# REVIEW

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# Harnessing peroxisome proliferator-activated receptor γ agonists to induce Heme Oxygenase-1: a promising approach for pulmonary inflammatory disorders

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## Abstract

The activation of peroxisome proliferator-activated receptor (PPAR)-v has been extensively shown to attenuate inflammatory responses in conditions such as asthma, acute lung injury, and acute respiratory distress syndrome, as demonstrated in animal studies. However, the precise molecular mechanisms underlying these inhibitory effects remain largely unknown. The upregulation of heme oxygenase-1 (HO-1) has been shown to confer protective effects, including antioxidant, antiapoptotic, and immunomodulatory effects in vitro and in vivo. PPARy is highly expressed not only in adipose tissues but also in various other tissues, including the pulmonary system. Thiazolidinediones (TZDs) are highly selective agonists for PPARy and are used as antihyperglycemic medications. These observations suggest that PPARy agonists could modulate metabolism and inflammation. Several studies have indicated that PPARy agonists may serve as potential therapeutic candidates in inflammation-related diseases by upregulating HO-1, which in turn modulates inflammatory responses. In the respiratory system, exposure to external insults triggers the expression of inflammatory molecules, such as cytokines, chemokines, adhesion molecules, matrix metalloproteinases, and reactive oxygen species, leading to the development of pulmonary inflammatory diseases. Previous studies have demonstrated that the upregulation of HO-1 protects tissues and cells from external insults, indicating that the induction of HO-1 by PPARy agonists could exert protective effects by inhibiting inflammatory signaling pathways and attenuating the development of pulmonary inflammatory diseases. However, the mechanisms underlying TZD-induced HO-1 expression are not well understood. This review aimed to elucidate the molecular mechanisms through which PPARy agonists induce the expression of HO-1 and explore how they protect against inflammatory and oxidative responses.

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### Introduction

Chronic inflammation is at the core of airway and pulmonary inflammatory diseases, including asthma, chronic obstructive pulmonary disease (COPD), acute lung injury, and acute respiratory distress syndrome (ARDS) [1]. Various proinflammatory mediators, such as cigarette smoke extract (CSE), adenosine-5'-triphosphate (ATP), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin-1 $\beta$ (IL-1 $\beta$ ), lipopolysaccharide (LPS), and lipoteichoic acid (LTA), play pivotal roles in initiating and sustaining



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inflammation in the pulmonary system [2-10]. These inflammatory responses are primarily triggered in two key types of lung cells. Airway smooth muscle cells, which are typically involved in ventilation regulation, become responsive to various external substances and proinflammatory mediators under pathological conditions such as asthma. Furthermore, alveolar epithelial type II cells, which are vital for gas exchange, are highly susceptible to oxidants. In response to exposure to proinflammatory mediators and oxidative stress, these lung cells release a cascade of cytokines, chemokines, and inflammatory mediators, including IL-1 $\beta$ , TNF- $\alpha$ , cyclooxygenase-2 (COX-2), cytosolic phospholipase  $A_2$ (cPLA<sub>2</sub>), and matrix metalloproteinase-9 (MMP-9) [11]. Previous studies have revealed the induction of proteins such as COX-2/prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) [2, 6, 12, 13], cPLA<sub>2</sub> [8, 14, 15], MMP-9 [3, 16, 17], intercellular adhesion molecule-1 (ICAM-1) [7, 18, 19], and vascular cell adhesion molecule-1 (VCAM-1) [5, 19-21] and reactive oxygen species (ROS) [7, 21, 22] in response to various stimuli and pathogens associated with pulmonary inflammation (Fig. 1). The induction of these factors is likely mediated by NADPH oxidase (NOX)/ROS, intracellular signaling pathways, and transcription factors, illustrating the complex interplay between ROS and the expression of inflammatory proteins induced by proinflammatory mediators (Fig. 1). Consequently, understanding these mechanisms is crucial for developing therapeutic strategies for pulmonary inflammatory diseases. Oxidative stress plays a pivotal role in pulmonary inflammation. Early interventions targeting oxidative stress could delay the onset and progression of inflammation. Despite extensive research, an effective therapeutic strategy to prevent pulmonary inflammation progression remains elusive. Therefore, the development of antioxidative and anti-inflammatory drugs has emerged as a promising avenue to inhibit the pathological progression of these diseases.

Peroxisome proliferator-activated receptor (PPAR) agonists, which are widely used in clinical settings, exert multiple effects, including anti-inflammatory and antioxidative effects, on various tissues and organs. PPARy is abundantly expressed in human adipose tissue, and its expression levels are comparatively lower in bone marrow, skeletal muscle, liver, heart, and stromal cells [23, 24]. PPARy plays a crucial role in glucose metabolism, adipocyte differentiation, and the inflammatory response and is important in various physiological processes. Its therapeutic applications extend to the treatment of conditions such as diabetes, fatty liver disease, cardiovascular disease, and neuroinflammation [25–30]. PPAR agonists, while exhibiting increased therapeutic efficacy and improved glycemic control, are part of a complex clinical landscape due to a spectrum of associated adverse reactions. Notably, these adverse events include an increased risk of cardiovascular events, including congestive heart failure, cardiovascular death, and myocardial infarction [31–33]. Additionally, users may face increased risk of



**Fig. 1** Pathways in pulmonary inflammation and potential therapeutic interventions. During pulmonary inflammation, proinflammatory factors like CSE, ATP, peptides (e.g., bradykinin, BK and *endothelin-1, ET-1*), cytokines (TNF-α and IL-1β), and endotoxins (e.g., LTA and LPS) increase. These factors induce inflammatory mediators (e.g., MMPs, adhesion molecules, COX-2, or cPLA<sub>2</sub>) through various signaling molecules, including mitochondrial or NOX-derived ROS generation, MAPKs activation, transactivation of growth factor receptors, and transcription factors in pulmonary resident cells (alveolar epithelial cells and tracheal smooth muscle cells). These changes lead to pathological alterations in these cells. Furthermore, potential therapeutic drugs such as PPAR agonists might protect against pulmonary inflammation by inducing antioxidant proteins like HO-1. It is hypothesized that these drugs induce HO-1 expression through ROS-dependent signals, directly preventing lung injury and inflammation

fractures, bladder cancer, and macular edema and weight gain [31–33]. It is imperative to approach the clinical use of PPAR agonists with a nuanced understanding of the potential hepatotoxicity associated with their use [31– 33]. This consideration of therapeutic benefits and associated risks underscores the importance of careful patient selection and monitoring when using PPAR agonists in various medical contexts. By leveraging these pleiotropic effects, the use of PPAR agonists is a potential strategy for managing pulmonary inflammatory diseases (Fig. 1). Further research and clinical exploration in this direction may lead to innovative treatments for these challenging conditions.

### The anti-inflammatory and antioxidative effects of PPAR agonists in diverse inflammatory disorders

Recently, research has identified three distinct PPAR isoforms (PPAR $\alpha$ , PPAR $\beta$ , and PPAR $\gamma$ ), which are encoded by unique genes. Notably, two PPARy isoforms are generated through alternative splicing of exons A1, A2, and B1, giving rise to  $\gamma$ 1,  $\gamma$ 2, and  $\gamma$ 3 variants, respectively [34]. These PPARs are widely expressed in various animal and human tissues. In adult rodents, PPARa is prominently found in tissues with high peroxisome-dependent activities or rapid fatty acid catabolism, such as hepatocytes, cardiomyocytes, proximal tubules of the kidney cortex, the intestinal mucosa, and brown adipose tissue [35, 36]. In humans, PPAR $\alpha$  is notably expressed in skeletal muscle, heart, liver, kidney, intestine, and pancreas. Additionally, it is present, albeit at lower levels, in adipose tissue, lung, and placenta [23]. PPARa's primary functions include the regulation of fatty acid oxidation and lipoprotein metabolism and involvement in anti-inflammatory/antioxidant pathways. PPAR $\beta$ , on the other hand, is ubiquitously expressed in most rodent tissues and in higher quantities than PPARa and PPARy [37]. This widespread presence extends to human tissues, making PPAR $\beta$  a key player in fatty acid metabolism regulation and the suppression of macrophage-mediated inflammation. PPARy, which is prominently expressed in adipose tissue and moderately in the large intestine and select immune system components such as the spleen in rodents, is abundantly found in human adipose tissue. However, in humans, it is expressed at lower levels in bone marrow, skeletal muscle, liver, heart, and stromal cells [23, 24]. PPARy plays a pivotal role in glucose metabolism, adipocyte differentiation, and inflammatory responses.

