt: wild-type AR

Acknowledgements

We thank Dr. Feng (University of Houston) for providing MCF-7-EDR cells and Drs. Weigel (Baylor College of Medicine) and Frigo (MD Anderson Cancer Center) for sharing luciferase reporter constructs and providing guidance with interpreting results. We thank Saathvika Ravi and Lynna Nguyen for their technical assistance.

Authors' contributions

TBK and SB designed the experiments. SB performed the majority of the experiments, analyzed data and prepared Figs. HT purified GST recombinant proteins for in vitro SUMOylation studies, MQ performed RT-PCR for epithelial/mesenchymal transcripts, AK performed scratch assays, JR generated the SUMO-fused-AR construct, DD and SK helped with cell cultures, and JR did in vivo SUMOylation of AR. SB and TBK wrote the manuscript and TBK supervised the study. All authors read and approved the final manuscript.

Funding

Not applicable.

Availability of data and materials

Not applicable.

Ethics approval and consent to participate

Not applicable.

Consent for publication

All authors agreed to publish this study.

Competing interests

There is no potential conflict of interest to disclose.

Author details

¹Center for Nuclear Receptors & Cell Signaling, Department of Biology & Biochemistry, University of Houston, 3517 Cullen Blvd, SERC Bldg, Rm 3010, Houston, TX 77204-5056, USA. ²Department of Pharmacy Practice and Translational Research, College of Pharmacy, University of Houston, Houston, TX 77204, USA. ³Department of Biology, Stanford University, Stanford, CA 94305, USA. ⁴Center for Precision Medicine, Department of Medicine, University of Missouri, Columbia, MO 65212, USA.

Received: 8 April 2020 Accepted: 24 August 2020 Published online: 18 September 2020

References

- Christopoulos PF, Vlachogiannis NI, Vogkou CT, Koutsilieris M. The role of the androgen receptor signaling in breast malignancies. Anticancer Res. 2017;37(12):6533–40.
- Gao W, Bohl CE, Dalton JT. Chemistry and structural biology of androgen receptor. Chem Rev. 2005;105(9):3352–70.
- Vera-Badillo FE, Templeton AJ, de Gouveia P, Diaz-Padilla I, Bedard PL, Al-Mubarak M, et al. Androgen Receptor Expression and Outcomes in Early Breast Cancer: A Systematic Review and Meta-Analysis. J Natl Cancer Inst. 2014;106(1):djt319-djt.
- Cochrane DR, Bernales S, Jacobsen BM, Cittelly DM, Howe EN, D'Amato NC, et al. Role of the androgen receptor in breast cancer and preclinical analysis of enzalutamide. Breast Cancer Res. 2014;16(1):R7.
- De Amicis F, Thirugnansampanthan J, Cui Y, Selever J, Beyer A, Parra I, et al. Androgen receptor overexpression induces tamoxifen resistance in human breast cancer cells. Breast Cancer Res Treat. 2010;121(1):1–11.
- D'Amato NC, Gordon MA, Babbs B, Spoelstra NS, Carson Butterfield KT, Torkko KC, et al. Cooperative dynamics of AR and ER activity in breast Cancer. Mol Cancer Res. 2016;14(11):1054–67.
- Chia K, Milioli H, Portman N, Laven-Law G, Coulson R, Yong A, et al. Noncanonical AR activity facilitates endocrine resistance in breast cancer. Endocr Relat Cancer. 2019;26(2):251–64.
- 8. Ponnusamy S, Asemota S, Schwartzberg LS, Guestini F, McNamara KM, Pierobon M, et al. Androgen Receptor Is a Non-canonical Inhibitor of Wild-Type and Mutant Estrogen Receptors in Hormone Receptor-Positive Breast Cancers. iScience. 2019;21:341–58.
- Krop I, Abramson V, Colleoni M, Traina T, Holmes F, Estevez L, et al. Abstract GS4-07: results from a randomized placebo-controlled phase 2 trial evaluating exemestane ± enzalutamide in patients with hormone receptor– positive breast cancer. Cancer Res. 2018;78(4 Supplement):GS4-07.
- Gioeli D, Paschal BM. Post-translational modification of the androgen receptor. Mol Cell Endocrinol. 2012;352(1–2):70–8.
- van der Steen T, Tindall DJ, Huang H. Posttranslational modification of the androgen receptor in prostate cancer. Int J Mol Sci. 2013;14(7):14833–59.
- Gareau JR, Lima CD. The SUMO pathway: emerging mechanisms that shape specificity, conjugation and recognition. Nat Rev Mol Cell Biol. 2010;11(12):861–71.
- Poukka H, Karvonen U, Jänne OA, Palvimo JJ. Covalent modification of the androgen receptor by small ubiquitin-like modifier 1 (SUMO-1). Proc Natl Acad Sci. 2000;97(26):14145–50.
- Rytinki M, Kaikkonen S, Sutinen P, Paakinaho V, Rahkama V, Palvimo JJ. Dynamic SUMOylation is linked to the activity cycles of androgen receptor in the cell nucleus. Mol Cell Biol. 2012;32(20):4195–205.
- Bawa-Khalfe T, Cheng J, Wang Z, Yeh ET. Induction of the SUMO-specific protease 1 transcription by the androgen receptor in prostate cancer cells. J Biol Chem. 2007;282(52):37341–9.
- 16. Cheng J, Bawa T, Lee P, Gong L, Yeh ET. Role of desumoylation in the development of prostate cancer. Neoplasia. 2006;8(8):667–76.
- Cheng J, Wang D, Wang Z, Yeh ET. SENP1 enhances androgen receptordependent transcription through desumoylation of histone deacetylase 1. Mol Cell Biol. 2004;24(13):6021–8.

