REVIEW

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Sex differences in colorectal cancer: with a focus on sex hormone–gut microbiome axis

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Abstract

Sexual dimorphism has been observed in the incidence and prognosis of colorectal cancer (CRC), with men generally exhibiting a slightly higher incidence than women. Research suggests that this difference may be attributed to variations in sex steroid hormone levels and the gut microbiome. The gut microbiome in CRC shows variations in composition and function between the sexes, leading to the concept of 'microgenderome' and 'sex hormone–gut microbiome axis.' Conventional research indicates that estrogens, by promoting a more favorable gut microbiota, may reduce the risk of CRC. Conversely, androgens may have a direct pro-tumorigenic effect by increasing the proportion of opportunistic pathogens. The gut microbiota may also influence sex hormone levels by expressing specific enzymes or directly affecting gonadal function. However, this area remains controversial. This review aims to explore the differences in sex hormone in CRC incidence, the phenomenon of sexual dimorphism within the gut microbiome, and the intricate interplay of the sex hormone–gut microbiome axis in CRC. The objective is to gain a better understanding of these interactions and their potential clinical implications, as well as to introduce innovative approaches to CRC treatment.

Keywords Colorectal cancer, Sex differences, Sex hormones, Gut microbiome, Sexual dimorphism

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Introduction

Colorectal cancer (CRC) represents a major public health challenge worldwide. It ranks as the third most frequently diagnosed cancer and stands as the second leading cause of cancer-related deaths in the United States. Projections for the year 2023 estimate approximately 153,020 new cases and 52,550 deaths from CRC [1]. The etiology of CRC is multifactorial, involving genetic, environmental, and lifestyle factors [2]. However, sex hormones and sex contribute to differences in CRC incidence and outcomes, underscoring the potential role of sex-specific risk factors. Recent cancer statistics have revealed that between 2015 and 2019, the average annual CRC incidence rate was 33% higher and the overall mortality rate was 43% higher in men than in women. Additionally, women exhibit a slightly higher five-year relative survival rate than men [1, 3]. These sex-specific trends can be attributed to various factors. Historically, women, particularly pregnant women, were often excluded from pivotal clinical research to protect the developing fetus from potential harm. This exclusion led to an underestimation of the influence of sex on tumor biology and clinical outcomes [4]. Moreover, dietary habits have played a role in this divergence. Men tend to consume larger quantities of meat and alcohol than women, further elevating the risk of CRC in men [5].

Recently, there has been a growing interest in the potential role of sex hormones and the gut microbiome in intestinal tumors. Previous studies have revealed an association between sex steroids and colon physiology and pathophysiology. Intestinal epithelial cells possess the ability to metabolize sex steroids, particularly estrogens, which could influence the development of CRC [6, 7]. Estrogens appear to confer protection against CRC progression, whereas androgens have been linked to an increased risk of CRC. However, controversy in this field persists, and the precise mechanisms remain unclear [4, 8]. The emerging concept of the 'microgenderome' underscores the interactions between sex hormones and the gut microbiota [9]. Recent investigations have indicated discernible compositional and functional differences in the gut microbiome between men and women. Generally, women exhibit higher diversity and richness of gut microbiota than men [10], and this divergence may underlie the observed sex-based differences in CRC risk and outcomes. The gut microbiome, a complex ecosystem composed of trillions of microorganisms, exerts crucial influences on host physiology, metabolism, and immune function [11]. Gut microbiota has been postulated to influence CRC development through several mechanisms, including the fermentation of undigested dietary fibers into short-chain fatty acids (SCFAs), modulation of the host immune system, and production of carcinogenic compounds [12, 13]. New evidence suggests the existence of a sex hormone–gut microbiome axis, where sex hormones regulate the composition and function of the gut microbiome and its metabolites, while the gut microbiome significantly influences sex hormone levels [10, 14].

Despite the growing evidence linking sex hormones and the gut microbiota to the development of CRC, there is a lack of research on their interactions and potential contributions to sex-based differences in CRC risk. Therefore, this review aims to summarize the current evidence on sex hormone differences in CRC incidence and the phenomenon of sexual dimorphism within the gut microbiome. Additionally, it explores the intricate interplay of the sex hormone–gut microbiome axis in CRC. The objective of this review is to better understand the interactions among sex hormones, the gut microbiome, and CRC, as well as to explore potential clinical implications and introduce innovative approaches to CRC treatment.

Sex hormone differences in CRC Female sex hormones and CRC Estradiol and estrone

Estrogens, primarily estradiol (E2) and estrone (E1), have been proposed as protective factors against the development of CRC. However, the relationship between estrogen and CRC remains controversial [15] (Table 1).

Clinical research Clinical studies have validated that postmenopausal CRC patients have significantly lower levels of free estradiol, total estradiol, and estrone levels, as well as more advanced tumor stage compared to premenopausal CRC patients. This suggests an inverse association between estradiol and estrone levels and CRC risk [16-19]. An investigation into the expression and single nucleotide polymorphisms of the TLR4/NF-KB pathway in relation to colon cancer has found that the TLR4 rs2770150 variant is associated with colon cancer in women over 50 years old. This association is strongly linked to the decline in female sex hormone levels in postmenopausal women [17]. Previous studies have indicated a connection between a family history of diabetes and an increased risk of CRC, and that men are more susceptible to CRC compared to women. Moreover, male patients with a family history of diabetes exhibit more significant decreases in estradiol and sex hormonebinding globulin (SHBG) levels. These associations are particularly prominent in men under 60 years old [20]. Studies have also shown that serum estradiol levels decrease by 80% after bilateral oophorectomy in premenopausal women. Women who have previously undergone oophorectomy or hysterectomy have a 24-30% higher risk of CRC compared to the general population [21, 22]. Colorectal anastomotic leakage is a common and serious complication after CRC surgery, with a higher incidence in men. Besides differences in pelvic anatomy, sex hormone differences between men and women have been suggested to play a role. Recent studies have found a lower incidence of anastomotic leakage in postmenopausal women with CRC who received preoperative estrogen replacement therapy, which may explain some of the differences in leakage rates between men and women [23]. These findings suggest a potential protective role of estrogen against CRC in women.

Animal and cellular models In vitro experiments have demonstrated that 17β-estradiol (E2) can inhibit the migration and proliferation of DLD1 cells independently of miR-34a-mediated actions [24]. E2 has also been found to reduce inflammation in the female human colonic epithelial cell line CCD841CoN. This is achieved by decreasing the expression of inflammatory factors, such as NF-KB and COX-2, while increasing the expression of antioxidant enzymes [25]. Similarly, In the azoxymethane/dextran sodium sulfate (AOM/DSS) model, estradiol supplementation was observed to reduce the severity of colitis. Additionally, at weeks 10 and 16, male (estradiol-supplemented) and female mice had significantly lower numbers of colonic polyps and tumors compared to male mice without estradiol supplementation [26]. Furthermore, ovariectomy increases the incidence of CRC in female mouse model, but estrogen supplementation reduces the number of tumors and migration rates [27, 28]. At the molecular level, estradiol significantly increases the expression of Nrf2, which contributes to the anti-inflammatory effects of estrogen by directly regulating estrogen receptor-beta (ERB) expression. Additionally, it upregulates Nrf2-associated antioxidant enzymes, ER β , and NLRP3 inflammasome and downregulates the expression of ER α and NF- κ B in the mouse colon. These actions ameliorate impaired associations with E-cadherin and β-catenin, ultimately inhibiting colorectal carcinogenesis and metastasis [26-29] (Fig. 1a). In the MC38 colon tumor model mice, males displayed faster colon tumor growth and a higher proportion of PD-L1 expression, M2 phenotype tumor-associated macrophages (TAMs), and cancer-associated fibroblasts than females. Furthermore, compared with anti-PD-L1 antibody alone, the combination of E2 with anti-PD-L1 antibody significantly inhibited MC38 tumor growth and decreased PD-L1 expression and M2/M1 TAM ratio. This

Table 1 Sex hormone-based differences in CRC

| Sex hormones | Model organism | Findings | References |
|--|--|---|------------------|
| Estradiol/17β-estradiol (E2) Estrone | Human CRC tissue or blood sample (from postmenopausal | Endogenous estradiol and estrone levels are inversely associated with CRC risk and complication. | [16–23] |
| | women) | Pre-diagnostic estrogen and other sex steroid levels are positively associated with mortality risk in female CRC survivors. | [34] |
| | | Estradiol levels are not associated with CRC risk in postmenopausal women. | [36–38] |
| | | Pathobiological role of estrogen in postmenopausal CRC varies depending on patient age and tumor characteristics. | [42] |
| | AOM/DSS, OVX_AOM/DSS, and OVX_Min/+ mice | Estradiol prevents colorectal carcinogenesis and metastasis by inhib- iting inflammatory pathways, regulating Nrf2-related signaling, and ameliorating impaired associations with E-cadherin and β -catenin. | [26–28] |
| | MC38 and OVX_MC38 tumor model mice | E2 inhibits MC38 tumor growth by regulating tumor-associated cell populations and reducing PD-L1 expression. Obesity, macrophage- associated inflammation, and TAMs are potential mechanisms for inducing CRC in females lacking estrogen. | [30, 31] |
| | | Estrogen is implicated in hepatic immunosuppression within the tumor microenvironment and promotes metastatic expansion. | [41] |
| | AOM-male mice DLD1, HT-29, SW480, SW620 cells and CSCs | E2 inhibits the migration and proliferation of DLD1 cells independently of miR-34a-mediated actions. | [24] |
| | | Combined use of E2 and progesterone treatment promotes cell cycle arrest and apoptosis by stimulating the expression of ER β and PGR and inhibiting ER α -regulated oncogenic pathways. | [32, 33] |
| | | Combined use of E2 and 5-fluorouracil treatment exhibits superior anticancer effects than monotherapy on female and male primary CRC cells. E2 monotherapy exhibits the most substantial effects on male metastatic cells. | [33] |
| | | E2 induces the expression of estrogen receptors on CSCs, promoting their migration and metastasis. | [40] |
| ERβ | Human CRC, sporadic polyps, and FAP tissue | $\text{ER}\beta$ expression is reduced in colorectal precancerous stages and plays a key role in inhibiting the development of CRC. | [44, 45, 47–49] |
| | AOM/DSS ERβ _KO mice SW480 and HT-29 cells | $ER\beta$ knockdown significantly induces $TNF\alpha$ expression and affects NF- κB inflammatory signals. | |
| Progesterone PGR | Human CRC tissue or blood sample (from postmenopausal | Progesterone is generally not associated with CRC risk in postmeno- pausal women. | [16, 37, 38, 50] |
| | women) | Progesterone and PGR expression levels positively correlate with the prognosis of CRC. | [48, 51, 52] |
| | Xenograft tumor model CRC cell lines | Progesterone activates the GADD45α/JNK pathway, arrests the cell cycle, and induces apoptosis, thereby inhibiting CRC progression. | |
| Testosterone SHBG Androstenedione DHEA | CRC/adenoma patients, tissue or blood samples | Higher levels of circulating testosterone and SHBG are associated with lower CRC risk, whereas free testosterone levels are positively associated with CRC risk. | [19, 37, 54–57] |
| Androstenedione DHEA | | Patients with prostate cancer who undergo androgen deprivation therapy have an increased risk of CRC | [58] |
| | | Free testosterone levels are negatively associated with CRC incidence and mortality in both men and women. | [34, 57, 60, 61] |
| | | Circulating concentrations of testosterone, SHBG, androstenedione, and DHEA are not associated with the risk of early precursor lesions in the colon or colon cancer | [38, 62, 63] |
| | AOM/DSS and ORX_AOM/DSS mice | Testosterone enhances AOM/DSS-induced CRC development. | [8, 32] |
| | Pirc/+ rat, Min/+ mice, and AOM mice | Sex differences in colon adenoma development may result from an indirect tumor-promoting effect of testosterone rather than a protective effect of estrogen. | [59] |