Natural ligands of PPARs include fatty acids and eicosanoids. Leukotriene B4 activates PPAR $\alpha$ , while prostaglandin J<sub>2</sub> (PGJ<sub>2</sub>) serves as a ligand for PPAR $\gamma$ . Additionally, synthetic ligands such as antidiabetic glitazones and hypolipidemic fibrates activate PPAR $\gamma$  and PPAR $\alpha$ , respectively. Research has shown that PPARa regulates the oxidation of fatty acids in the liver by controlling genes containing peroxisome proliferator response elements (PPREs) in their promoter regions [38]. Mice lacking PPARa show prolonged responses to inflammatory mediators. Furthermore, PPAR agonists inhibit the activation of genes associated with the inflammatory response, such as IL-2, IL-6, IL-8, TNF- $\alpha$ , and metalloproteases, by modulating signaling pathways involving nuclear factor κ-light-chain-enhancer of activated B cells  $(NF-\kappa B)$ , activator protein 1 (AP-1), and signal transducer and activator of transcription proteins (STATs) [39]. The activation of PPARy also mediates adipogenesis; studies have demonstrated that synthetic PPARy ligands such as rosiglitazone [40] and natural ligands such as 15d-PGJ<sub>2</sub> [41] reduce serum levels and transcription of TNF- $\alpha$ .

In summary, PPARs play multiple roles in the  $\beta$ -oxidation of fatty acids and the metabolism of arachidonic acid metabolites. Beyond these functions, they exhibit significant anti-inflammatory potential. These findings strongly suggest that PPARs are key regulators that control inflammation, paving the way for potential therapeutic interventions in inflammatory diseases.

# Elucidating the mechanisms by which PPARy agonists induce heme oxygenase-1 (HO-1) expression

PPARs hold significant promise as therapeutic agents for treating lung inflammation [42, 43]. In particular, PPARy ligands can inhibit the release of proinflammatory cytokines from airway epithelial cells and reduce airway hyperresponsiveness in murine models of asthma [44]. A noteworthy example is rosiglitazone, a PPARy agonist that is known not only for its role in maintaining glucose and lipid homeostasis but also for its capacity to attenuate airway inflammation through the upregulation of HO-1 [45]. HO-1, which is a direct transcriptional target of PPARy, exhibits potent anti-inflammatory, antioxidant, and apoptosis-regulating properties [46, 47]. Furthermore, the upregulation of HO-1 by PPARy agonists has been linked to the inhibition of pulmonary cell proliferation and remodeling [48]. However, the precise mechanisms governing rosiglitazone-induced PPARy/HO-1 expression in human pulmonary alveolar epithelial cells (HPAEpiCs) remain unclear. In the following sections, we will delve into the molecular mechanisms through which PPARy agonists induce HO-1 expression. We will examine the activation of various signaling components and transcription factors that contribute to this regulatory process. This understanding will not only shed light on the potential therapeutic applications of PPARy agonists in managing pulmonary inflammation but also unveil

the underlying molecular mechanisms that govern these therapeutic effects.

#### NOX and HO-1 expression

ROS are pivotal messengers during normal physiological functions, and their effects oscillate between beneficial and inflammatory, depending on their concentrations [49]. HO-1, which is a sentinel enzyme, is activated in response to intracellular oxidative stress or disruptions in intracellular reduction-oxidation (redox) equilibrium, serving as a protector against inflammatory responses [50]. Notably, activated NOX is a prominent source of ROS, which act as secondary messengers that induce the expression of HO-1, a potent defense mechanism against inflammation [51-53]. Many stimuli can induce NOX/ ROS-dependent HO-1 expression, underscoring the complexity of this regulatory pathway [54, 55]. Intriguingly, PPAR agonists activate protein kinases through ROS-dependent pathways, thereby regulating a multitude of genes, including HO-1 [56, 57]. Furthermore, endogenous PPARy ligands such as 15d-PGJ<sub>2</sub> have been shown to induce ROS generation, stimulating the expression of HO-1 via the nuclear factor E2-related factor 2 (Nrf2) pathway [58]. PPARy ligands, with their multifaceted effects, have been shown to inhibit airway inflammation and hyperresponsiveness through PPARy-independent mechanisms in mouse models challenged with allergens [59]. The NOX/ROS system, which is a complex web of signaling components, induces HO-1 expression through a choreographed activation process. In response to stimulation, cytosolic regulatory subunits such as p40<sup>phox</sup>, p47<sup>phox</sup>, p67<sup>phox</sup>, Ras-related C3 botulinum toxin substrate (Rac)1, and Rac2 translocate to the membrane. There, they assemble with membrane-bound components, including gp91<sup>phox</sup>/p22<sup>phox</sup>, leading to the generation of ROS  $(O_2^{\bullet-}/H_2O_2)$  [60]. Specifically, NOX2, which is a prominent NOX enzyme in pulmonary alveolar epithelial cells, recruits p22<sup>phox</sup> and p47<sup>phox</sup>, thereby producing  $O_2^{\bullet-}$  [60]. Phosphorylation of p47<sup>phox</sup> is a pivotal event that is essential for NOX activation and ROS generation [61]. In our studies of HPAEpiCs, we discovered a significant connection. Rosiglitazone, which is a PPARy agonist, upregulates HO-1 through NOX-dependent ROS generation [62]. This relationship was underscored by the attenuation of rosiglitazone-induced HO-1 expression when p47<sup>phox</sup> or NOX was inhibited, as well as in the presence of ROS scavengers. Moreover, NOX inhibitors exerted their effects by preventing p47<sup>phox</sup> phosphorylation or membrane translocation, effectively inhibiting NOX activation and ROS generation [63]. These findings emphasize the vital role of NOX/ROS signaling in the responses induced by rosiglitazone, further illuminating the underlying mechanisms by which PPARy agonists protect against pulmonary inflammation. However, the precise mechanisms by which rosiglitazone triggers the phosphorylation of p47<sup>phox</sup> and the recruitment of NOX subunits to the plasma membrane, ultimately leading to ROS generation, remain unknown. Further investigations will unveil critical insights, paving the way for a deeper understanding of these complex molecular interactions.

### Protein tyrosine kinases and HO-1 expression

Protein tyrosine kinases are pivotal players in intracellular signaling and are categorized as receptor tyrosine kinases (e.g., EGFR and PDGFR) and nonreceptor tyrosine kinases (e.g., c-Src and Pyk2). These kinases modulate diverse cellular functions. Notably, NOX/ROS production activates downstream components such as c-Src and Pyk2, influencing cellular activities [64, 65]. Specifically, c-Src acts as an upstream regulator, phosphorylating Pyk2, which is stimulated by thiazolidinediones (TZDs) and regulated by ROS. This phosphorylation event triggers HO-1 expression, protecting various cell types from oxidative damage [65-69]. Our investigations of HPAEpiCs showed that rosiglitazone-induced HO-1 expression involved the phosphorylation of c-Src and Pyk2. The nonreceptor tyrosine kinases c-Src, and Pyk2 are pivotal players in the protective effects of rosiglitazone against pulmonary inflammation [70]. ROSdependent phosphorylation of c-Src and Pyk2 is critical for HO-1 induction by rosiglitazone, offering a promising avenue for therapeutic interventions against pulmonary inflammation. Notably, c-Src and Pyk2 are distinct from receptor tyrosine kinases such as EGFR and PDGFR. Hyperoxia-induced phosphorylation of EGFR has been shown to induce HO-1 expression in pulmonary epithelial cells [71]. This intriguing interplay between PPARy agonists and receptor tyrosine kinases such as EGFR and PDGFR presents a compelling area for future exploration. Understanding whether rosiglitazone stimulates the phosphorylation of EGFR or PDGFR, thereby triggering HO-1 expression in pulmonary resident cells, holds the key to unraveling another layer of the complex molecular landscape underpinning the protective effects of PPARy agonists against pulmonary inflammation. In essence, the interplay between nonreceptor tyrosine kinases, receptor tyrosine kinases, and PPARy agonists in the regulation of HO-1 expression opens up exciting avenues for further research. These findings not only deepen our understanding of the molecular mechanisms but also offer potential targets for therapeutic strategies to mitigate pulmonary inflammation.