- Sutinen P, Malinen M, Heikkinen S, Palvimo JJ. SUMOylation modulates the transcriptional activity of androgen receptor in a target gene and pathway selective manner. Nucleic Acids Res. 2014;42(13):8310–9.
- Bawa-Khalfe T. Isolation of in vivo SUMOylated chromatin-bound proteins.
 In: Rodriguez MS, editor. SUMO: methods and protocols. New York: Springer New York; 2016. p. 205–16.
- Gyorffy B, Lanczky A, Eklund AC, Denkert C, Budczies J, Li Q, et al. An online survival analysis tool to rapidly assess the effect of 22,277 genes on breast cancer prognosis using microarray data of 1,809 patients. Breast Cancer Res Treat. 2010:123(3):725–31.
- Morrissey JJ, Raney S. A metastatic breast tumor cell line, Gl-101a, is estrogen receptor positive and responsive to estrogen but resistant to Tamoxifen. Cell Biol Int. 1998;22(6):413–9.
- 22. Chelouche Lev D, Kiriakova G, Price JE. Selection of more aggressive variants of the Gl101A human breast cancer cell line: a model for analyzing the metastatic phenotype of breast cancer. Clin Experimental Metastasis. 2003; 20(6):515–23
- Fukuda I, Ito A, Hirai G, Nishimura S, Kawasaki H, Saitoh H, et al. Ginkgolic Acid Inhibits Protein SUMOylation by Blocking Formation of the E1-SUMO Intermediate. Chem Biol. 2009;16(2):133–40.
- Kaikkonen S, Jaaskelainen T, Karvonen U, Rytinki MM, Makkonen H, Gioeli D, et al. SUMO-specific protease 1 (SENP1) reverses the hormone-augmented SUMOylation of androgen receptor and modulates gene responses in prostate cancer cells. Mol Endocrinol. 2009;23(3):292–307.
- Nishida T, Yasuda H. PIAS1 and PIASxalpha function as SUMO-E3 ligases toward androgen receptor and repress androgen receptor-dependent transcription. J Biol Chem. 2002;277(44):41311–7.
- Qin Y, Xu J, Aysola K, Begum N, Reddy V, Chai Y, et al. Ubc9 mediates nuclear localization and growth suppression of BRCA1 and BRCA1a proteins. J Cell Physiol. 2011;226(12):3355–67.
- Zoubeidi A, Zardan A Fau Beraldi E, Beraldi E Fau Fazli L, Fazli L Fau Sowery R, Sowery R Fau Rennie P, Rennie P Fau Nelson C, et al.
 Cooperative interactions between androgen receptor (AR) and heat-shock protein 27 facilitate AR transcriptional activity. Can Res. 2007;67(21):10455–65
- Toropainen S, Malinen M, Kaikkonen S, Rytinki M, Jääskeläinen T, Sahu B, et al. SUMO ligase PIAS1 functions as a target gene selective androgen receptor coregulator on prostate cancer cell chromatin. Nucleic Acids Res. 2015;43(2):848–61.