AOM/DSS Azoxymethane/dextran sodium sulfate, CRC Colorectal cancer, CSCs Cancer stem cells, DHEA Dehydroepiandrosterone, E2 Estradiol/17ß-estradiol, ERß Estrogen receptor-beta, FAP Familial adenomatous polyposis, OVX Ovariectomy, ORX Orchiectomy, PGR Progesterone receptor, SHBG Sex hormone-binding globulin, TIME Tumor immune microenvironment



Fig. 1 Sex hormone differences in CRC risk and outcome. a Molecular mechanisms underlying the protective effect of estradiol against CRC. b Association between sex hormones and CRC. CRC, colorectal cancer. ER, estrogen receptor. E2, Estradiol/17β-estradiol. ORX, orchiectomy. OVX, ovariectomy. PGR, progesterone receptor. P4, progesterone. SHGB, sex hormone-binding globulin. T, testosterone. TAMs, tumor-associated macrophages. TIME, Tumor immune microenvironment

observation suggests that E2 may synergistically enhance the anti-tumor ability of anti-PD-L1 antibodies by downregulating PD-L1 expression and regulating the number of tumor-associated cells [30]. In MC38 model mice subjected to a high-fat diet and oophorectomy, the subcutaneous adipose tissue of oophorectomized (OVX) mice exhibited more severe macrophage-associated inflammation and higher expression of M2-like genes in TAMs than that of non-OVX female mice. This observation implies that obesity, macrophage-associated inflammation, and TAMs are potential mechanisms for inducing CRC in estrogen-deficient obese women [31]. Mahbub et al. [32, 33] demonstrated that CRC is associated with abnormally altered levels of E2, progesterone (P4), and their corresponding nuclear receptors in the colon. Both monotherapy and combination therapy with E2 and P4 decreased the number of malignant lesions and ERa expression. These treatments also increased the expression of $ER\beta$ and progesterone receptor (PGR), promoting cell cycle arrest in the Sub-G1 phase and apoptosis in AOM-male mice and human male colon cancer cell lines. Additionally, the combined use of E2 and 5-fluorouracil treatment exhibited superior anticancer effects than monotherapy on female and male primary CRC cells, with E2 monotherapy demonstrating the most considerable effects on male metastatic cells (SW620). These findings confirm the potential anti-tumor effects of estrogen in males [33].

Controversial findings Numerous studies have demonstrated the potential protective effect of estrogen against CRC; however, this relationship remains controversial. Yang et al. reported that pre-diagnosis levels of total estradiol, free estradiol, and estrone are positively associated with mortality risk in female CRC survivors [34]. Additionally, estrogen metabolite 2-MeO-E1 and COMT gene variants have been linked to CRC susceptibility in men [35]. In contrast, some studies failed to establish a significant association between estradiol levels and CRC risk [36-38]. Initial WHI trials suggested that short-term administration of estrogen plus progestin in postmenopausal women reduced CRC risk but led to more advanced-stage CRC diagnoses [39]. Consistently, an in vitro experiment demonstrated that exposure of colon cancer stem cells (CSCs) to 100 nM E2 for 48 h did not significantly affect cell viability and proliferation. However, prolonged exposure to E2 induced estrogen receptor expression on CSCs and inhibited the

interaction between CSCs and HUVEC, thereby promoting CSC migration and increasing metastasis rates [40]. These results suggest that extended estrogen use may promote malignant CRC progression, ultimately increasing mortality risk. Studies in ovariectomized mice have revealed a significant reduction in CRC liver metastases, an effect augmented by estradiol supplementation. Mechanistically, estrogen deficiency results in decreased accumulation of hepatic myeloid-derived suppressor cells (MDSCs), reduced levels of immunomodulatory active proteins, and increased CD8⁺ T cell populations, all of which inhibit CRC liver metastasis [41]. To address the controversy between estrogen and CRC, a study validated the role of estrogen in CRC pathobiology based on tumor characteristics. Their findings indicated that the pathobiological role of estrogen in postmenopausal CRC varies depending on patient age, tumor location (right or left hemicolon), pathological stage, histology, E2 concentration, ER β expression, and mismatch repair (MMR) status. Specifically, estrogen inhibits tumors characterized by L-Ca<70 (left-sided tumors and individuals aged<70 years), with non-medullary/mucinous (Med/Muc) histology or proficient MMR. Conversely, it promotes tumors with $R-Ca \ge 70$ (right-sided tumors and individuals aged \geq 70 years), Med/Muc histology, or deficient MMR [42]. These findings underscore the importance of considering patient age and tumor characteristics when assessing the role of estrogen in CRC.

ERβ

The involvement of sex hormones in the pathogenesis of CRC has been linked to ER β , which is a nuclear receptor expressed predominantly in the colonic mucosa and ubiquitous in the intestinal tract. It primarily exerts antiproliferative and apoptotic effects, contributing to the maintenance of normal epithelial function and tissue integrity [43]. Estrogen exerts its effects on colon cells mainly through ER β . The expression of ER β is reduced in human CRC tissues, adenomatous tissue, and familial adenomatous polyposis (FAP) tissues compared to normal tissues. This reduction in ERB appears to manifest early in the development of colorectal precancerous lesions [44–48]. Survival rate seems to be more favorable in patients with $ER\beta$ -positive CRC than in those with ERβ-negative CRC, with approximately 49% of the significant CpG sites being hypermethylated in $ER\beta$ -positive tumor [44]. Furthermore, among the ER β isoforms, the expression levels of ERB1 and ERB5 subtypes correlate positively with improved disease-free survival [45]. In an intestine-specific Er\beta-knockout (ERβ-KO) AOM/ DSS mouse model, both male and female mice exhibited increased intestinal ulcers and tumors, with male mice particularly affected. Subsequent in vitro organoid cultures and experiments with CRC cell lines validated that ERβ knockdown significantly induces TNFα expression and influences inflammatory signals such as NF- κ B [47]. Naringenin, a natural estrogen disruptor, binds to ERβ. Co-treatment of HT-29 cells with naringenin and the carcinogen bisphenol A revealed that naringenin, along with its fermented extract, reduces cell viability and induces apoptosis. Mechanistically, naringenin upregulates Erβ, subsequently regulating the expression of key genes in the P53 signaling pathway. This includes the upregulation of pro-apoptotic genes, such as FASLG and CASP2, and the downregulation of anti-apoptotic genes, such as BCL2 and MIR141, ultimately activating both extrinsic and intrinsic apoptosis pathways [49]. Collectively, these results suggest that intestinal ER^β plays a crucial role in repairing inflammatory damage in epithelial cells and inhibiting CRC development.

Progesterone and PGR

Progesterone is another female sex hormone that is critically involved in colorectal carcinogenesis, with its effects mediated through binding to the PGR. The existing literature on the association between progesterone and CRC is limited and contradictory. Some studies have reported no significant association between progesterone and CRC, and this consistency holds when analyzing colon and rectal cancer separately [16, 37, 38, 50]. Although a trend indicating an increased risk of CRC with 17-hydroxypregnenolone has been noted, it is not statistically significant [50]. However, other studies have suggested that low levels of progesterone and PGR are linked to a poorer prognosis in CRC [48, 51, 52]. Progesterone upregulates growth arrest and DNA damage-inducible protein alpha (GADD45 α), subsequently activating the JNK pathway, arresting the cell cycle, and inducing apoptosis, thereby inhibiting CRC progression [51]. Higher levels of $ER\beta$ and PGR in non-malignant tumor tissues from women aged \leq 50 years than in men and women aged \geq 60 years may contribute to the lower incidence of CRC in premenopausal women, as frequently reported [48]. Additionally, research has demonstrated that activation of the PGR is necessary for folic acid to inhibit the proliferation and migration of CRC cell lines [52, 53]. Collectively, these studies yield inconsistent results, warranting further investigation.

Briefly, most studies substantiate that estrogen exerts a protective effect against CRC in both men and women. A significant decline in estrogen levels in postmenopausal women is strongly linked to an elevated risk of CRC. Mechanistically, estrogen promotes cell cycle arrest and apoptosis primarily by inhibiting the NF-κB inflammatory pathway and tumor-associated cell populations, particularly M2-type macrophages and PD-L1-expressing cells. It regulates Nrf2-associated signaling, upregulates ER β and PGR, and suppresses ER α -regulated oncogenic pathways, thus preventing CRC development and progression. However, some studies have suggested a positive association between estrogen and CRC risk, suggesting that prolonged estrogen use may promote malignant progression. The pathobiological role of estrogen in postmenopausal CRC varies depending on patient age and tumor background. Conversely, the relationship between progesterone and CRC risk lacks significance, and the evidence remains insufficient. Therefore, the association of estrogen and progesterone with CRC is controversial. The duration of sex hormone use, patient age, and tumor characteristics must all be comprehensively considered when analyzing the role of female sex hormones in CRC (Fig. 1b).

Androgen and CRC

Androgen has been increasingly linked to CRC (Table 1). Numerous clinical studies have shown that higher levels of circulating testosterone and SHBG are associated with a decreased risk of CRC, while elevated free testosterone levels are positively correlated with CRC risk [19, 37, 54-57]. In addition, patients with prostate cancer who undergo androgen deprivation therapy have an increased risk of CRC, particularly distal colon adenocarcinoma [58]. In an AOM/DSS-induced colitisassociated cancer model, serum testosterone levels were significantly increased in orchiectomized (ORX) male and female mice after AOM/DSS treatment compared with those in controls. Orchiectomy (ORX) significantly reduced the incidence of colitis and distal colonic tumors in male mice. When supplemented with testosterone, the expression of inflammatory mediators COX-2 and iNOS increased, exacerbating colonic inflammation and greatly contributing to the development of submucosal invasive carcinoma [8]. Previous studies have confirmed that certain animal models of colon cancer, including Apc^{Pirc/+} rats, Apc^{Min/+} mice, and AOM-mice, exhibit increased male susceptibility to colon adenoma development [32, 59]. The reduction in endogenous hormone levels after ovariectomy did not alter the prevalence of adenomas in females, whereas androgen depletion during orchiectomy significantly inhibited adenoma development in Apc^{Pirc/+} rats and AOM-mice. Furthermore, androgen receptors were not detected in the colon and adenomas, suggesting that the sex difference in colon adenoma development may be an indirect tumor-promoting effect of testosterone rather than a protective effect of estrogen [59].