### Protein kinase Cs (PKCs) and HO-1 expression

The PKC family is a diverse group of at least eleven isoforms that are categorized into three groups and are finely tuned to cellular signals for precise regulation [72]. Notably, classical PKCs such as PKC $\alpha$ ,  $\beta/\delta$ , and  $\gamma$  are activated by diacylglycerol (DAG) in a calcium-dependent manner, serving as pivotal players in various pathophysiological responses in different cellular contexts [73]. These serine- and threonine-specific protein kinases are fundamental to multiple physiological processes and gene expression in various cellular contexts [74]. Their activation results in the phosphorylation of specific target proteins, which is involved in various pathological conditions, including pulmonary inflammatory diseases [75]. Furthermore, PKC isoforms modulate both pro- and anti-inflammatory systems, underscoring their vital roles in cellular functions and gene regulation [76]. Numerous studies have demonstrated the regulatory role of PKCs, especially PKC $\alpha$ , in orchestrating the expression of HO-1 in response to diverse stimuli in many cell types [77, 78]. Specifically, PKCa actively participates in immune signaling pathways, influencing the expression of inflammatory genes [79, 80]. By enhancing the phosphorylation of several protein kinases, PKCa facilitates the expression of specific proteins, which has been extensively observed in various tissues and cells [79]. Furthermore, the PKC $\alpha$ / Erk1/2 pathway has been identified as a key regulator of HO-1 expression induced by compounds such as curcumin [81]. Our research substantiates these findings. Using small interfering RNA (siRNA) to silence PKCa significantly decreases rosiglitazone-induced HO-1 expression in HPAEpiCs through PKCα phosphorylation [70]. Moreover, PKC $\alpha$  plays a crucial role in the Nrf2/ HO-1 pathway by protecting against oxidative stress [82]. Collectively, our results underscore the pivotal role of phosphorylated PKCs, particularly PKCα, in mediating rosiglitazone-induced HO-1 expression, offering promising avenues for therapeutic interventions against pulmonary inflammatory diseases. However, unraveling the effects of other PKC isoforms on HO-1 expression induced by TZDs is a critical area that requires further investigation.

### AMPKa and HO-1 expression

5'-Adenosine monophosphate-activated protein kinase α (AMPKα) is a crucial cellular sensor that is activated in response to reduced ATP levels and plays a pivotal role in cellular homeostasis. This enzyme not only enhances cyclic adenosine monophosphate (cAMP) production, protecting endothelial cells against proliferation and angiogenesis [83], but also orchestrates a myriad of cellular processes through the phosphorylation of target proteins and the regulation of gene expression. Notably, AMPKα is a potent inducer of Nrf2/HO-1 gene expression, effectively inhibiting inflammatory responses in animal models [84]. The multifaceted role of AMPKα is

immune regulation is underscored by its ability to induce the production of IL-10, negatively regulate ΙκΒα degradation, inhibit glycogen synthase kinase (GSK) 3β, and activate Akt and cAMP response element-binding protein (CREB). These actions culminate in the promotion of an anti-inflammatory phenotype, indicating that AMPKα is a key player in immune modulation [85]. Moreover, AMPK $\alpha$ 's influence extends to airway inflammation, where it uses multiple pathways to suppress the release of proinflammatory cytokines and inhibit the NF-KB pathway, thereby mitigating pulmonary inflammation and emphysema in lung disease models [86, 87]. The protective and anti-inflammatory effects of AMPK $\alpha$  are often mediated by the induction of HO-1 expression in lung epithelial cells, which serves as a linchpin in the cellular defense against inflammatory insult [88, 89]. In the specific context of our study, we discovered that rosiglitazone activated AMPKα, leading to the induction of HO-1 expression and subsequent anti-inflammatory effects on HPAEpiCs [70]. This finding highlights the pivotal role of AMPKa phosphorylation in mediating the protective effects of rosiglitazone against pulmonary inflammatory diseases. Furthermore, the intriguing prospect that AMPKa influences downstream components, particularly mitogen-activated protein kinases (MAPKs), in the context of HO-1 expression opens a promising avenue for future research. Understanding the interplay between AMPK $\alpha$  and MAPKs could lead to the discovery of novel therapeutic strategies, potentially revolutionizing our approach to combating pulmonary inflammatory disorders [90, 91]. This network of signaling pathways presents an exciting frontier for exploration, promising not only deeper insights into cellular responses but also innovative therapeutic interventions for inflammatory diseases.

### MAPKs and HO-1 expression

MAPKs form a pivotal signaling network that regulates many cellular functions, including proliferation, apoptosis, motility, differentiation, immunity, and responses to oxidative stress. MAPKs include three distinct groups in mammalian cells: extracellular signal-regulated protein kinases (ERKs: ERK1/2), c-Jun NH2-terminal kinases (JNKs: JNK1, JNK2, and JNK3), and p38 MAP kinases (p38 $\alpha$ , p38 $\beta$ , p38 $\gamma$ , and p38 $\delta$ ). In the context of HO-1 regulation, MAPK pathways are conduits through which various stimuli induce HO-1 expression via downstream protein kinases and transcription factors [92-95]. Interestingly, HO-1 induction is linked to the activation of MAPKs mediated by protein phosphorylation and redox reactions. Sodium arsenite triggers HO-1 expression by activating JNK1/2 in rat hepatocytes, while in chicken hepatoma cells, arsenite uses both the ERK1/2 and p38 MAPK pathways to induce HO-1 expression [96, 97].

The ischemia-reperfusion lung model implicates JNK1/2 and p38 MAPK in HO-1 expression [98], and in various cell types, p38 MAPK activation upregulates HO-1, as observed in porcine renal epithelial cells and human hepatoma cells stimulated by curcumin [93, 99]. Furthermore, 15d-PGJ<sub>2</sub>, which is an endogenous PPARy agonist, induces HO-1 expression through p38 MAPKa in rat vascular smooth muscle cells [100]. Consistent with these findings, our research uncovered the involvement of p38 MAPKa phosphorylation in rosiglitazone-induced HO-1 expression in HPAEpiCs [70]. These observations underscore the significant role of MAPKs, including p38 MAPKa, in TZD-induced HO-1 expression. However, the distinct roles of ERK1/2 and JNK1/2 in rosiglitazoneinduced HO-1 expression in HPAEpiCs need further exploration. This interplay between different MAPK pathways and HO-1 induction is critical for future investigations, offering potential insights into cell-specific responses and paving the way for novel therapeutic interventions in inflammatory diseases.

# Phosphoinositide 3-kinase/Akt (PI3K/Akt) and HO-1 expression

The PI3K/Akt pathway, which is a pivotal antiapoptotic survival pathway, is regulated by various receptordependent mechanisms that are activated by growth factors and cytokines [101]. Notably, in models featuring PI3K genetic deficiency, this kinase has been implicated in the regulation of inflammatory reactions, emphasizing its importance in cellular responses [102]. A growing body of evidence underscores the multifaceted role of PI3K/Akt in HO-1 regulation. Activation of PI3K/ Akt not only upregulates HO-1 gene expression but also connects the protective effects of this signaling cascade to the beneficial effects of HO-1 [103]. Immune cells exhibit upregulated HO-1 gene expression mediated by PI3K/Akt activation in response to diverse stimuli [103–105]. Intriguingly, HO-1 induction has been linked to the activation of PI3K/Akt through a mitochondrial redox-dependent pathway in vascular endothelial cells, highlighting the nuanced regulatory mechanisms involved in HO-1 expression [106]. Furthermore, NOX/ ROS-dependent HO-1 expression has been shown to be mediated by PI3K/Akt activation, resulting in protection against oxidative stresses [107, 108]. Activated Akt, in turn, triggers the expression of Nrf2 and subsequently HO-1, increasing cellular defense mechanisms and promoting cell survival in response to stresses [107, 108]. Consistent with these findings, our research showed the critical role of Akt phosphorylation induced by rosiglitazone in facilitating HO-1 expression, and this effect was significantly attenuated by LY294002 treatment or transfection with Akt siRNA in HPAEpiCs [62]. This discovery underscores the pivotal role of PI3K/Akt in TZD-induced HO-1 expression in many cell types. However, the regulatory mechanisms of PI3K/Akt-mediated HO-1 induction in response to PPARy agonists remain unclear. Delving deeper into these mechanisms will not only enhance our understanding of the complexities of cellular signaling pathways but may also lead to novel therapeutic strategies to protect against inflammatory diseases.

### Janus kinase (JAK)/signal transducer and activator of transcription 3 (STAT3) and HO-1 expression

The intricacies of the JAK/STAT3 signaling cascade, which links HO-1 gene expression to cytokines and oxidative stress, remain unclear. However, the involvement of STAT3 activation in this regulatory pathway has been suggested based on existing evidence [109]. The JAK/STAT signaling pathway plays a pivotal role in the immune system, serving as a major mediator of cytokineactivated pathways. In endothelial cells, STAT3 has been shown to mediate HO-1-dependent protection against hyperoxic lung injury, underscoring its importance in cellular defense mechanisms [110]. Functional STAT3 elements have been identified in the promoter regions of both rat [111] and human [112] HO-1 genes, indicating a direct regulatory role of STAT3 in HO-1 expression. In rat hepatocytes, IL-6-induced HO-1 gene expression has been shown to be mediated by the JAK/STAT3 pathway, highlighting the interplay between this signaling cascade and HO-1 induction [111]. Notably, the specific interactions between JAK/STAT signaling and HO-1 expression in HPAEpiCs remain unexplored, representing a significant gap in our understanding. Therefore, the potential regulatory role of JAK/STAT in HO-1 gene expression induced by PPARy agonists, particularly in pulmonary resident cells such as HPAEpiCs, holds substantial promise for further investigation. Exploring this aspect will not only broaden our understanding of the molecular mechanisms of HO-1 regulation but also open avenues for targeted therapeutic interventions against inflammatory diseases.