- Cronauer MV, Schulz WA, Burchardt T, Ackermann R, Burchardt M. Inhibition of p53 function diminishes androgen receptor-mediated signaling in prostate cancer cell lines. Oncogene. 2004;23:3541.
- Feng J, Li L, Zhang N, Liu J, Zhang L, Gao H, et al. Androgen and AR contribute to breast cancer development and metastasis: an insight of mechanisms. Oncogene. 2017;36(20):2775–90.
- Liu H, Zhang HW, Sun XF, Guo XH, He YN, Cui SD, et al. Tamoxifen-resistant breast cancer cells possess cancer stem-like cell properties. Chin Med J. 2013;126(16):3030–4.
- Raffo D, Berardi DE, Pontiggia O, Todaro L, de Kier Joffe EB, Simian M. Tamoxifen selects for breast cancer cells with mammosphere forming capacity and increased growth rate. Breast Cancer Res Treat. 2013;142(3):537–48.
- Kessler JD, Kahle KT, Sun T, Meerbrey KL, Schlabach MR, Schmitt EM, et al. A SUMOylation-dependent transcriptional subprogram is required for Mycdriven tumorigenesis. Science. 2012;335(6066):348–53.
- Bogachek MV, Park JM, De Andrade JP, Lorenzen AW, Kulak MV, White JR, et al. Inhibiting the SUMO pathway represses the Cancer stem cell population in breast and colorectal carcinomas. Stem Cell Reports. 2016;7(6): 1140–51.
- Bogachek MV, Chen Y, Kulak MV, Woodfield GW, Cyr AR, Park JM, et al. Sumoylation pathway is required to maintain the basal breast cancer subtype. Cancer Cell. 2014;25(6):748–61.
- Brunet Simioni M, De Thonel A, Hammann A, Joly AL, Bossis G, Fourmaux E, et al. Heat shock protein 27 is involved in SUMO-2/3 modification of heat shock factor 1 and thereby modulates the transcription factor activity. Oncogene. 2009;28(37):3332–44.
- Ahner A, Gong X, Frizzell RA. Divergent signaling via SUMO modification: potential for CFTR modulation. Am J Physiol Cell Physiol. 2016;310(3):C175–80.
- Liu X, Feng C, Liu J, Cao L, Xiang G, Liu F, et al. Androgen receptor and heat shock protein 27 co-regulate the malignant potential of molecular apocrine breast cancer. J Exp Clin Cancer Res. 2018;37(1):90.

- Takagi K, Miki Y, Ishida T, Sasano H, Suzuki T. The interplay of endocrine therapy, steroid pathways and therapeutic resistance: importance of androgen in breast carcinoma. Mol Cell Endocrinol. 2017;466:31–37.
- Schwartzberg LS, Yardley DA, Elias AD, Patel M, LoRusso P, Burris HA, et al. A
 phase I/lb study of Enzalutamide alone and in combination with endocrine
 therapies in women with advanced breast Cancer. Clin Cancer Res. 2017;
 23(15):4046–54.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