Conversely, some studies suggest that androgens may have a protective effect against CRC [34, 57, 60, 61]. Both men and women showed a negative correlation between free testosterone levels and CRC morbidity and mortality [34, 57]. In men, the risk of CRC was increased due to obesity-induced reductions in SHBG and testosterone levels [60]. Additionally, testosterone therapy was found to be negatively associated with distant-stage CRC [61]. Mori et al. [37] suggested that the association between SHBG levels and CRC risk may vary depending on total isoflavone intake. However, several studies have reported that circulating concentrations of testosterone, SHBG, androstenedione, and dehydroepiandrosterone (DHEA) are not associated with early precursor lesions of the colon or the risk of colon cancer [38, 62, 63].

In summary, sex differences in CRC risk and outcomes exist, and sex hormones, primarily estrogen and testosterone, are thought to play a significant role in this sex dimorphism. Although the relationship between sex hormones and CRC risk remains controversial, existing studies can provide partial insights into the sex dimorphism observed in CRC (Fig. 1). Considering the strong association between the gut microbiome and CRC development, coupled with the bidirectional interaction between sex hormones and the gut microbiome, we will further explore sexual dimorphism in the gut microbiome in CRC in the next section.

Sexual dimorphisms in the gut microbiome in CRC Influence of the gut microbiome on CRC

The gut microbiome is made up of approximately $10^{13}-10^{14}$ microorganisms that play a role in regulating host metabolism, inflammatory responses, immune system, and intestinal barrier function. Research has shown that the composition of the gut microbiome is strongly influenced by sex and age [9, 64]. In older adults, there is a decrease in Bifidobacteria and Lactobacilli, and a significant increase in Bacteroidetes and E. coli in the intestinal bacteria [65]. Among the primary microbial phyla in the gut microbiota, Firmicutes (F) and Bacteroidetes (B) are particularly important, and the F/B ratio is used as an indicator of the overall balance of the gut microbiota [66]. An elevated F/B ratio is generally associated with obesity and dietary habits, while a reduced F/B ratio is typically observed in cases of inflammatory bowel disease. However, the evidence for a correlation between this ratio and these diseases is not yet sufficient [66, 67].

In recent years, it has been increasingly recognized that the gut microbiome is involved in CRC development [13, 68, 69]. It has been shown that ecological dysregulation of the gut microbiome is a risk factor for CRC [70]. Previous studies have indicated that the F/B ratio increases from birth to adulthood and then gradually decreases with age [65]. However, new evidence suggests that the F/B ratio sequentially increases in younger volunteers, older volunteers, and older CRC patients [71]. These findings indicate that the same bacteria may have different functions in different populations. CRC is more prevalent in middle-aged and elderly populations, and specific bacteria colonizing the gut of young people may have a beneficial role in preventing inflammation and tumorigenesis. On the other hand, bacteria enriched in the guts of older individuals may promote colorectal tumorigenesis. At the phylum level, the dominant genera in CRC include Bacteroidetes, Faecalibacterium, E. coli, Fusobacterium nucleatum, Streptococcus gallolyticus, and Peptostreptococcus [72-74]. Several studies have shown that bacteria such as enterotoxigenic Bacteroides fragilis, E. coli, and Fusobacterium nucleatum are involved in the development of chronic colitis-associated cancers [74-76]. Moreover, higher abundance of Bacteroidetes has been associated with chemotherapy-related adverse effects and poorer prognosis [77, 78]. Additionally, among patients treated with immune checkpoint inhibitors, those with high abundance of Bacteroidetes have shown shortened progression-free survival compared to those with low abundance [78, 79]. Mechanistically, patients with relatively high abundance of Bacteroidetes exhibit limited intratumoral lymphocyte and myeloid cell infiltration, as well as diminished antigen-presenting capacity, resulting in impaired systemic and antitumor immunity [78]. These findings highlight the significant contribution of intestinal bacteria, particularly Bacteroidetes, in the progression of CRC and the potential for therapeutic intervention by modulating the gut microbiome in patients undergoing immune checkpoint inhibitor treatment.

Sex differences in the gut microbiome in clinical samples

Recent studies have highlighted differences in the gut microbiome composition between males and females [80–85]. These differences may contribute to the observed variations in CRC risk and outcomes between the sexes. To investigate the correlation between intestinal flora and sex, Lin et al. [80] generated a heatmap based on the sex of CRC patients. They discovered that females had higher levels of five bacterial species, including Prevotella sp. Marseille-P2931, Clostridium colinum, and Bifidobacterium pseudocatenulatum. On the other hand, male patients showed significant enrichment of 11 bacterial species, including Fusobacterium mortiferum, Bifidobacterium adolescentis, and Succinatimonas hippei. Another clinical study confirmed that Bacteroides are important bacteria associated with CRC [81]. In male CRC patients, the most detected bacteria were Bacteroides, Eubacterium, and Faecalibacterium, while in female patients, they were Bacteroides, Subdoligranulum, and Eubacterium. Additionally, Blautia, Barnesiella, and Anaerostipes were identified as the three bacteria that differed the most between male and female patients [81]. Liao et al. [82] also observed significant variations in microbial diversity, community structure, and microbial symbiosis between males and females during CRC development. They found that males had more stable gut microbial communities compared to females. As CRC advanced, the gut microbial β-diversity increased in males, without significant changes in α -diversity, resulting in more stochastic gut microbial communities with complex microbial symbioses. Conversely, both α - and β-diversity decreased significantly in females, leading to more deterministic microbial communities but with the absence of key species [82]. Previous research has demonstrated that Escherichia coli containing polyketide synthase (*pks*⁺*E. coli*) can promote the progression of CRC. In a cross-sectional study investigating the relationship between dietary intake and the prevalence of *pks*⁺*E. coli*, it was found that men had a significantly higher prevalence compared to women. This disparity may partially explain the difference in CRC incidence between the sexes [83]. A recent study discovered that healthy men had lower levels of intestinal lactobacilli and butyrateproducing bacteria compared to healthy women. However, these sex-related differences were not observed in patients with colonic adenomas and CRC. This suggests that the absence of certain probiotics could contribute to the higher incidence of CRC in men [85].

The gut microbiome in CRC not only exhibits sexual dimorphism in its composition and function but also influences sex-specific aspects of preventive and therapeutic approaches based on intestinal microbiome. For example, serum vitamin D levels are negatively associated with CRC risk and prognosis. Female patients with CRC are more susceptible to Fusarium nucleatum infections following vitamin D supplementation than their male counterparts [84]. Secondary bile acids, such as deoxycholic acid (DCA), are tumor-promoting bile acids produced by anaerobic colonic bacteria via 7α-dehydroxylation. Ursodeoxycholic acid (UDCA) mitigates the effects of DCA and inhibits colon cancer activity in mouse models [86]. In patients treated with UDCA, an increased abundance of Faecalibacterium prausnitzii and a deficiency of Ruminococcus gnavus were significantly associated with a higher risk of colon adenomas in men. However, such associations were not observed in women. These results imply that sex alters the activity of UDCA in the colon [87].

Sexual dimorphism in the gut microbiome of animal models

Sex differences

The sex-biased gut microbiome in CRC is also prevalent in animal models. Previous studies have indicated the presence of sex dimorphism in the epigenetic genes of the colons of prepubescent mice. However, these studies did not identify significant sex-based differences in the composition of the gut microbiome; rather, they suggested that sex primarily influences the dominance of specific taxa [88]. Recent investigations have revealed that Apc^{Min/+} and AOM/DSS male mice exhibit larger and more numerous colon tumors and greater severity of gut inflammation than female mice. Additionally, in male and pseudo-germ mice that received fecal samples from male mice or men, the intestinal pathogenic bacterium Akkermansia muciniphila was significantly enriched. Conversely, the abundance of the probiotic bacterium Parabacteroides goldsteinii was significantly reduced, leading to a deterioration in intestinal barrier function. Mechanistically, the predominant male-associated intestinal microbial metabolites activate the glycerophospholipid metabolic pathway, ultimately exacerbating CRC tumorigenesis. Therefore, the modulation of sex-biased gut microbiomes and associated metabolites may represent a potentially effective, sex-targeted strategy for both the prevention and treatment of CRC [89].

Microgenderome

The "microgenderome," referring to interactions between sex hormones and the gut microbiome, is an emerging area of study. Research data from various animal models have established the role of the "microgenderome" in CRC incidence. Song et al. [27] discovered that E2 inhibits AOM/DSS-induced tumorigenesis in male mice. They conducted a study to further investigated the effects of E2 on the gut microbiome. They found that E2 supplementation in males, AOM/DSS+E2 treatment in males and females, led to a significant increase in microbial diversity compared to normal male mice. This increase was indicated by OTU counts and the Chao 1 index. Furthermore, they observed a significant reduction in Bacteroides abundance and F/B ratio in AOM/DSS+E2 male mice compared to AOM/DSS mice [90]. Bacteroides are major promoters and facilitators of human CRC and have the potential to exacerbate tumorigenesis in the AOM/DSS model [76, 91]. Additionally, the ratio of commensal bacteria to opportunistic pathogens (C/O)was higher in E2-supplemented males and females than in normal males and females_OVX [90]. Building upon their discovery that E2 can inhibit MC38 tumor growth by reducing PD-L1 expression, Song et al. also found that E2 pretreatment prior to anti-PD-L1 treatment induced changes in the intestinal microbiota of MC38 mice. These changes included a decrease in the abundance of the Muribaculaceae family and opportunistic pathogens (Enterobacteriaceae group) and an increase in the abundance of the Ruminococcaceae family and commensal bacteria (Lactobacillus murinus group and P. goldsteinii).

These alterations may contribute to enhanced anti-tumor immunotherapy [92]. These findings indicate that estradiol has the ability to induce changes in the gut microbiome of colon tumor model mice. These changes include a decrease in the abundance of *Bacteroides* and the F/B ratio, an increase in the C/O ratio, and alterations in microbiota diversity. These alterations are closely linked to an improvement in anti-tumor immunity and a lower risk of CRC [90, 92].

Intestinal ER β serves as a protective factor against colitis-associated cancers. Studies have shown that colitis-induced CRC reduces gut microbiota diversity and enriches gram-negative bacteria, particularly in the absence of ERB. In AOM/DSS ERB-KO mice, males exhibited an enriched microbiota with functions related to cell motility, membrane transport, and carbohydrate metabolism, whereas females exhibited a reduced microbiota with functions primarily associated with metabolism and the endocrine system. This observation suggests that $ER\beta$ facilitates a more favorable gut microbiome, which, in turn, may contribute to the prevention of CRC development [93]. Prior research has demonstrated that Nrf2 can enhance the anti-inflammatory effects of estrogen by directly regulating ERß expression. Recent studies have also indicated that Nrf2 genotypes can alter gut microbiome composition. In AOM/DSS Nrf2_KO mice, changes in the abundance of Akkermansia muciniophila, L. murinus, and Bacteroides vulgatus were observed. These changes varied between sexes and the extent of CRC induction. Specifically, B. vulgatus abundance increased in both male and female AOM/DSS mice, whereas L. murinus abundance was decreased only in Nrf2 KO male mice. Furthermore, the abundance of (A)muciniophila increased in male mice, regardless of Nrf2 knockout. Here, L. murinus abundance was negatively correlated with the number of colon tumors, whereas (B) vulgatus abundance was positively correlated with inflammatory status, tumor count, and adenoma grade [94]. These findings suggest that Nrf2 may reduce the incidence and progression of colitis-associated cancers by regulating ER β expression and shaping a more favorable gut microbiota.

