CORRECTION Open Access

Correction to: PTH Derivative promotes wound healing via synergistic multicellular stimulating and exosomal activities



Yi-Fan Shen^{1†}, Jing-Huan Huang^{1†}, Kai-Yang Wang¹, Jin Zheng³, Lin Cai², Hong Gao^{1*}, Xiao-Lin Li^{1*} and Jing-Feng Li^{2*}

Correction to: Cell Commun Signal (2020) 18:40 https://doi.org/10.1186/s12964-020-00541-w

Following publication of the original article [1], two mistakes were noticed in Fig. 4 and Fig. 6. The pictures describing the effects of 0.1 nM PTHrP-2 group on migration of HUVEC in Fig. 4 and Control and HFF-1-Exos groups on migration of HFF-1 cells in Fig. 6 are incorrect. The correct figures are supplied below in this correction article. The figure legends were not changed.

The authors sincerely apologize for having this unintentional error in the article, and apologize for any inconvenience caused.

Author details

¹Department of Orthopaedic Surgery, Shanghai Jiao Tong University Affiliated Sixth People's Hospital, Shanghai, People's Republic of China. ²Department of Orthopedics, Zhongnan Hospital of Wuhan University, Wuhan, People's Republic of China. ³Department of Neurology, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, People's Republic of China.

Published online: 19 May 2020

Reference

 Shen Y, Huang J, Wang K, et al. PTH derivative promotes wound healing via synergistic multicellular stimulating and exosomal activities. Cell Commun Signal. 2020;18:40 https://doi.org/10.1186/s12964-020-00541-w.

The original article can be found online at https://doi.org/10.1186/s12964-020-00541-w.

Full list of author information is available at the end of the article



© The Author(s). 2020 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

^{*} Correspondence: honggao630@163.com; lixiaolin@sjtu.edu.cn; jingfengli@whu.edu.cn

[†]Yi-Fan Shen and Jing-Huan Huang contributed equally to this work. ¹Department of Orthopaedic Surgery, Shanghai Jiao Tong University Affiliated Sixth People's Hospital, Shanghai, People's Republic of China ²Department of Orthopedics, Zhongnan Hospital of Wuhan University, Wuhan, People's Republic of China

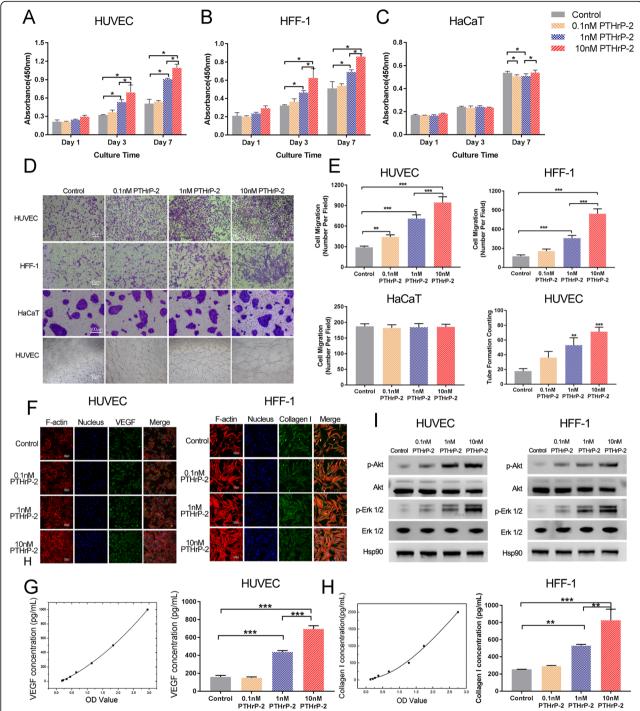


Fig. 4 Proliferation of HUVECs (**a**), HFF-1 cells (**b**), HaCaTs (**c**) incubated for 0, 1, 3, or 7 days in conditioned medium with different drug concentrations from days 0 and 6. **d** Effects of PTHrP-2 on migration of HUVECs, HFF-1 cells and HaCaTs and the tube formation assay of HUVECs. **e** Quantitation of HUVECs, HFF-1 cells and HaCaTs migration (violet stained cells) using a Transwell chamber. The quantitative evaluation of the number of nodes formed in the culture plate with different drug concentrations after 8 h. **f** Immunofluorescence images of HUVECs and HFF-1 incubated in each group on day 3. Cytoskeleton and cell nuclei are stained red and blue, VEGF and Collagen I are stained green in the picture taken by the laser scanning confocal microscopy. **g** VEGF and Collagen I secretion by HUVEC and HFF-1 incubated for 3 days in media with different drug concentrations. **h** Akt and Erk1/2 phosphorylation level in HUVEC and HFF-1 treated with different drug concentrations

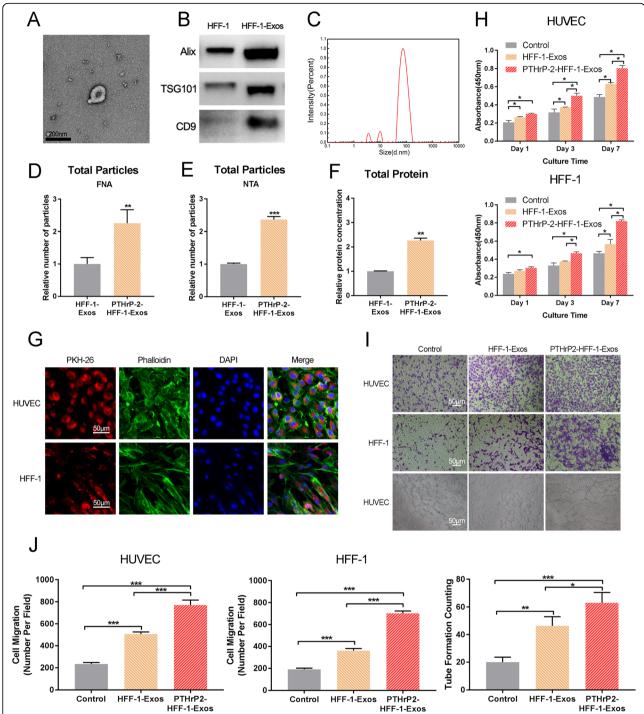


Fig. 6 a TEM images of HFF-1-Exos. **b** Exosome surface markers detected by Western blotting (Alix, Tsg101, CD9). The experiment was repeated three times in order to confirm the stability of the phenomena. **c** Size distribution of exosomes. Particle concentration, particle size and video frame of exosomes were analyzed by FNA (**d**) and NTA (**e**). Total protein levels (**f**) in HFF-1-Exos and PTHrP-2-HFF-1-Exos. **g** The uptake of exosomes by HUVECs and HFF-1 cells. Cytoskeleton, exosomes and cell nuclei are stained green, red and blue in the picture taken by the laser scanning confocal microscopy. **h** Proliferation of HUVECs and HFF-1 cells incubated for 0, 1, 3, or 7 days in conditioned medium with HFF-1-Exos and PTHrP-2-HFF-1-Exos from days 0 and 6. **l** Effects of HFF-1-Exos on migration of HUVECs and HFF-1 cells and the tube formation assay of HUVECs. **j** Quantitation of HUVECs and HFF-1 cells migration (violet stained cells) using a Transwell chamber. The quantitative evaluation of the number of nodes formed in the culture plate with different conditions of culture after 8 h