# Transcriptional regulation of HO-1 gene expression: key transcription factors and their functions

Understanding the machinery of HO-1 expression is crucial for targeted anti-inflammatory treatments. HO-1, which is a versatile enzyme, is triggered by various stimuli, such as oxidative stress, cytokines, bacterial compounds, and growth factors. Its expression is primarily governed at the transcriptional level, and multiple cisacting regulatory elements within the HO-1 promoter control its basal and inducible expression in different species [104, 113, 114]. Within the promoter regions of HO-1, two upstream enhancer regions, namely, E1 and E2, play pivotal roles in redox-dependent induction [113]. These enhancer regions house several antioxidant response elements (AREs) [115], which are also found in other stress-inducible antioxidant and phase 2 detoxifying gene promoters [116, 117]. Notably, a GT-microsatellite polymorphism in the proximal human HO-1 gene promoter distinguishes it from its rodent counterpart. This polymorphism, which is characterized by varying numbers of GT repeats, significantly impacts HO-1 inducibility; fewer GT repeats correlate with increased HO-1 expression in response to stressors. Individuals harboring this allele are protected against cardiovascular disorders [118, 119]. In this comprehensive review, we focus on the pivotal roles of major transcription factors such as Nrf2, silent information regulator type-1 (SIRT1)/peroxisome proliferator-activated receptor gamma coactivator  $1-\alpha$  (PGC1 $\alpha$ ), PPAR $\gamma$ , Sp1, and AP-1 in the regulation of HO-1 gene expression, specifically in response to PPARy agonists. Delving into these regulatory pathways will not only enhance our understanding of HO-1 modulation but also pave the way for refining anti-inflammatory therapeutic strategies.

# Decoding HO-1 expression: unraveling coordinated activation via the Nrf2 signaling pathway

The regulation of HO-1, a critical player in oxidative stress management, is governed by a complex network involving multiple factors and pathways. At the heart of this system is Nrf2, which is essential for the transcriptional regulation of the *hmox1* gene that encodes HO-1 [50, 51]. Normally, Nrf2 is restrained in the cytoplasm by its interaction with Kelch-like ECH-associated protein (Keap1). However, when cells encounter oxidative stress, marked by an increase in ROS, this interaction is disrupted [51]. As a result, Nrf2 is released and accumulates in the nucleus, where it forms dimers with small Maf proteins and binds to AREs in the promoters of genes like hmox1, driving the transcription of genes involved in antioxidant defense [120-122]. This response to oxidative stress serves to bolster the cell's antioxidant capacity, with HO-1 being a key component. Alongside Nrf2, the heme-binding protein BTB and CNC homologue 1 (Bach1), which negatively regulates hmox1 transcription, is also involved [123]. Bach1 competes with Nrf2 for binding sites on DNA. Interestingly, heme influences this interaction; it not only promotes the release of Nrf2 from Keap1 but also aids in the export of Bach1 from the nucleus, thereby facilitating Nrf2's activity [124].

The role of the PPARy pathway in regulating HO-1 is another crucial aspect. PPARy agonists, such as rosiglitazone, have been shown to upregulate HO-1 expression. This not only aids in maintaining redox balance but also exhibits anti-inflammatory effects, suggesting a therapeutic potential in conditions characterized by oxidative stress. Our research has highlighted that rosiglitazone can induce HO-1 expression in HPAEpiCs through mechanisms that are both dependent on and independent of PPARy. This includes the activation of PPARy leading to increased HO-1 expression, which in turn helps reduce inflammation and tissue remodeling in the airways [45]. Interestingly, this process seems to be unaffected by factors such as NOX/ROS, c-Src/Pyk2, Akt, and even Nrf2 itself [70]. Further, we found that rosiglitazone's stimulation of HO-1 expression involves the activation of Nrf2 via a pathway that does not rely on PPARy. This was evident from the increased binding of Nrf2 to the ARE in the HO-1 promoter, a process that was sensitive to Nrf2 siRNA and pharmacological inhibitors but not to PPARy antagonists. Additionally, a decrease in Keap1 levels corresponded with an increase in HO-1 protein in HPAEpiCs, highlighting the significant role of Nrf2 in mediating the response to rosiglitazone.

This intricate interplay between various factors emphasizes the delicate balance between prooxidant forces and antioxidant defenses in the cell. The upregulation of HO-1 is a key protective mechanism against oxidative stress, but the effectiveness of this response is dependent on the specific conditions of stress, including its intensity and duration. This underscores the need for further research to explore the dual PPAR $\gamma$ -dependent and independent mechanisms involved in rosiglitazone-induced HO-1 expression and to understand their wider implications in the management of oxidative stress.

### Regulation of HO-1 expression by SIRT1/PGC1a

SIRT1, which is a member of the NAD-dependent deacetylases known as sirtuins, plays a pivotal role in anti-inflammatory responses by suppressing the production of various proinflammatory cytokines [125, 126]. Recent research highlights the protective effects of SIRT1-mediated HO-1 induction, which not only suppresses inflammatory responses but also activates antiapoptotic pathways [127]. The activation of SIRT1, which is often facilitated by compounds such as resveratrol, effectively inhibits bronchial inflammation induced by cigarette smoke in the lungs [128, 129]. Additionally, SIRT1 prevents multiple inflammatory responses by enhancing PGC1a activation and downregulating NF-κB [130]. Overexpression of SIRT1 reduces the acetylation levels of PGC1a, thereby protecting against neuroinflammation [131]. Nuclear receptors interact with coactivators such as PGC1a and corepressors such as NCoR. PGC1a overexpression has been shown to protect endothelial cells against oxidative stress [132]. In contrast, NCoR not only prevents inflammatory target gene expression in the absence of stimuli but is also essential for PPAR ligand-dependent transcription [133]. SIRT1's interaction with NCoR could negatively regulate PPARy when PGC1 $\alpha$  levels are low [134]. Our observations align with these findings. Rosiglitazone induces the phosphorylation of SIRT1 specifically in the cytosol, facilitating the deacetylation of Ac-PGC1 $\alpha$ . This process promotes the translocation of PGC1 $\alpha$  from the cytosol to the nucleus, where it interacts with PPARy [70]. Furthermore, there is a reduction in NCoR levels in the nucleus, enhancing the phosphorylation of PPARy. This interplay between the transcriptional corepressor NCoR and the coactivator PGC1 $\alpha$  underscores their involvement in rosiglitazone-induced HO-1 expression through a PPARy-dependent pathway in HPAEpiCs [70].

# Regulation of HO-1 expression by PPARy: insights and mechanisms

PPARy belongs to the nuclear hormone receptor family and functions as a transcriptional activator, relying on ligand binding to activate its domain. All PPAR members share similar structural domains: a ligand-dependent activation domain, a DNA-binding domain, and an amino-terminal region enabling ligand-independent activation [34]. PPAR-mediated transactivation occurs when PPAR and 9-cis retinoic acid receptor (RXR) bind to PPAR-response elements (PPREs) in response to ligand activation. Ligand binding induces conformational changes, facilitating coactivator recruitment and releasing corepressors such as NCoR. Ligand binding or other activation processes, including phosphorylation, induce conformational shifts in PPAR, particularly in the ligand-binding domain (LBD) and the C-terminal helix activation function-2 (AF-2). This alteration generates new protein-protein interaction surfaces, allowing the recruitment of specific coactivators such as steroid receptor coactivator 1 (SRC-1), CREB-binding protein (CBP)/p300, and PGC-1. Subsequently, the regulatory effects of this complex are transduced to the basal transcriptional machinery [34]. Phosphorylation at the Ser<sup>112</sup> residue of PPARy has been shown to influence its binding affinity with PPARy ligands and its interactions with coactivators and posttranslational events [135, 136]. Notably, rosiglitazone protects against acute lung injury and inflammation through HO-1 expression [137, 138]. Consistent with these findings, our study reveals that rosiglitazone-induced HO-1 expression occurs through a PPARy-dependent mechanism in HPAEpiCs [62]. However, the interactions among various nuclear components that initiate HO-1 gene expression are highly complex and require further clarification.