Recent reports suggest that the dysregulation of the intestinal microecology induced by testosterone may play a crucial role in the differences in colorectal carcinogenesis between sexes [95]. In the control group, male mice exhibited higher levels of *Firmicutes*, lower levels of *Bacteroidetes*, and higher F/B ratios compared to female and ORX mice. In the AOM/DSS and AOM/DSS + testosterone propionate (TP) groups, there were no significant differences in F/B ratios between females and males. However, in the ORX AOM/DSS group, the addition of TP did not result in a difference in *Bacteroidetes*, but it led to a decrease in the abundance of *Firmicutes*, which in turn decreased the F/B ratio. Additionally, a decline in microbial diversity and the C/O ratio was observed [95]. Testosterone supplementation increased the probability of infection with opportunistic pathogens (*Mucispirillum schaedleri* or *A. muciniphila*) in AOM/DSS females and ORX mice [95].

Microgenderome-based therapeutic approaches

Sexual dimorphism in gut flora plays a role in CRC treatment. Recent studies have revealed that anti-PD-L1 treatment reduces testosterone levels in male MC38 mice while significantly altering the composition of the gut microbiome in female mice [96]. When male mice were treated with narrow-spectrum antibiotics, they exhibited a more robust recovery of microbiota dysbiosis index (MDI), improved colitis, mitigation of testicular lipid metabolism disorders, and enhanced efficacy of anti-PD-L1 immunotherapy efficacy compared with those in female mice. At the genus level, the abundance of Lachnospiraceae was significantly lower in male mice than in females, whereas the converse was true for the Muribaculaceae family [96, 97]. Lachnospiraceae has been linked to immune checkpoint inhibitor responses [98], whereas Muribaculaceae plays a key role in AOM/DSS-induced CRC [99]. These findings underscore the potential significance of sex hormones as targets for enhancing the antitumor efficacy of anti-PD-L1. Moreover, they highlight the importance of considering sex differences in the gut microbiome when using antibiotics to manage immune checkpoint inhibitor-associated colitis.

Zearalenone (ZEA), an estrogenic mycotoxin, has shown promise in inhibiting colitis-associated colorectal tumorigenesis in male mice. It achieves this by increasing the abundance of SCFA-producing bacteria like unidentified Ruminococcaceae, Blautia, and Parabacteroidies while concurrently inhibiting the RAS/RAF/ERK pathway [100]. Additionally, the weakly estrogenic compound isoxanthohumol, along with its intestinal microbial metabolite 8-PN, demonstrates the ability to disrupt the cell cycle and inhibit the proliferation and invasion of colon cancer cells [101]. Helicobacter spp. is suspected to act as a deleterious stimulus, creating an environment of chronic intestinal inflammation that drives CRC development. Wolfe et al. uncovered complex dynamic changes in the intestinal microbiome when mice inoculated with the Th17-enhanced commensal candidate Candidatus Savagella, referred to as segmented filamentous bacteria (SFB), were exposed to *Helicobacter* spp. SFB⁺ male mice exhibited a significantly lower incidence of CRC than SFB⁺ female mice. The Enterobacteriaceae family, which ultimately developed CRC, was significantly increased in abundance exclusively in SFB⁺ mice. Moreover, the relative abundance of *Enterobacteriaceae* was higher in female mice than in male mice. These results suggest that SFB stabilizes the transgene against changes induced by *Helicobacter* spp. following inoculation and exerts a sexdependent protective effect in male mice [102].

Collectively, the gut microbiome in CRC is sexually dimorphic in composition, function, and microbebased therapeutics. The interplay among between sex, sex hormones, gut microbiota, and CRC is intricate and multifaceted (Fig. 2). In broad terms, estrogen and $ER\beta$ may mitigate CRC risk by fostering a more favorable gut microbiota. This includes altering the F/B ratio, increasing the C/O ratio, and enriching microbiota diversity, thereby enhancing anti-tumor immunity. Conversely, testosterone-induced dysregulation in the intestinal microecological may constitute a pivotal factor contributing to sex-based differences in colorectal carcinogenesis (Table 2). Understanding these relationships can aid in identifying individuals at risk for CRC development and optimizing treatment strategies for this disease. In the next section, we will delve further into the mechanisms governing the interaction between sex hormones and the gut microbiome.

Sex hormone-gut microbiome axis and its role in CRC

Generalizing the differences in the gut microbiome based on sex is challenging due to variations in study findings. However, researchers had identified several gut flora that were related to sex hormones. Emerging evidence supports the existence of the sex hormone–gut microbiome axis, which describes a bidirectional interaction between sex hormones and the gut microbiome. This interaction suggests that sex hormones can influence the composition and function of the gut microbiome and its metabolites, while the gut microbiome can also have a significant impact on sex hormone levels [9, 69] (Fig. 3).

Influence of sex hormones on the gut microbiome

The sex hormone–gut microbiome axis operates through multiple mechanisms (Fig. 3a). One important mechanism is the regulation of the gut microbiota composition by sex hormones. Estrogen, a major regulator of the gut microbiota, modulates the composition of gut microbiome by promoting the growth of bacteria that produce SCFAs (e.g., butyrate). These SCFAs play a role in inhibiting inflammatory signaling pathways, enhancing gut barrier function, and improving energy metabolism [92, 103, 104]. Progesterone supplementation increases the relative abundance of intestinal *Bifidobacterium* in women and mice during late pregnancy [105]. Conversely, testosterone increases the abundance of *Firmicutes* and decreases that of *Bacteroidetes* [95, 106, 107]. A study



Fig. 2 Sexual dimorphism in the gut microbiome in CRC. CRC, colorectal cancer. ERβ, estrogen receptor-beta. E2, Estradiol/17β-estradiol. ORX, orchiectomy. OVX, ovariectomy. SCFAs, short chain fatty acids. SFB, Segmented Filamentous Bacteria. T, testosterone. ZEA, Zearalenone

found that individuals with high estradiol or testosterone levels exhibited highly diverse gut microbiota. In men, testosterone levels positively correlated with the abundance of *Ruminococcus, Acinetobacter, Megamonas*, and *Dorea*. Conversely, women with higher estradiol levels exhibited a higher *Bacteroidetes* abundance and a lower *Firmicutes* abundance [108].

Sex hormones can also influence the immune response through the gut microbiota, which plays a crucial role in the development and function of the immune system. Commensal bacteria support the maturation of the intestinal mucosal immune system, while pathogenic bacteria can cause immune dysfunction [109]. Sex hormone receptors are widely expressed in immune cells, allowing sex hormones to directly regulate bacterial metabolism via these receptors [110]. Animal studies have revealed that E2 and estrogen-like compounds enhance the prevention of low-grade intestinal chronic inflammation by upregulating the expression and activity of intestinal alkaline phosphatase (IAP), an intestinal microbial-regulated anti-microbial peptide. This reduces the abundance

of Proteobacteria and lipopolysaccharide biosynthesis [111]. Knockdown of ER β in female mice disrupts the gut microbiota, enriching microbes that affect the immune system [93, 112]. Additionally, the gut microbiota influences T-cell development and antibody production in a sex-dependent manner [113]. However, earlier studies suggested that β -estradiol promotes IL-12 and IFN- γ production by dendritic cells, activating pro-inflammatory pathways. It also prolongs B-cell survival, creating an inflammatory microenvironment with altered intestinal permeability, which leads to the migration of gut microbiota into the lamina propria, promoting inflammation [9]. Androgens are generally considered immunosuppressive, but some studies have found that testosterone and DHEA promote the activation of regulatory T cells, thereby suppressing Th17-type responses [114]. Others have demonstrated that testosterone inhibits T-cell proliferation, modulates macrophage function, and promotes inflammatory responses [115]. These seemingly contradictory results align with previous research indicating that the roles of sex hormones and the gut

| Table 2 Summary of se | xual dimorphism in | the gut microbiome in CRC | | | |
|-----------------------------|------------------------------|---|--|--|------------|
| Model organism | Sex dimorphism | Change in Gut microbiome | | Findings | References |
| | | Increase (Enrich) | Reduce | | |
| CRC/adenoma patients | male | Emortiferum, B. adolescentis, S. hippei, P. gingivalis, A. intestini, Clostridium sp. A14, D. propionicifacien, M. smithii, B. massiliensis, F. varium, L. bacterium | | The composition and relative abundance of the gut microbiome in CRC would be influenced by sex | [80] |
| | female | Prevotella sp. Marseille, C. colinum, B. pseudo- catenulatum, Gordonibacter sp. Marseille, C. Saccharibacteria | | | |
| | male | Bacteroides, Eubacterium, Faecalibacterium | | | [81] |
| | female | Bacteroides, Subdoligranulum, Eubacterium | | | |
| CRC/adenoma patients | male | microbial β-diversity, rare species, more stochastic community structure | | The gut microbial communities are more stable in males than in females dur- | [82] |
| | female | more deterministic community structure | microbial $lpha$ - and eta -diversity, key species | ing the development of CRC | |
| Patients with pks + E. coli | male | pks + E. coli | | The prevalence of <i>pks</i> + <i>E. coli</i> was signifi- | [83] |
| | female | pks + E. coli | | cantly higher in men than women | |
| CRC patients | female | F. nucleatum | | Women are more susceptible to F. nuclea- tum following Vitamin D supplementation | [84] |
| Adenoma patients | male | F. prausnitzii | R. gnavus | Gender alters UDCA activity in the CRC | [87] |
| | female | no difference | | | |
| Min/+ and AOM/DSS mice | male | C.aerofaciens, D. bacterium, A. muciniphila, A. inops | P. goldsteinii, L.taiwanensis, L. fermentum | Gender-biased gut microbiome and metabolites favor sex dimorphism | [89] |
| | female | microbial diversity, beneficial bacteria | harmful bacteria | in CRC | |
| AOM/DSS mice | E2-treated male male | microbial diversity C/O ratio | Bacteroides, F/B ratio | E2 may induce changes in the gut micro- biota, thereby reducing the risk of CRC | [06] |
| | female_OVX | | C/O ratio | | |
| MC38 tumor model mice | αPD-L1 + E2 male | family <i>Ruminococcaceae, L. murinus</i> group and <i>P. goldsteinii</i> | family <i>Muribaculaceae</i> , Enterobacteriaceae group | E2 pre-treatment prior to anti-PD-L1 therapy induces changes in the gut micro- | [92] |
| | αPD-L1 male αPD-L1 female | P. goldsteinii | | biome of MC38 mice, thereby contributing to anti-tumor therapy | |
| AOM/DSS mice | male | Parasutterella | microbiota diversity | ERB facilitates a more favorable gut micro- | [93] |
| | fenale | Prevotellaceae_UCG_001 | microbiota diversity | biome, which may prevent the develop- | |
| | ERB_KO | Rikenellaceae_RC9; Lachnospiracae_ UCG_010 | microbiota α- diversity, Chao-1index | ment of נאני | |