Regulation of HO-1 expression by the transcription factor Sp1 Sp1, which is a versatile transcription factor, responds to many signals, including oxidative stress [139, 140]. This factor affects vital physiological processes, controlling cell cycle dynamics, growth modulation, hormonal responses, apoptosis, and angiogenesis [141]. Beyond self-regulation, Sp1 governs genes that are essential for cellular homeostasis, forging direct connections with critical elements such as TATA-binding protein-associated factors [142], cAMP response domains [143], NF-κB [144], and vascular endothelial growth factor receptor-2 [145]. This regulatory role is modulated by dynamic modifications, such as phosphorylation, acetylation, and methylation, which fine-tune Sp1 protein levels, transactivation potential, and DNA binding affinity [139, 146]. In the context of PPARy agonist-induced responses, Sp1 is a linchpin, orchestrating the complex events that culminate in HO-1 induction in pulmonary resident cells. The involvement of Sp1 indicates a sophisticated regulatory network in which Sp1 acts as a pivotal mediator, coordinating PPARy signaling. Unraveling Sp1's role in this context would not only shed light on fundamental molecular mechanisms but also holds promise for the development of therapeutic strategies for treating diverse pulmonary disorders.

### The involvement of AP-1 in HO-1 expression

The AP-1 transcription factor complex, which is composed of members of the Jun (c-Jun, JunB, JunD), Fos (c-Fos, FosB, Fra1, Fra2), and activating TF (ATF) families, has emerged as a central player in cellular signaling, orchestrating various responses to oxidative and inflammatory stimuli [147, 148]. Activation of AP-1, which is induced by prooxidant and proinflammatory triggers, influences cellular outcomes. Notably, AP-1's involvement in the induction of the mouse HO-1 gene underscores its role in stress-responsive gene expression, indicating its importance in cellular stress adaptation [113, 149]. This process involves cooperative interactions between AP-1 and other essential transcription factors, such as USF2 and Sp1, which are bound to the regulatory regions of the HO-1 promoter [150, 151]. This interplay emphasizes the multifaceted nature of inducer-dependent HO-1 expression. In the context of pulmonary biology, where oxidative stress and inflammation converge, deciphering the nuanced connections between AP-1 and PPARy agonists is of paramount importance. These interactions, which affect HO-1 regulation, offer promising avenues for novel therapeutic interventions. Exploring the intricacies of AP-1-mediated HO-1 expression would not only advance our understanding of cellular stress responses but also identify potential strategies for mitigating oxidative stress-related disorders within the pulmonary milieu.

# The positive impact of PPARy agonists on inflammatory lung disorders

In the past decade, evidence has emerged regarding the potential advantages of PPARy agonists in treating various pulmonary inflammatory diseases. A cohort study conducted by Rinne et al. examined the impact of TZDs on COPD exacerbation in US veterans with COPD and diabetes [132]. Their research revealed a decrease in the risk of COPD exacerbation in patients treated with TZDs, and the incidence rate ratio was 0.86 (95% CI: 0.81-0.92) compared to those using non-TZDs [152]. Another study by the same team focused on diabetic veterans with asthma comorbidity and showed a significant reduction in the risk of asthma exacerbation in patients exposed to TZDs (OR = 0.79, 95% CI: 0.62-0.99) compared to those not exposed to TZDs. Moreover, patients in good compliance with diabetes medications experienced an even more substantial reduction in the risk of asthma exacerbation (OR=0.64, 95% CI: 0.47-0.85) [153]. These findings underscore the substantial reduction in the risk of COPD and asthma exacerbation in diabetic patients with COPD and asthma due to TZD administration. The consistent observation of the beneficial effects of TZDs in reducing severe COPD exacerbation is evident in the two most recent observational studies [154, 155]. Furthermore, there was a notable decrease in PPAR-gamma gene expression and a median decrease of 45% (P=0.008) observed from baseline to postchallenge in the bronchoalveolar lavage (BAL) fluid of allergic asthma patients. It is important to highlight that this decrease was not evident in the healthy control group [156]. Another study involving smokers with asthma demonstrated that rosiglitazone improved lung function, particularly in forced expiratory volume in 1 s (FEV(1)), compared to the effects of inhaled beclometasone dipropionate [157]. In contrast to these positive results, it is imperative to shed light on findings that deviate from the expected outcomes. For instance, Richards et al. conducted a single-center, double-blind, randomized, placebo-controlled, two-period crossover study exploring the effects of rosiglitazone, which is a PPARy agonist, on the late asthmatic reaction in an allergen challenge model of asthma [158]. Surprisingly, their results showed only a modest (15%) reduction, which did not reach statistical significance. We acknowledge that several factors, including the duration of the washout period, the dose of rosiglitazone (4 mg), and the severity of the patient conditions, may have contributed to this nonsignificant outcome. This divergence from the expected positive outcome prompts a more critical examination of the nuances surrounding the experimental design and patient characteristics. However, despite these positive outcomes, the mechanisms by which PPARy agonists affect lung inflammation remain unclear. Numerous studies have shown the anti-inflammatory effects of PPARy agonists on lung tissues. For example, in a mouse model of asthma induced by ovalbumin (OVA) inhalation, PPARy agonists such as rosiglitazone and pioglitazone reduced the increases in IL-17 mRNA and protein expression, airway inflammation, and bronchial hyperresponsiveness. Additionally, these agents abrogated increase in NF- $\kappa$ B activity in this asthma model [159]. Another study demonstrated that PPARy ligands such as rosiglitazone and pioglitazone reduced TNF- $\alpha$  and CC chemokine ligand-5 in the alveolar macrophages of COPD patients. Rosiglitazone also increased the gene expression of M2 macrophages and facilitated the clearance of apoptotic airway neutrophils in an in vivo model of pulmonary inflammation [160]. Moreover, rosiglitazone and SB219994 could inhibit airway neutrophilia induced by LPS and associated chemoattractants (keratinocyte-derived chemokine and granulocytecolony-stimulating factor) in an animal model [161]. A previous study suggested that a reduction in the expression of PPAR-y protein in the lungs could lead to pulmonary inflammation and lung injury. This study revealed that a decrease in PPAR-y protein expression and an increase in NF-KB activation occurred in LPSinduced acute lung injury in vivo [162]. In an asthma model induced by OVA, rosiglitazone alleviated airway inflammation by reducing peribronchiolar inflammatory cell infiltration, goblet cell hyperplasia, and mucus secretion. These effects were attributed to the inhibition of both NF-KB expression and the activation of the toll-like receptor 2 (TLR2)/nod-like receptor with pyrin domain-containing 3 (NLRP3) inflammatory corpuscle pathway by the PPARy agonist [163]. Furthermore, accumulating evidence suggests that rosiglitazone exerts anti-inflammatory effects by upregulating HO-1 in the pulmonary system [47, 108, 164, 165], indicating a potential therapeutic strategy for inflammatory diseases [43]. Our previous study demonstrated that rosiglitazone suppressed LPS-induced nuclear translocation of phosphorylated NF-KB (p65) and the expression of adhesion molecules through PPARydependent HO-1 upregulation. Additionally, we found that rosiglitazone inhibited LPS-induced lung inflammation through alternative PPARy-independent HO-1 induction [62]. Consequently, our results indicate that rosiglitazone-induced HO-1 expression was mediated by activation of the Nrf2 or PPARy cascade and



**Fig. 2** Schematic pathways for rosiglitazone-induced HO-1 expression in HPAEpiCs. Rosiglitazone triggers HO-1 expression in HPAEpiCs through two distinct pathways: PPARy-dependent and PPARy-independent mechanisms. PPARy-dependent pathway: rosiglitazone enhances HO-1 expression by activating a series of events, including PKCa, AMPKa, p38 MAPKa, SIRT1, Ac-PGC1a deacetylation, NCoR fragmentation, and direct binding of activated PPARy to the HO-1 promoter's responsive element. PPARy-independent pathway: in this pathway, rosiglitazone-induced HO-1 expression occurs via NOX/ROS-dependent phosphorylation of c-Src/Pyk2/Akt, leading to Nrf2 activation. Nrf2 then binds to the ARE region of the HO-1 promoter, stimulating HO-1 expression. Upregulation of HO-1 exerts anti-inflammatory effects, particularly on the expression of adhesion molecules like ICAM-1 and VCAM-1, associated with monocyte/leukocyte accumulation in pulmonary resident cells challenged with LPS. These pathways are adapted from prior studies [62, 70] and represent crucial mechanisms in mitigating inflammation in the lungs

suppressed inflammatory responses triggered by LPS (Fig. 2). The positive effects of TZDs on attenuating inflammatory responses by upregulating HO-1 suggest that rosiglitazone could be a potent treatment for pulmonary inflammatory diseases [45, 160, 166].

Beyond their well-established anti-inflammatory actions, PPAR agonists, particularly PPAR $\gamma$  agonists like rosiglitazone, play a significant role in the resolution phase of inflammation, a process crucial for restoring tissue homeostasis [167]. This phase involves not just the

suppression of proinflammatory signals but also actively promotes reparative and restorative processes in inflamed tissues. PPAR $\gamma$  agonists contribute to the resolution of inflammation through multiple mechanisms. They promote the polarization of macrophages towards the M2 phenotype, which are anti-inflammatory and essential for tissue repair and resolving inflammation [168]. These M2 macrophages play a crucial role in efferocytosis, the phagocytic clearance of apoptotic cells, preventing secondary necrosis and further inflammation. Additionally, PPARy agonists suppress the expression of proinflammatory mediators like cytokines and chemokines and activate anti-inflammatory signaling pathways, including the inhibition of NF-κB and AP-1 [169]. These actions help in controlling the balance between pro- and antiinflammatory signals. Furthermore, PPARy agonists modulate the recruitment of immune cells to inflammation sites by regulating the expression of adhesion molecules and chemokines, thus influencing the immune cell population towards an anti-inflammatory environment [170]. The study by Lea et al. in the European Respiratory Journal in 2014 underscores these findings, demonstrating that rosiglitazone, a PPARy agonist, increases M2 macrophage polarization and enhances efferocytosis in COPD alveolar macrophages. The study showed that COPD alveolar macrophages exhibit a skewed M2 phenotype and that treatment with rosiglitazone inhibited LPS-induced TNF- $\alpha$  and CCL-5 production [160]. It also increased the expression of CD36, HO-1, and PPARy, key elements in the phagocytosis of apoptotic neutrophils and the resolution of inflammation [171]. In a subchronic tobacco smoke mouse model, rosiglitazone significantly reduced inflammatory cells in bronchoalveolar lavage, further illustrating its role in resolving pulmonary inflammation [172]. These findings collectively indicate that PPARy agonists like rosiglitazone play a critical role in resolving inflammation by modulating macrophage function and enhancing efferocytosis, contributing to the restoration of pulmonary homeostasis and offering potential therapeutic avenues for chronic pulmonary diseases.

## Discussion

Understanding the mechanisms governing pulmonary inflammation is paramount in determining effective therapeutic strategies for diseases such as asthma, COPD, acute lung injury, and ARDS. This review examined the role of PPAR agonists, specifically PPARa, PPARB, and PPARy, in combating inflammatory and oxidative stress cascades in various pulmonary disorders. PPARa, PPARβ, and PPARγ exhibit differential tissue expression, indicating their involvement in different physiological functions. PPAR activation by endogenous and synthetic ligands can modulate inflammatory responses in the lungs. PPAR agonists inhibit crucial inflammatory mediators by disrupting pathways such as NF-KB, AP-1, and STATs. By interrupting these pathways, PPAR agonists exert potent anti-inflammatory effects, thereby halting chronic inflammation in the lungs. Pulmonary inflammation is characterized by oxidative stress. PPARs, especially PPARy, regulate oxidative stress by modulating the NOX/ROS pathway. Additionally, PPAR agonists enhance antioxidant defenses, such as HO-1, thereby protecting against oxidative damage. PPAR agonists, with their dual actions against inflammation and oxidative stress, offer a promising therapeutic approach for pulmonary diseases. Synthetic PPAR agonists, which allow for precise dose control, have potential as future treatments. Clinical translation of PPAR agonists requires addressing safety concerns, patient variability, and long-term effects. Rigorous trials are essential for establishing optimal doses and safety profiles. Exploring novel delivery methods and combination therapies, as well as identifying predictive biomarkers, is crucial for future advancements. The balance between inflammation and oxidative stress in pulmonary diseases necessitates comprehensive therapeutic strategies. PPAR agonists, with their multifaceted effects, represent a promising avenue for managing these conditions. Ongoing research and clinical exploration play pivotal roles in unlocking the potential of PPAR agonists and present novel possibilities for patients grappling with these complex disorders.

In this comprehensive review, we showed a complex web of signaling pathways and transcription factors that orchestrate the protective effects of PPARy agonists against pulmonary inflammation. The multifaceted nature of this regulation underscores the potential of targeting HO-1 expression as a therapeutic strategy for managing inflammatory lung diseases. PPARy, which is a nuclear receptor with diverse physiological functions, stands at the nexus of this regulatory network [34, 35]. Its activation by rosiglitazone not only induces HO-1 expression but also mitigates LPS-mediated inflammatory responses [62], demonstrating the dual protective effects of PPARy agonists against pulmonary inflammation. Our study shows that rosiglitazone-induced HO-1 expression occurs through both PPARy-dependent and PPARy-independent pathways, underscoring the complexity of PPARy-mediated responses. While the PPARy-dependent pathway involves Nrf2 activation and subsequent binding to ARE sites in the HO-1 promoter, the PPARy-independent pathway, intriguingly, does not rely on canonical mediators such as NOX/ROS, c-Src/ Pyk2, Akt, or Nrf2. These findings challenge existing paradigms and emphasize the need for further investigations to unravel the precise molecular mechanisms driving PPARy-independent responses in pulmonary epithelial cells. On the other hand, the overexpression of HO-1 in cells can have several negative consequences, necessitating strict regulatory mechanisms to maintain a balanced cellular environment [173]. Firstly, excessive HO-1 activity can disrupt cellular homeostasis. This enzyme plays a key role in the degradation of heme into biliverdin, carbon monoxide, and free iron. While these byproducts have protective roles at moderate levels, their overproduction can lead to toxicity and disrupt various cellular

processes, including oxidative stress responses and iron homeostasis [174]. Secondly, the accumulation of HO-1 byproducts such as carbon monoxide and biliverdin can be harmful. Carbon monoxide, although beneficial in small quantities as a signaling molecule, can be toxic at high concentrations, potentially impairing mitochondrial function and cellular respiration [175]. Biliverdin and its subsequent conversion to bilirubin are known for their antioxidant properties, but in excess, they can contribute to cellular stress and damage [174]. Finally, excessive HO-1 expression may trigger cytotoxic effects. While HO-1 is generally considered cytoprotective, primarily due to its role in reducing oxidative stress, its overexpression could paradoxically lead to cellular damage [176]. This could be due to an imbalance in prooxidant and antioxidant activities, leading to oxidative stress, or the dysregulation of other cellular pathways such as apoptosis or inflammation. In summary, while HO-1 is crucial for cellular protection against oxidative stress, its overexpression must be carefully regulated to prevent disturbances in cellular homeostasis, toxic accumulation of its byproducts, and potential cytotoxic effects. This underscores the importance of a balanced expression of HO-1 for maintaining cellular health and function.

Our study highlights Nrf2 as a pivotal mediator of rosiglitazone-induced HO-1 expression, particularly in the context of PPARy-independent pathways [62]. Nrf2, which is activated in response to oxidative stress, controls the expression of antioxidant genes, including HO-1, by binding to ARE sites [50, 51]. Rosiglitazone enhances Nrf2 binding to the ARE site in the HO-1 promoter, verifying its role in mediating HO-1 induction. This Nrf2mediated response is an attractive target for therapeutic interventions, and Nrf2 activation not only induces HO-1 but also regulates many antioxidant and cytoprotective genes, offering comprehensive cellular defense against oxidative insults. The interplay between SIRT1, PGC1 $\alpha$ , and HO-1 is a crucial mechanism in PPARy agonistinduced responses [127, 130]. Rosiglitazone-induced SIRT1 phosphorylation leads to the deacetylation of PGC1 $\alpha$ , facilitating its translocation to the nucleus [70]. Inside the nucleus, PGC1 $\alpha$  interacts with PPARy, resulting in the phosphorylation of PPARy. Additionally, a reduction in NCoR levels in the nucleus further enhances PPARy phosphorylation. This interplay underscores the nuanced regulation of HO-1 expression, highlighting the cross-talk between various transcriptional regulators in the cellular response to PPARy agonists. Sp1 and AP-1, which are transcription factors that are sensitive to oxidative stress and inflammatory signals, modulate HO-1 expression [147, 148, 150, 151]. Sp1, which is a central mediator of cellular responses, responds to a plethora of signals, orchestrating a finely tuned regulatory network [141, 142]. Our study showed the pivotal role of Sp1 in PPARy agonist-induced HO-1 expression, offering a potential target for therapeutic interventions. Similarly, AP-1, which is composed of diverse family members, serves as a central player in cellular signaling, responding to oxidative and inflammatory stimuli [147, 148, 150]. Deciphering the interactions between AP-1 and PPARy agonists provides insights into stress-responsive gene expression, paving the way for innovative strategies to mitigate oxidative stress-related disorders.