| Model organism | Sex dimorphism | Change in Gut microbiome | | Findings | References |
|--------------------------------|--|---|---|---|------------|
| | | Increase (Enrich) | Reduce | | |
| AOM/DSS mice | male | A. muciniophila; B. vulgatus | microbiota a- diversity | Alterations in intestinal flora composi- | [94] |
| | female | <i>B</i> . abundance | L. murinus | tion by NrT2 depend on gender and LKC induction | |
| | male Nrf2_KO | A. muciniophila | microbial α-diversity, L. murinus | | |
| | female Nrf2_ KO | No difference | | | |
| AOM/DSS mice | male_ORX | microbial diversity; F/B ratio | | Testosterone-induced dysbiosis of the gut | [95] |
| | Male_ORX +TP | | microbial diversity, F/B ratio; C/O ratio | microbiota may be a factor in sex differ- | |
| | TP- male and female | opportunistic pathogens (<i>M. schaedleri</i> or <i>A. muciniphila</i>) | C/O ratio | בוורבא ווו רסוסוברומו רמורוווסלבוובאא | |
| | female | microbial diversity, beneficial bacteria | harmful bacteria | | |
| MC38 tumor model mice | male | Family <i>Muribaculaceae</i> | microbial a-diversity, <i>Lachnospiraceae</i> group | Sex differences in the gut microbiota should be considered when applying | [76, 97] |
| | female | Lachnospiraceae group | microbial a-diversity, Family <i>Muribacu-</i> <i>laceae</i> | antibiotics for the treatment of immune checkpoint inhibitor-associated colitis | |
| AOM/DSS mice | ZEA ⁺ male | unidentified <i>Ruminococcaceae, Parabac-</i> <i>teroidies, Blautia</i> ; microbial community stability | Microbial community vulnerability | Zearalenone increases SCFAs-producing intestinal microbiome with good inhibitory effects on CRC | [100] |
| Smad3 ^{-/-} CAC model | SFB ⁺ female SFB ⁺ female | Helicobacter spp., SFB Helicobacter spp., SFB, family Enterobacte- riaceae | | SFB has a sex-dependent protective effect in CRC male mice | [102] |

AUMUAS AZOXJMENTARE/GEXTRA SOCIUM SUITARE, LAL CORRECTAI CANCET, C/O COMMENSAI DECRENA/OPPORTUNISTIC PARIAGIO/ 1/15-ESTRAGIO/, LAG ESTOGEN RECEPTOR-DETA, F/B FITMICUTES/BACTERFORDERES, OVX Ovariectomy, ORX Orchiectomy, pks + E. coli Escherichia coli containing polyketide synthase, SCFAs Short chain fatty acids, SFB Segmented Filamentous Bacteria, TP Testosterone propionate, UDCA Ursodeoxycholic acid, ZEA Zearalenone



Fig. 3 Sex hormone–gut microbiome axis and its role in CRC. a Influence of sex hormones on the composition and function of the gut microbiome. b Potential mechanisms through which the gut microbiome regulates sex hormone levels. AR, androgen receptor. CRC, colorectal cancer. ER, estrogen receptor. P4, progesterone. UGT, UDP-glucuronosyltransferase

microbiome in the development of CRC are intricate and diverse. Consequently, further studies are required to validate these findings.

Potential mechanisms through which the gut microbiome regulates sex hormone levels

Sex hormones have the ability to impact the composition and diversity of the gut microbiome. This, in turn, plays a role in regulating the excretion and cycling of sex hormones. The gut microbiome can regulate host sex hormone levels through a variety of potential mechanisms (Fig. 3b). Firstly, specific enzymes called UDP-glucuronosyltransferases (UGTs) in the liver catalyze the glucuronidation of sex hormones, which are then excreted into the urine or eliminated through the bile duct into the intestine [116]. These conjugated estrogens and androgens are deconjugated by β -glucuronidase, which is an enzyme responsible for breaking down sex hormones into their active forms. This process results in the formation of biologically active free sex hormones that are reabsorbed through the enterohepatic circulation, ultimately influencing downstream physiological effects. Dysregulation of the intestinal microbiota can lead to reduced deconjugation, resulting in decreased levels of free estrogens [103, 117–119]. The gut microbiome can metabolize both endogenous and exogenous estrogens, with the metabolites produced having an impact on the host. When estrogen-metabolizing bacteria are deficient, circulating estrogen levels decrease, which may be a risk factor for certain conditions, including cancer. The gut microbiota is a crucial regulator of androgen metabolism. Compared with female mice, male mice exhibit significantly elevated β -glucuronidase activity, high concentrations of glucuronidase testosterone and dihydrotestosterone (DHT) in their small intestinal contents, and increased levels of free DHT in the distal intestine or feces. However, germ-free mice show low levels of free DHT in the distal intestine or feces that the gut microbiota influences the intestinal metabolism of testosterone and DHT [9, 117].

Secondly, emerging evidence strongly supports the potential involvement of the gut microbiota in the biosynthesis of various sex steroid hormones. This is primarily achieved through the activity of specific bacterial enzymes, including 17, 20 lyase, hydroxysteroid dehydrogenase (HSD), and steroid reductase [120, 121]. However, the available evidence on bacterial enzymes is currently limited, and further research is required to confirm these findings. Additionally, human fecal bacteria can oxidize, reduce, and hydrolyze estrogens and androgens [14]. Thirdly, the gut microbiome may influence sex hormone levels by directly affecting gonadal function. Certain mucus-degrading bacteria play a role in maintaining mucus function. Disruption of the gut mucosal barrier can facilitate the entry of microbiota from the intestinal lumen into the circulation, potentially inducing systemic chronic inflammation. Such inflammation can inhibit testosterone production by testicular cells [14, 122]. In contrast, some intestinal microorganisms are involved in regulating the secretion of neuromodulators in the gut–brain axis, which can influence the production of endogenous androgens and estrogens via the hypothalamic–pituitary–testis or ovary axis [123, 124].

Role of the sex hormone-gut microbiome axis in CRC

Sex steroid hormones, particularly estrogen and testosterone, are believed to play a pivotal role in the sexual dimorphism of CRC risk and outcome. Extensive research consistently shows that estrogen has a protective effect against CRC in both men and women. Conversely, higher levels of free testosterone are associated with an increased risk of CRC. Recent studies have identified differences in the gut microbiome between males and females in the context of CRC. New evidence now supports the existence of a sex hormone-gut microbiome axis, which may explain the observed disparities in CRC risk and outcomes between the sexes. Mechanistically, estrogen enhances anti-tumor immunity by promoting a more favorable gut microbiota. This involves increasing the C/O ratio and enriching microbiota diversity by inhibiting inflammatory pathways, enhancing Nrf2-associated signaling, and modulating the populations of tumor-associated immune cell. Conversely, testosterone directly promotes tumor growth by triggering inflammatory responses, reducing microbiota diversity, and increasing the proportion of opportunistic pathogens. At the molecular level, there are bidirectional interactions between sex hormones and the gut microbiome. Sex hormones not only regulate the composition and diversity of the gut microbiome but also modulate the host immune response through the gut microbiome. The gut microbiome, in turn, may be involved in the biosynthesis and metabolism of sex hormones by expressing certain enzymes, such as β -glucuronidase and bacterial enzymes. Additionally, the gut microbiota may directly affect gonadal function. However, the available studies present conflicting findings. Some propose that estrogens may increase CRC risk, while androgens seem to confer protection against CRC. Nevertheless, most of these findings are based on clinical samples, and further validation through animal and cellular models is needed.

The prospect of combination therapy, targeting both sex hormones and modulation of the gut microbiome, holds great potential for enhancing outcomes in patients with CRC. Hormonal therapy can be complemented by interventions that modify the gut microbiome composition towards a more protective profile. For example, supplementation of E2 before anti-PD-L1 treatment has been shown to alter the gut microbial composition of MC38 colon tumor-bearing mice, creating a favorable microecological environment and thereby enhancing the anti-tumor efficiency of anti-PD-L1 treatment [92]. On the other hand, manipulating the gut microbiome and its metabolites can also enhance anti-tumor effects. For instance, a combination of anti-PD-L1 with antibiotic colistin treatment significantly reduced testosterone levels in male MC38 mice, leading to increased immunotherapy efficiency [96]. Common methods for modulating the gut microbiome, such as fecal microbiota transplantation and probiotic supplementation, have the potential to modulate sex hormone levels, thereby enhancing the efficiency of immunotherapy [14]. The identification of specific microbial taxa, metabolites, and drugs that augment the antitumor effects of hormonal therapy is an active area of research that paves the way for novel CRC treatment strategies. Future studies should further explore the intricate interactions among sex hormones, the gut microbiome, and host immunity to provide innovative approaches to CRC treatment.

Conclusion

This review emphasizes the significant influence of sex hormones and the gut microbiome in the development of CRC. Modulating sex hormones or the gut microbiome could have potential clinical implications in preventing and treating CRC. A more thorough comprehension of the interactions among sex hormones, the gut microbiome, and colorectal carcinogenesis could pave the way for novel therapeutic strategies. However, it is important to note that current research in this field is still in its early stages and remains controversial. Further investigation is imperative to gain a deeper understanding of the underlying mechanisms and to optimize therapeutic approaches in this context.

Abbreviations

| AOM/DSS | Azoxymethane/dextran sodium sulfate |
|---------|--|
| C/O | Commensal bacteria/opportunistic pathogens |
| CRC | Colorectal cancer |
| CSCs | Cancer stem cells |
| DCA | Deoxycholic acid |
| DHEA | Dehydroepiandrosterone |
| DHT | Dihydrotestosterone |
| E2 | Estradiol/17β-estradiol |
| ERβ | Estrogen receptor- beta |
| F/B | Firmicutes/Bacteroidetes |
| FAP | Familial adenomatous polyposis |
| HSD | Hydroxysteroid dehydrogenase |
| MDI | Microbial dysbiosis index (MDI) |
| MDSC | Myeloid-derived suppressor cells |
| Med/Muc | Medullary/mucinous histology |

| MMR | Mismatch repair |
|-------|--------------------------------|
| ORX | Orchiectomy |
| OVX | Ovariectomy |
| PGR | Progesterone receptor |
| SCFAs | Short chain fatty acids |
| SFB | Segmented filamentous bacteria |
| SHBG | Sex hormone-binding globulin |
| TAMs | Tumor-associated macrophages |
| TIME | Tumor immune microenvironment |
| TP | Testosterone propionate |
| UDCA | Ursodeoxycholic acid |
| UGT | UDP-glucuronosyltransferase |
| ZEA | Zearalenone |

Acknowledgements

Not applicable.

Authors' contributions

Z.H.W. and Y.Q.H. designed the review protocol, conducted the search, drafted the manuscript, and prepared all Figures and tables. R.Y.Z. and C.Z. screened potentially eligible studies, extracted and analyzed data, and updated reference lists. C.X. and X.K.L. were responsible for the funding acquisition. F.M.Y., M.W., and X.K.L. contributed to the design of the review protocol, arbitrating potentially eligible studies, and interpreting results. C.X. and X.K.L. provided feedback on the report. All authors read and approved the final manuscript.

Funding

This research was supported by the Natural Science Foundation of Sichuan, grant number 2023NSFSC1830, and the China Postdoctoral Science Foundation, grant number 2022MD723718.

Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

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Received: 28 October 2023 Accepted: 1 March 2024 Published online: 07 March 2024

References

- Siegel RL, Wagle NS, Cercek A, Smith RA, Jemal A. Colorectal cancer statistics, 2023. CA Cancer J Clin. 2023;73(3):233–54.
- Patel SG, Karlitz JJ, Yen T, Lieu CH, Boland CR. The rising tide of earlyonset colorectal cancer: a comprehensive review of epidemiology, clinical features, biology, risk factors, prevention, and early detection. Lancet Gastroenterol Hepatol. 2022;7(3):262–74.
- Siegel RL, Miller KD, Wagle NS, Jemal A. Cancer statistics, 2023. CA Cancer J Clin. 2023;73(1):17–48.
- Baraibar I, Ros J, Saoudi N, Salva F, Garcia A, Castells MR, Tabernero J, Elez E. Sex and gender perspectives in colorectal cancer. ESMO Open. 2023;8(2):101204.

- 5. Wele P, Wu X, Shi H. Sex-dependent differences in Colorectal Cancer: with a focus on obesity. Cells. 2022;11(22):3688.
- Banibakhsh A, Sidhu D, Khan S, Haime H, Foster PA. Sex steroid metabolism and action in colon health and disease. J Steroid Biochem Mol Biol. 2023;233:106371.
- Mahbub AA. Therapeutic strategies and potential actions of female sex steroid hormones and their receptors in Colon cancer based on preclinical studies. Life (Basel). 2022;12(4):605.
- Song C-H, Kim N, Nam RH, Choi SI, Yu JE, Nho H, Shin E, Lee H-N, Surh Y-J. Testosterone strongly enhances azoxymethane/dextran sulfate sodium-induced colorectal cancer development in C57BL/6 mice. Am J cancer Res. 2021;11(6):3145–62.
- 9. Yoon K, Kim N. Roles of sex hormones and gender in the gut microbiota. J Neurogastroenterol Motil. 2021;27(3):314–25.
- Kim YS, Unno T, Kim BY, Park MS. Sex differences in gut microbiota. World J Mens Health. 2020;38(1):48–60.
- 11. Heintz-Buschart A, Wilmes P. Human gut microbiome: function matters. Trends Microbiol. 2018;26(7):563–74.
- DeDecker L, Coppedge B, Avelar-Barragan J, Karnes W, Whiteson K. Microbiome distinctions between the CRC carcinogenic pathways. Gut Microbes. 2021;13(1):1854641.
- Kim J, Lee HK. Potential role of the gut Microbiome in Colorectal Cancer Progression. Front Immunol. 2021;12:807648.
- Wang L, Tang L, Zhai D, Song M, Li W, Xu S, Jiang S, Meng H, Liang J, Wang Y, et al. The role of the sex hormone-gut microbiome axis in tumor immunotherapy. Gut Microbes. 2023;15(1):2185035.
- Ko SH, Jung Y. Energy Metabolism Changes and dysregulated lipid metabolism in Postmenopausal Women. Nutrients. 2021;13(12):4556.
- Murphy N, Strickler HD, Stanczyk FZ, Xue X, Wassertheil-Smoller S, Rohan TE, Ho GY, Anderson GL, Potter JD, Gunter MJ. A prospective evaluation of endogenous sex hormone levels and colorectal Cancer risk in Postmenopausal Women. J Natl Cancer Inst. 2015;107(10):djv210.
- Semlali A, Reddy Parine N, Arafah M, Mansour L, Azzi A, Al Shahrani O, Al Amri A, Shaik JP, Aljebreen AM, Alharbi O, et al. Expression and polymorphism of toll-like receptor 4 and effect on NF-kappaB mediated inflammation in Colon cancer patients. PLoS ONE. 2016;11(1):e0146333.
- Razak S, Alam I, Afsar T, Abulmeaty MMA, Almajwal A, Jahan S. A Prospective Evaluation of Serum Vitamin D (1, 25(OH)(2) D(3)) and endogenous sex hormone levels in Colorectal Cancer patients. Front Oncol. 2019;9:468.
- Lin JH, Zhang SM, Rexrode KM, Manson JE, Chan AT, Wu K, Tworoger SS, Hankinson SE, Fuchs C, Gaziano JM, et al. Association between sex hormones and colorectal cancer risk in men and women. Clin Gastroenterol Hepatol. 2013;11(4):419–424e411.
- Ma W, Song M, Kvaerner AS, Prescott J, Chan AT, Giovannucci EL, Zhang X. Sex-specific association between Family History of Diabetes and Risk of Colorectal Cancer: two prospective cohort studies. Cancer Prev Res (Phila). 2018;11(9):535–44.
- Luo G, Zhang Y, Wang L, Huang Y, Yu Q, Guo P, Li K. Risk of colorectal cancer with hysterectomy and oophorectomy: a systematic review and meta-analysis. Int J Surg. 2016;34:88–95.
- Segelman J, Lindstrom L, Frisell J, Lu Y. Population-based analysis of colorectal cancer risk after oophorectomy. Br J Surg. 2016;103(7):908–15.
- Rutegard M, Moshtaghi-Svensson J, Weibull CE, Ottander U, Nordenvall C, Sund M. Exposure to oestrogen and risk of anastomotic leakage after colorectal cancer surgery - A clue to the different leak rates in men and women. Colorectal Dis. 2023;25(1):9–15.
- Moravcik R, Olejarova S, Zlacka J, Herichova I. Effect of miR-34a on the expression of clock and clock-controlled genes in DLD1 and Lovo human cancer cells with different backgrounds with respect to p53 functionality and 17beta-estradiol-mediated regulation. PLoS ONE. 2023;18(10):e0292880.
- Son HJ, Kim N, Song CH, Lee SM, Lee HN, Surh YJ. 17beta-Estradiol reduces inflammation and modulates antioxidant enzymes in colonic epithelial cells. Korean J Intern Med. 2020;35(2):310–9.
- Son HJ, Sohn SH, Kim N, Lee HN, Lee SM, Nam RH, Park JH, Song CH, Shin E, Na HY, et al. Effect of Estradiol in an Azoxymethane/Dextran sulfate sodium-treated mouse model of Colorectal Cancer: implication for sex difference in Colorectal Cancer Development. Cancer Res Treat. 2019;51(2):632–48.

- Song CH, Kim N, Lee SM, Nam RH, Choi SI, Kang SR, Shin E, Lee DH, Lee HN, Surh YJ. Effects of 17beta-estradiol on colorectal cancer development after azoxymethane/dextran sulfate sodium treatment of ovariectomized mice. Biochem Pharmacol. 2019;164:139–51.
- Javid SH, Moran AE, Carothers AM, Redston M, Bertagnolli MM. Modulation of tumor formation and intestinal cell migration by estrogens in the apc(Min/+) mouse model of colorectal cancer. Carcinogenesis. 2005;26(3):587–95.
- Song CH, Kim N, Kim DH, Lee HN, Surh YJ. 17-beta estradiol exerts antiinflammatory effects through activation of Nrf2 in mouse embryonic fibroblasts. PLoS ONE. 2019;14(8):e0221650.
- Song CH, Kim N, Nam RH, Choi SI, Jang JY, Kim JW, Na HY, Lee HN. Combination treatment with 17beta-estradiol and anti-PD-L1 suppresses MC38 tumor growth by reducing PD-L1 expression and enhancing M1 macrophage population in MC38 colon tumor model. Cancer Lett. 2022;543:215780.
- Bader J, Carson M, Enos R, Velazquez K, Sougiannis, Singh U, Becker W, Nagarkatti M, Fan D, Alexander, et al. High-fat diet-fed ovariectomized mice are susceptible to accelerated subcutaneous tumor growth potentially through adipose tissue inflammation, local insulin-like growth factor release, and tumor associated macrophages. Oncotarget. 2020;11(49):4554–69.
- Mahbub AA, Aslam A, Elzubier ME, El-Boshy M, Abdelghany AH, Ahmad J, Idris S, Almaimani R, Alsaegh A, El-Readi MZ, et al. Enhanced anticancer effects of oestrogen and progesterone co-therapy against colorectal cancer in males. Front Endocrinol (Lausanne). 2022;13:941834.
- Mahbub AA. 17beta-estradiol enhances 5-Fluorouracil anti-cancer activities in Colon cancer cell lines. Med Sci (Basel). 2022;10(4):62.
- Yang W, Giovannucci EL, Hankinson SE, Chan AT, Ma Y, Wu K, Fuchs CS, Lee IM, Sesso HD, Lin JH, et al. Endogenous sex hormones and colorectal cancer survival among men and women. Int J Cancer. 2020;147(4):920–30.
- Li S, Chen Y, Xie L, Meng Y, Zhu L, Chu H, Gu D, Zhang Z, Du M, Wang M. Sex hormones and genetic variants in hormone metabolic pathways associated with the risk of colorectal cancer. Environ Int. 2020;137:105543.
- Lavasani S, Chlebowski RT, Prentice RL, Kato I, Wactawski-Wende J, Johnson KC, Young A, Rodabough R, Hubbell FA, Mahinbakht A, et al. Estrogen and colorectal cancer incidence and mortality. Cancer. 2015;121(18):3261–71.
- Mori N, Sawada N, Iwasaki M, Yamaji T, Goto A, Shimazu T, Inoue M, Murphy N, Gunter MJ, Tsugane S. Circulating sex hormone levels and colorectal cancer risk in Japanese postmenopausal women: the JPHC nested case-control study. Int J Cancer. 2019;145(5):1238–44.
- Mori N, Keski-Rahkonen P, Gicquiau A, Rinaldi S, Dimou N, Harlid S, Harbs J, Van Guelpen B, Aune D, Cross AJ, et al. Endogenous circulating sex hormone concentrations and Colon cancer risk in Postmenopausal women: a prospective study and Meta-analysis. JNCI Cancer Spectr. 2021;5(6):pkab084.
- Chlebowski RT, Wactawski-Wende J, Ritenbaugh C, Hubbell FA, Ascensao J, Rebecca J, Rodabough, Rosenberg CA, Taylor VM, Harris R, Chen C, et al. Estrogen plus Progestin and Colorectal Cancer in Postmenopausal Women. N Engl J Med. 2004;350(10):991–1004.
- Zamani ARN, Avci CB, Ahmadi M, Pouyafar A, Bagheri HS, Fathi F, Heidarzadeh M, Rezaie J, Mirhosseini Y, Saberianpour S, et al. Estradiol modulated colorectal cancer stem cells bioactivity and interaction with endothelial cells. Life Sci. 2020;257:118078.
- Milette S, Hashimoto M, Perrino S, Qi S, Chen M, Ham B, Wang N, Istomine R, Lowy AM, Piccirillo CA, et al. Sexual dimorphism and the role of estrogen in the immune microenvironment of liver metastases. Nat Commun. 2019;10(1):5745.
- 42. Honma N, Arai T, Matsuda Y, Fukunaga Y, Akishima-Fukasawa Y, Yamamoto N, Kawachi H, Ishikawa Y, Takeuchi K, Mikami T. Estrogen concentration and estrogen receptor-beta expression in postmenopausal colon cancer considering patient/tumor background. J Cancer Res Clin Oncol. 2022;148(5):1063–71.
- Garcia-Villatoro EL, Allred CD. Estrogen receptor actions in colitis. Essays Biochem. 2021;65(6):1003–13.
- 44. Neumeyer S, Popanda O, Edelmann D, Butterbach K, Toth C, Roth W, Blaker H, Jiang R, Herpel E, Jakel C, et al. Genome-wide DNA

methylation differences according to oestrogen receptor beta status in colorectal cancer. Epigenetics. 2019;14(5):477–93.