This comprehensive understanding of the molecular intricacies of PPARy agonist-induced HO-1 expression opens exciting avenues for future research and therapeutic interventions. Further investigations into PPARy-independent pathways to delineate the roles of unexplored mediators hold the key to identifying novel therapeutic targets. Additionally, exploring the interplay between different transcription factors and coregulators in the context of HO-1 regulation opens promising avenues for targeted therapies against pulmonary inflammatory diseases. Our previous studies provide a detailed roadmap of the signaling pathways and transcriptional regulators involved in PPARy agonist-induced HO-1 expression in pulmonary alveolar epithelial cells. These findings not only deepen our understanding of the molecular mechanisms governing pulmonary inflammation but also offer potential targets for innovative therapeutic strategies. As we continue to dissect the complexities of these regulatory networks, we move closer to revolutionizing our approach to combating pulmonary inflammatory disorders, ultimately improving the quality of life of patients worldwide.

In this review, we underscored the potential of TZDs, which are PPARy agonists, in shedding light on the molecular mechanisms governing HO-1 expression for the prevention and treatment of lung and airway inflammatory diseases (Fig. 2). The upregulation of HO-1 by rosiglitazone is mediated by both PPARy-dependent and PPARy-independent pathways in HPAEpiCs. In the PPARy-dependent pathway, rosiglitazone enhances HO-1 expression by activating a cascade involving PKCα, AMPKα, p38 MAPKα, SIRT1, Ac-PGC1α deacetylation, NCoR fragmentation, and the direct binding of activated PPARy to the HO-1 promoter's responsive element. Conversely, in the PPARy-independent pathway, rosiglitazone-induced HO-1 expression occurs via NOX/ROSdependent phosphorylation of c-Src/Pyk2/Akt, leading to Nrf2 activation and its binding to the ARE region of the HO-1 promoter. Upregulating HO-1 exerts anti-inflammatory effects, particularly affecting the expression of adhesion molecules such as ICAM-1 and VCAM-1, which are associated with monocyte/leukocyte accumulation in pulmonary resident cells challenged with LPS.

These findings highlight a novel approach for managing inflammatory pulmonary disorders and offers potential solutions to the increasing burden of chronic lung diseases worldwide.

Note: The pathways depicted in the figure are adapted from previous studies [62,70] and suggest that enhancing HO-1 by using TZDs is a promising strategy for addressing the challenges posed by chronic lung diseases globally.

#### Abbreviations

| ΑΜΡΚα             | 5'-adenosine monophosphate-activated protein kinase $\alpha$                   |
|-------------------|--|
| AP-1              | activator protein 1  |
| ARDS              | acute respiratory distress syndrome  |
| AREs              | antioxidant response elements  |
| ATP               | adenosine-5'-triphosphate  |
| cAMP              | cyclic adenosine monophosphate   |
| COPD              | chronic obstructive pulmonary disease  |
| COX-2             | cyclooxygenase-2   |
| cPLA <sub>2</sub> | cytosolic phospholipase $A_2$  |
| CREB              | cAMP response element-binding protein  |
| CBP               | CREB-binding protein   |
| CSE               | cigarette smoke extract  |
| DAG               | diacylglycerol   |
| EGFR              | epidermal growth factor receptor   |
| ERKs              | extracellular signal-regulated protein kinases                                 |
| GSK               | glycogen synthase kinase   |
| HO-1              | heme oxygenase-1   |
| HPAEpiCs          | human pulmonary alveolar epithelial cells                                      |
| ICAM-1            | intercellular adhesion molecule-1  |
| IL-1β             | interleukin-1β   |
| JNKs              | c-Jun NH2-terminal kinases   |
| LBD               | ligand-binding domain  |
| LPS               | lipopolysaccharide   |
| LTA               | lipoteichoic acid  |
| MAPKs             | mitogen-activated protein kinases  |
| MMP-9             | matrix metalloproteinase-9   |
| NCoR              | nuclear receptor co-repressor  |
| NF-ĸB             | nuclear factor к-light-chain-enhancer of activated B cells                     |
| NLRP3             | nod-like receptor with pyrin domain containing 3                               |
| NOX               | NADPH oxidase  |
| Nrf2              | nuclear factor E2-related factor 2   |
| OVA               | ovalbumin  |
| PDGFR             | platelet-derived growth factor receptor  |
| PGC-1a            | peroxisome proliferator-activated receptor gamma coactivator $1\text{-}\alpha$ |
| PGE <sub>2</sub>  | prostaglandin E <sub>2</sub>   |
| PGJ <sub>2</sub>  | prostaglandin J <sub>2</sub>   |
| PKC               | protein kinase C   |
| PPAR              | peroxisome proliferator-activated receptor                                     |
| PPRE              | peroxisome proliferator-activated receptor-response elements                   |
| Rac               | Ras-related C3 botulinum toxin substrate                                       |
| Redox             | reduction-oxidation  |
| REs               | regulatory elements  |
| ROS               | reactive oxygen species  |
| RXR               | 9-cis retinoic acid receptor   |
| siRNA             | small interfering RNA  |
| SIRT1             | silent information regulator type-1  |
| SRC-1             | steroid receptor coactivator 1   |
| STAT              | signal transducer and activator of transcription protein                       |
|                   | transcription factor   |
| TLR2              | toll-like receptor 2   |
| INF-α             | tumor necrosis factor-α  |
| 1ZDs              | thiazolidinediones   |
| VCAM-1            | vascular cell adhesion molecule-1  |

#### Authors' contributions

All authors conceived and designed the review, drafted the manuscript, prepared the figures, and edited the final manuscript.

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### Declarations

#### **Competing interests**

The authors declare no competing interests.

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#### References

- Fredenburgh LE, Perrella MA, Mitsialis SA. The role of heme oxygenase-1 in pulmonary disease. Am J Respir Cell Mol Biol. 2007;36:158–65.
- Lin CC, Lee IT, Yang YL, Lee CW, Kou YR, Yang CM. Induction of COX-2/ PGE<sub>2</sub>/IL-6 is crucial for cigarette smoke extract-induced airway inflammation: Role of TLR4-dependent NADPH oxidase activation. Free Radic Biol Med. 2010;48:240–54.
- Lee IT, Lin CC, Wu YC, Yang CM. TNF-α induces matrix metalloproteinase-9 expression in A549 cells: role of TNFR1/TRAF2/PKCα-dependent signaling pathways. J Cell Physiol. 2010;224:454–64.
- Liang KC, Lee CW, Lin WN, Lin CC, Wu CB, Luo SF, Yang CM. Interleukin-1β induces MMP-9 expression via p42/p44 MAPK, p38 MAPK, JNK, and nuclear factor-κB signaling pathways in human tracheal smooth muscle cells. J Cell Physiol. 2007;211:759–70.
- Lin WN, Luo SF, Lee CW, Wang CC, Wang JS, Yang CM. Involvement of MAPKs and NF-kB in LPS-induced VCAM-1 expression in human tracheal smooth muscle cells. Cell Signal. 2007;19:1258–67.
- Lin CC, Lee IT, Wu WL, Lin WN, Yang CM. Adenosine triphosphate regulates NADPH oxidase activity leading to hydrogen peroxide production and COX-2/PGE<sub>2</sub> expression in A549 cells. Am J Physiol Lung Cell Mol Physiol. 2012;303:L401–12.
- Cho RL, Yang CC, Lee IT, Lin CC, Chi PL, Hsiao LD, Yang CM. Lipopolysaccharide induces ICAM-1 expression via a c-Src/NADPH oxidase/ROSdependent NF-κB pathway in human pulmonary alveolar epithelial cells. Am J Physiol Lung Cell Mol Physiol. 2016;310:L639–57.
- Lee IT, Lee CW, Tung WH, Wang SW, Lin CC, Shu JC, Yang CM. Cooperation of TLR2 with MyD88, PI3K, and Rac1 in lipoteichoic acid-induced cPLA<sub>2</sub>/COX-2-dependent airway inflammatory responses. Am J Pathol. 2010;176:1671–84.
- Masjedy A, Salesi M, Ahmadi A, Salimian J, Azimzadeh Jamalkandi S. Association between single-nucleotide polymorphism of cytokines genes and chronic obstructive pulmonary disease: A systematic review and meta-analysis. Cytokine. 2023;171:156352.
- Page MJ, Kell DB, Pretorius E. The Role of Lipopolysaccharide-Induced Cell Signalling in Chronic Inflammation. Chronic Stress (Thousand Oaks). 2022;6:24705470221076390.
- 11. Lee IT, Yang C-M. Inflammatory Signalings Involved in Airway and Pulmonary Diseases. Mediators Inflamm. 2013;2013:791231.
- Cheng SE, Lee IT, Lin CC, Wu WL, Hsiao LD, Yang CM. ATP mediates NADPH oxidase/ROS generation and COX-2/PGE<sub>2</sub> expression in A549 cells: role of P2 receptor-dependent STAT3 activation. PLoS One. 2013;8:e54125.
- Ferrer MD, Busquets-Cortés C, Capó X, Tejada S, Tur JA, Pons A, Sureda A. Cyclooxygenase-2 Inhibitors as a Therapeutic Target in Inflammatory Diseases. Curr Med Chem. 2019;26:3225–41.
- Cheng SE, Lin CC, Lee IT, Hsu CK, Kou YR, Yang CM. Cigarette smoke extract regulates cytosolic phospholipase A2 expression via NADPH oxidase/MAPKs/AP-1 and p300 in human tracheal smooth muscle cells. J Cell Biochem. 2011;112:589–99.
- Lee CW, Lin CC, Lee IT, Lee HC, Yang CM. Activation and induction of cytosolic phospholipase A2 by TNF-α mediated through Nox2, MAPKs,