- 45. Stevanato Filho PR, Aguiar Junior S, Begnami MD, Kuasne H, Spencer RM, Nakagawa WT, Bezerra TS, Kupper BC, Takahashi RM, Barros Filho M, et al. Oestrogen receptor beta isoform expression in sporadic colorectal cancer, familial adenomatous polyposis and progressive stages of colorectal cancer. BMC Cancer. 2017;17(1):754.
- Das PK, Saha J, Pillai S, Lam AK, Gopalan V, Islam F. Implications of estrogen and its receptors in colorectal carcinoma. Cancer Med. 2023;12(4):4367–79.
- Hases L, Indukuri R, Birgersson M, Nguyen-Vu T, Lozano R, Saxena A, Hartman J, Frasor J, Gustafsson JA, Katajisto P, et al. Intestinal estrogen receptor beta suppresses colon inflammation and tumorigenesis in both sexes. Cancer Lett. 2020;492:54–62.
- Refaat B, Aslam A, Idris S, Almalki AH, Alkhaldi MY, Asiri HA, Almaimani RA, Mujalli A, Minshawi F, Alamri SA, et al. Profiling estrogen, progesterone, and androgen receptors in colorectal cancer in relation to gender, menopausal status, clinical stage, and tumour sidedness. Front Endocrinol (Lausanne). 2023;14:1187259.
- 49. Lozano-Herrera SJ, Luna-Barcenas G, Guevara-Gonzalez RG, Campos-Vega R, Solis-Sainz JC, Hernandez-Puga AG, Vergara-Castaneda HA. Fermentation extract of Naringenin increases the expression of estrogenic receptor beta and modulates genes related to the p53 signalling pathway, miR-200c and miR-141 in human Colon cancer cells exposed to BPA. Molecules. 2022;27(19):6588.
- Michels KA, Geczik AM, Bauer DC, Brinton LA, Buist DSM, Cauley JA, Dallal CM, Falk RT, Hue TF, Lacey JV, editors. Jr. : Endogenous Progestogens and Colorectal Cancer Risk among Postmenopausal Women. *Cancer Epidemiol Biomarkers Prev* 2021, 30(6):1100–1105.
- Zhang YL, Wen XD, Guo X, Huang SQ, Wang TT, Zhou PT, Li W, Zhou LF, Hu YH. Progesterone suppresses the progression of colonic carcinoma by increasing the activity of the GADD45alpha/JNK/c–Jun signalling pathway. Oncol Rep. 2021;45(6):95.
- Wang HC, Huo YN, Lee WS. Activation of progesterone receptor is essential for folic acid-regulated cancer cell proliferation and migration. J Nutr Biochem. 2023;112:109205.
- Kuo CT, Lee WS. Progesterone receptor activation is required for folic acid-induced anti-proliferation in colorectal cancer cell lines. Cancer Lett. 2016;378(2):104–10.
- Liu Z, Zhang Y, Lagergren J, Li S, Li J, Zhou Z, Hu Z, Xie SH. Circulating sex hormone levels and risk of gastrointestinal Cancer: systematic review and Meta-analysis of prospective studies. Cancer Epidemiol Biomarkers Prev. 2023;32(7):936–46.
- 55. Harbs J, Rinaldi S, Gicquiau A, Keski-Rahkonen P, Mori N, Liu X, Kaaks R, Katzke V, Schulze MB, Agnoli C, et al. Circulating sex hormone levels and Colon cancer risk in men: a nested case-control study and Metaanalysis. Cancer Epidemiol Biomarkers Prev. 2022;31(4):793–803.
- Hang D, He X, Kvaerner AS, Chan AT, Wu K, Ogino S, Hu Z, Shen H, Giovannucci EL, Song M. Plasma sex hormones and risk of conventional and serrated precursors of colorectal cancer in postmenopausal women. BMC Med. 2021;19(1):18.
- McMenamin UC, Liu P, Kunzmann AT, Cook MB, Coleman HG, Johnston BT, Cantwell MM, Cardwell CR. Circulating sex hormones are Associated with gastric and colorectal cancers but not esophageal adenocarcinoma in the UK Biobank. Am J Gastroenterol. 2021;116(3):522–9.
- Shore R, Zhang J, Ye W, Stattin P, Lindblad M. Risk of colorectal adenocarcinoma in men receiving androgen deprivation therapy for prostate cancer; a nationwide cohort study. Cancer Causes Control. 2023;34(11):949–61.
- Amos-Landgraf JM, Heijmans J, Wielenga MC, Dunkin E, Krentz KJ, Clipson L, Ederveen AG, Groothuis PG, Mosselman S, Muncan V, et al. Sex disparity in colonic adenomagenesis involves promotion by male hormones, not protection by female hormones. Proc Natl Acad Sci U S A. 2014;111(46):16514–9.
- Dashti SG, Viallon V, Simpson JA, Karahalios A, Moreno-Betancur M, English DR, Gunter MJ, Murphy N. Explaining the link between adiposity and colorectal cancer risk in men and postmenopausal women in the UK Biobank: a sequential causal mediation analysis. Int J Cancer. 2020;147(7):1881–94.
- 61. Butler EN, Zhou CK, Curry M, McMenamin U, Cardwell C, Bradley MC, Graubard BI, Cook MB. Testosterone therapy and cancer risks

among men in the SEER-Medicare linked database. Br J Cancer. 2023;128(1):48–56.

- 62. Figueiredo JC, Gresham G, Barry EL, Mott LA, Passarelli MN, Bradshaw PT, Anderson CW, Baron JA. Circulating sex hormones and risk of colorectal adenomas and serrated lesions in men. Cancer Epidemiol Biomarkers Prev. 2022;31(1):293–5.
- Dimou N, Mori N, Harlid S, Harbs J, Martin RM, Smith-Byrne K, Papadimitriou N, Bishop DT, Casey G, Colorado-Yohar SM, et al. Circulating levels of testosterone, sex hormone binding globulin and colorectal Cancer risk: observational and mendelian randomization analyses. Cancer Epidemiol Biomarkers Prev. 2021;30(7):1336–48.
- 64. Ghosh TS, Shanahan F, O'Toole PW. The gut microbiome as a modulator of healthy ageing. Nat Rev Gastro Hepat. 2022;19(9):565–84.
- Mariat D, Firmesse O, Levenez F, Guimarães VD, Sokol H, Doré J, Corthier G, Furet JP. The Firmicutes/Bacteroidetes ratio of the human microbiota changes with age. BMC Microbiology. 2009;9(1):123.
- Magne F, Gotteland M, Gauthier L, Zazueta A, Pesoa S, Navarrete P, Balamurugan R. The Firmicutes/Bacteroidetes ratio: a relevant marker of gut dysbiosis in obese patients? Nutrients. 2020;12(5):1474.
- 67. Stojanov S, Berlec A, Strukelj B. The influence of Probiotics on the Firmicutes/Bacteroidetes ratio in the Treatment of Obesity and inflammatory bowel disease. Microorganisms. 2020;8(11):1715.
- Wong SH, Yu J. Gut microbiota in colorectal cancer: mechanisms of action and clinical applications. Nat Rev Gastroenterol Hepatol. 2019;16(11):690–704.
- Santos-Marcos JA, Mora-Ortiz M, Tena-Sempere M, Lopez-Miranda J, Camargo A. Interaction between gut microbiota and sex hormones and their relation to sexual dimorphism in metabolic diseases. Biol Sex Differ. 2023;14(1):4.
- Janney A, Powrie F, Mann EH. Host-microbiota maladaptation in colorectal cancer. Nature. 2020;585(7826):509–17.
- Zhang YK, Zhang Q, Wang YL, Zhang WY, Hu HQ, Wu HY, Sheng XZ, Luo KJ, Zhang H, Wang M, et al. A comparison study of Age and Colorectal Cancer-related gut Bacteria. Front Cell Infect Microbiol. 2021;11:606490.
- 72. Rebersek M. Gut microbiome and its role in colorectal cancer. BMC Cancer. 2021;21(1):1325.
- Fong W, Li Q, Yu J. Gut microbiota modulation: a novel strategy for prevention and treatment of colorectal cancer. Oncogene. 2020;39(26):4925–43.
- Quaglio AEV, Grillo TG, De Oliveira ECS, Di Stasi LC, Sassaki LY. Gut microbiota, inflammatory bowel disease and colorectal cancer. World J Gastroenterol. 2022;28(30):4053–60.
- Yang Y, Gharaibeh RZ, Newsome RC, Jobin C. Amending microbiota by targeting intestinal inflammation with TNF blockade attenuates development of colorectal cancer. Nat Cancer. 2020;1(7):723–.
- Hwang S, Lee CG, Jo M, Park CO, Gwon SY, Hwang S, Yi HC, Lee SY, Eom YB, Karim B, et al. Enterotoxigenic infection exacerbates tumorigenesis in AOM/DSS mouse model. Int J Med Sci. 2020;17(2):145–52.
- Huang F, Li S, Chen W, Han Y, Yao Y, Yang L, Li Q, Xiao Q, Wei J, Liu Z, et al. Postoperative probiotics administration attenuates gastrointestinal complications and gut microbiota dysbiosis caused by chemotherapy in colorectal cancer patients. Nutrients. 2023;15(2):356.
- Gopalakrishnan V, Spencer CN, Nezi L, Reuben A, Andrews MC, Karpinets TV, Prieto PA, Vicente D, Hoffman K, Wei SC, et al. Gut microbiome modulates response to anti-PD-1 immunotherapy in melanoma patients. Science. 2018;359(6371):97–103.
- Routy B, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillère R, Fluckiger A, Messaoudene M, Rauber C, Roberti MP, et al. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. Science. 2018;359(6371):91–.
- Lin C, Li B, Tu C, Chen X, Guo M. Correlations between Intestinal Microbiota and Clinical Characteristics in Colorectal Adenoma/Carcinoma. *Biomed Res Int* 2022, 2022;3140070.
- Yang X, Li P, Qu Z, Zhuang J, Wu Y, Wu W, Wei Q. Gut bacteria and sex differences in colorectal cancer. J Med Microbiol. 2023;72(6):1–13. https://doi.org/10.1099/jmm.0.001706.
- Liao H, Li C, Ai Y, Kou Y. Gut microbiome is more stable in males than in females during the development of colorectal cancer. J Appl Microbiol. 2021;131(1):435–48.
- 83. Watanabe D, Murakami H, Ohno H, Tanisawa K, Konishi K, Tsunematsu Y, Sato M, Miyoshi N, Wakabayashi K, Watanabe K, et al. Association

between dietary intake and the prevalence of tumourigenic bacteria in the gut microbiota of middle-aged Japanese adults. Sci Rep. 2020;10(1):15221.

- 84. Bellerba F, Serrano D, Johansson H, Pozzi C, Segata N, NabiNejad A, Piperni E, Gnagnarella P, Macis D, Aristarco V, et al. Colorectal cancer, Vitamin D and microbiota: a double-blind phase II randomized trial (ColoViD) in colorectal cancer patients. Neoplasia. 2022;34:100842.
- Song CH, Kim N, Nam RH, Choi SI, Jang JY, Kim EH, Choi J, Choi Y, Yoon H, Lee SM. The possible preventative role of lactate- and butyrateproducing bacteria in colorectal carcinogenesis. Gut Liver 2023. https:// doi.org/10.5009/gnl230385.
- Rezen T, Rozman D, Kovacs T, Kovacs P, Sipos A, Bai P, Miko E. The role of bile acids in carcinogenesis. Cell Mol Life Sci. 2022;79(5):243.
- Pearson T, Caporaso JG, Yellowhair M, Bokulich NA, Padi M, Roe DJ, Wertheim BC, Linhart M, Martinez JA, Bilagody C, et al. Effects of ursodeoxycholic acid on the gut microbiome and colorectal adenoma development. Cancer Med. 2019;8(2):617–28.
- Steegenga WT, Mischke M, Lute C, Boekschoten MV, Pruis MG, Lendvai A, Verkade HJ, Boekhorst J, Timmerman HM, Plösch T, et al. Sexually dimorphic characteristics of the small intestine and colon of prepubescent C57BL/6 mice. Biology sex Differences. 2014;5(11):11.
- Wang L, Tu YX, Chen L, Zhang Y, Pan XL, Yang SQ, Zhang SJ, Li SH, Yu KC, Song S, et al. Male-biased gut microbiome and metabolites aggravate colorectal Cancer Development. Adv Sci (Weinh). 2023;25:e2206238.
- Song CH, Kim N, Nam RH, Choi SI, Lee HN, Surh YJ. 17beta-Estradiol supplementation changes gut microbiota diversity in intact and colorectal cancer-induced ICR male mice. Sci Rep. 2020;10(1):12283.
- Hwang S, Jo M, Hong JE, Park CO, Lee CG, Rhee KJ. Protective effects of Zerumbone on colonic Tumorigenesis in Enterotoxigenic (ETBF)-Colonized AOM/DSS BALB/c Mice. Int J Mol Sci. 2020;21(3):857.
- Song CH, Kim N, Nam RH, Choi SI, Jang JY, Choi J, Lee HN. Anti-PD-L1 antibody and/or 17beta-Estradiol treatment induces changes in the gut microbiome in MC38 Colon tumor model. Cancer Res Treat. 2023;55(3):894–909.
- Ibrahim A, Hugerth LW, Hases L, Saxena A, Seifert M, Thomas Q, Gustafsson JA, Engstrand L, Williams C. Colitis-induced colorectal cancer and intestinal epithelial estrogen receptor beta impact gut microbiota diversity. Int J Cancer. 2019;144(12):3086–98.
- 94. Song CH, Kim N, Nam RH, Choi SI, Yu JE, Nho H, Surh YJ. Changes in Microbial Community Composition related to sex and Colon cancer by Nrf2 knockout. Front Cell Infect Microbiol. 2021;11:636808.
- Song CH, Kim N, Nam RH, Choi SI, Jang JY, Lee HN. Changes in gut Microbiome upon Orchiectomy and Testosterone Administration in AOM/DSS-Induced Colon Cancer Mouse Model. Cancer Res Treat. 2023;55(1):196–218.
- Wang L, Jiang G, Jing N, Liu X, Zhuang H, Zeng W, Liang W, Liu Z. Downregulating testosterone levels enhance immunotherapy efficiency. Oncoimmunology. 2021;10(1):1981570.
- 97. Jing N, Wang L, Zhuang H, Ai C, Jiang G, Liu Z. Sex-Biased Immune Responses to Antibiotics during Anti-PD-L1 Treatment in Mice with Colon Cancer. *J Immunol Res* 2022, 2022:9202491.
- Robinson I, Hochmair MJ, Schmidinger M, Absenger G, Pichler M, Nguyen VA, Richtig E, Rainer BM, Ay L, Jansen C, et al. Assessing the performance of a Novel Stool-based Microbiome Test that predicts response to First Line Immune checkpoint inhibitors in multiple Cancer types. Cancers (Basel). 2023;15(13):3268.
- Lin H, Ma X, Yang X, Chen Q, Wen Z, Yang M, Fu J, Yin T, Lu G, Qi J, et al. Natural shikonin and acetyl-shikonin improve intestinal microbial and protein composition to alleviate colitis-associated colorectal cancer. Int Immunopharmacol. 2022;111:109097.
- Leung HKM, Lo EKK, Chen C, Zhang F, Felicianna, Ismaiah MJ, El-Nezami H. Zearalenone attenuates colitis associated colorectal tumorigenesis through Ras/Raf/ERK pathway suppression and SCFA-producing bacteria promotion. Biomed Pharmacother. 2023;164:114973.
- Allsopp P, Possemiers S, Campbell D, Gill C, Rowland I. A comparison of the anticancer properties of isoxanthohumol and 8-prenylnaringenin using in vitro models of colon cancer. BioFactors. 2013;39(4):441–7.
- Wolfe AE, Moskowitz JE, Franklin CL, Wiemken TL, Ericsson AC. Interactions of segmented filamentous Bacteria (Candidatus Savagella) and bacterial drivers in colitis-associated colorectal cancer development. PLoS ONE. 2020;15(7):e0236595.

- Baker JM, Al-Nakkash L, Herbst-Kralovetz MM. Estrogen-gut microbiome axis: physiological and clinical implications. Maturitas. 2017;103:45–53.
- 104. Parida S, Sharma D. The Microbiome-Estrogen connection and breast Cancer risk. Cells. 2019;8(12):1642.
- Nuriel-Ohayon M, Neuman H, Ziv O, Belogolovski A, Barsheshet Y, Bloch N, Uzan A, Lahav R, Peretz A, Frishman S, et al. Progesterone increases Bifidobacterium relative abundance during late pregnancy. Cell Rep. 2019;27(3):730–736e733.
- 106. Matsushita M, Fujita K, Motooka D, Hatano K, Hata J, Nishimoto M, Banno E, Takezawa K, Fukuhara S, Kiuchi H, et al. Firmicutes in Gut Microbiota correlate with blood testosterone levels in Elderly men. World J Mens Health. 2022;40(3):517–25.
- Harada N, Hanaoka R, Hanada K, Izawa T, Inui H, Yamaji R. Hypogonadism alters cecal and fecal microbiota in male mice. Gut Microbes. 2016;7(6):533–9.
- Shin JH, Park YH, Sim M, Kim SA, Joung H, Shin DM. Serum level of sex steroid hormone is associated with diversity and profiles of human gut microbiome. Res Microbiol. 2019;170(4–5):192–201.
- 109. Shi N, Li N, Duan XW, Niu HT. Interaction between the gut microbiome and mucosal immune system. Military Med Res. 2017;4:14.
- Chakraborty B, Byemerwa J, Krebs T, Lim F, Chang CY, McDonnell DP. Estrogen Receptor Signaling in the Immune System. Endocr Rev. 2023;44(1):117–41.
- Kaliannan K, Robertson RC, Murphy K, Stanton C, Kang C, Wang B, Hao L, Bhan AK, Kang JX. Estrogen-mediated gut microbiome alterations influence sexual dimorphism in metabolic syndrome in mice. Microbiome. 2018;6(1):205.
- 112. Ma Y, Liu T, Li X, Kong A, Xiao R, Xie R, Gao J, Wang Z, Cai Y, Zou J, et al. Estrogen receptor beta deficiency impairs gut microbiota: a possible mechanism of IBD-induced anxiety-like behavior. Microbiome. 2022;10(1):160.
- 113. Fransen F, van Beek AA, Borghuis T, Meijer B, Hugenholtz F, van der Gaast-de Jongh C, Savelkoul HF, de Jonge MI, Faas MM, Boekschoten MV, et al. The impact of gut microbiota on gender-specific differences in immunity. Front Immunol. 2017;8:754.
- 114. Buendia-Gonzalez FO, Legorreta-Herrera M. The similarities and differences between the effects of Testosterone and DHEA on the Innate and Adaptive Immune Response. Biomolecules. 2022;12(12):1768.
- 115. Gomez A, Luckey D, Taneja V. The gut microbiome in autoimmunity: sex matters. Clin Immunol. 2015;159(2):154–62.
- Milani N, Qiu N, Fowler S. Contribution of UGT enzymes to Human Drug Metabolism Stereoselectivity: a case study of Medetomidine, RO5263397, Propranolol, and Testosterone. Drug Metab Dispos. 2023;51(3):306–17.
- 117. Collden H, Landin A, Wallenius V, Elebring E, Fandriks L, Nilsson ME, Ryberg H, Poutanen M, Sjogren K, Vandenput L, et al. The gut microbiota is a major regulator of androgen metabolism in intestinal contents. Am J Physiol Endocrinol Metab. 2019;317(6):E1182–92.
- Ervin SM, Li H, Lim L, Roberts LR, Liang X, Mani S, Redinbo MR. Gut microbial beta-glucuronidases reactivate estrogens as components of the estrobolome that reactivate estrogens. J Biol Chem. 2019;294(49):18586–99.
- Sui Y, Wu J, Chen J. The role of Gut Microbial beta-glucuronidase in estrogen reactivation and breast Cancer. Front Cell Dev Biol. 2021;9:631552.
- 120. Ly LK, Doden HL, Ridlon JM. Gut feelings about bacterial steroid-17,20-desmolase. Mol Cell Endocrinol. 2021;525:111174.
- 121. Vom Steeg LG, Klein SL. Sex steroids mediate bidirectional interactions between hosts and microbes. Horm Behav. 2017;88:45–51.
- 122. Tremellen K. Gut endotoxin leading to a decline IN gonadal function (GELDING) a novel theory for the development of late onset hypogonadism in obese men. Basic Clin Androl. 2016;26:7.
- Li X, Cheng W, Shang H, Wei H, Deng C. The interplay between Androgen and Gut Microbiota: is there a Microbiota-Gut-Testis Axis. Reprod Sci. 2022;29(6):1674–84.
- 124. Snigdha S, Ha K, Tsai P, Dinan TG, Bartos JD, Shahid M. Probiotics: potential novel therapeutics for microbiota-gut-brain axis dysfunction across gender and lifespan. Pharmacol Ther. 2022;231:107978.

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