NF-κB, and p300 in human tracheal smooth muscle cells. J Cell Physiol. 2011;226:2103–14.

- Lin CC, Kuo CT, Cheng CY, Wu CY, Lee CW, Hsieh HL, Lee IT, Yang CM. IL-1 beta promotes A549 cell migration via MAPKs/AP-1- and NFkappaB-dependent matrix metalloproteinase-9 expression. Cell Signal. 2009;21:1652–62.
- Lin CC, Tseng HW, Hsieh HL, Lee CW, Wu CY, Cheng CY, Yang CM. Tumor necrosis factor-alpha induces MMP-9 expression via p42/p44 MAPK, JNK, and nuclear factor-kappaB in A549 cells. Toxicol Appl Pharmacol. 2008;229:386–98.
- Lin FS, Lin CC, Chien CS, Luo SF, Yang CM. Involvement of p42/p44 MAPK, JNK, and NF-kappaB in IL-1beta-induced ICAM-1 expression in human pulmonary epithelial cells. J Cell Physiol. 2005;202:464–73.
- Singh V, Kaur R, Kumari P, Pasricha C, Singh R. ICAM-1 and VCAM-1: Gatekeepers in various inflammatory and cardiovascular disorders. Clin Chim Acta. 2023;548:117487.
- Lee CW, Lin CC, Luo SF, Lee HC, Lee IT, Aird WC, Hwang TL, Yang CM. Tumor necrosis factor-alpha enhances neutrophil adhesiveness: induction of vascular cell adhesion molecule-1 via activation of Akt and CaM kinase II and modifications of histone acetyltransferase and histone deacetylase 4 in human tracheal smooth muscle cells. Mol Pharmacol. 2008;73:1454–64.
- Luo SF, Chang CC, Lee IT, Lee CW, Lin WN, Lin CC, Yang CM. Activation of ROS/NF-kappaB and Ca2+/CaM kinase II are necessary for VCAM-1 induction in IL-1 beta-treated human tracheal smooth muscle cells. Toxicol Appl Pharmacol. 2009;237:8–21.
- 22. Yang X, Liu X, Nie Y, Zhan F, Zhu B. Oxidative stress and ROS-mediated cellular events in RSV infection: potential protective roles of antioxidants. Virol J. 2023;20:224.
- Auboeuf D, Rieusset J, Fajas L, Vallier P, Frering V, Riou JP, Staels B, Auwerx J, Laville M, Vidal HJD. Tissue distribution and quantification of the expression of mRNAs of peroxisome proliferator–activated receptors and liver X receptor-α in humans: no alteration in adipose tissue of obese and NIDDM patients. Diabetes. 1997;46:1319–27.
- Mukherjee R, Jow L, Croston GE. Paterniti JRJJoBC: Identification, characterization, and tissue distribution of human peroxisome proliferator-activated receptor (PPAR) isoforms PPARy2 versus PPARy1 and activation with retinoid X receptor agonists and antagonists. J Biol Chem. 1997;272:8071–6.
- Chen H, Tan H, Wan J, Zeng Y, Wang J, Wang H, Lu X. PPAR-γ signaling in nonalcoholic fatty liver disease: Pathogenesis and therapeutic targets. Pharmacol Ther. 2023;245:108391.
- Gu L, Ju Y, Hu M, Zheng M, Li Q, Zhang X. Research progress of PPARγ regulation of cholesterol and inflammation in Alzheimer's disease. Metab Brain Dis. 2023;38:839–54.
- Ji X, Zhang W, Yin L, Shi Z, Luan J, Chen L, Liu L. The Potential Roles of Post-Translational Modifications of PPARγ in Treating Diabetes. Biomolecules. 2022;12(12):1832.
- Qiu YY, Zhang J, Zeng FY, Zhu YZ. Roles of the peroxisome proliferatoractivated receptors (PPARs) in the pathogenesis of nonalcoholic fatty liver disease (NAFLD). Pharmacol Res. 2023;192:106786.
- Wagner N, Wagner KD. Pharmacological Utility of PPAR Modulation for Angiogenesis in Cardiovascular Disease. Int J Mol Sci. 2023;24:2345.
- Yang XF, Shang DJ. The role of peroxisome proliferator-activated receptor γ in lipid metabolism and inflammation in atherosclerosis. Cell Biol Int. 2023;47:1469–87.
- 31. Lebovitz HE. Differentiating members of the thiazolidinedione class: a focus on safety. Diabetes Metab Res Rev. 2002;18(Suppl 2):S23–9.
- 32. Scheen AJ. Hepatotoxicity with thiazolidinediones: is it a class effect? Drug Saf. 2001;24:873–88.
- Xu B, Xing A, Li S. The forgotten type 2 diabetes mellitus medicine: rosiglitazone. Diabetol Int. 2022;13:49–65.
- 34. Desvergne B, Wahli W. Peroxisome proliferator-activated receptors: nuclear control of metabolism. Endocr Rev. 1999;20:649–88.
- Kliewer SA, Forman BM, Blumberg B, Ong ES, Borgmeyer U, Mangelsdorf DJ, Umesono K, Evans RM. Differential expression and activation of a family of murine peroxisome proliferator-activated receptors. Proc Natl Acad Sci U S A. 1994;91:7355–9.
- Beck F, Plummer S, Senior P, Byrne S, Green S. Brammar WJPotRSoLSBBS: The ontogeny of peroxisome-proliferator-activated receptor gene expression in the mouse and rat. Proc Biol Sci. 1992;247:83–7.

- 37. Braissant O, Wahli WJE. Differential expression of peroxisome proliferator-activated receptor- $\alpha$ ,- $\beta$ , and- $\gamma$  during rat embryonic development. Endocrinology. 1998;139:2748–54.
- Schoonjans K, Staels B, Auwerx J. The peroxisome proliferator activated receptors (PPARS) and their effects on lipid metabolism and adipocyte differentiation. Biochim Biophys Acta. 1996;1302:93–109.
- Chinetti G, Fruchart JC, Staels B. Peroxisome proliferator-activated receptors (PPARs): nuclear receptors at the crossroads between lipid metabolism and inflammation. Inflamm Res. 2000;49:497–505.
- Li J, Shen X. Effect of rosiglitazone on inflammatory cytokines and oxidative stress after intensive insulin therapy in patients with newly diagnosed type 2 diabetes. Diabetol Metab Syndr. 2019;11:35.
- Engdahl R, Monroy MA, Daly JM. 15-Deoxy-Delta12,14-prostaglandin J2 (15d-PGJ2) mediates repression of TNF-alpha by decreasing levels of acetylated histone H3 and H4 at its promoter. Biochem Biophys Res Commun. 2007;359:88–93.
- 42. Belvisi MG, Hele DJ. Peroxisome proliferator-activated receptors as novel targets in lung disease. Chest. 2008;134:152–7.
- 43. Belvisi MG, Mitchell JA. Targeting PPAR receptors in the airway for the treatment of inflammatory lung disease. Br J Pharmacol. 2009;158:994–1003.
- 44. Ryu SL, Shim JW, Kim DS, Jung HL, Park MS, Park SH, Lee J, Lee WY, Shim JY. Expression of peroxisome proliferator-activated receptor (PPAR)-α and PPAR-γ in the lung tissue of obese mice and the effect of rosiglitazone on proinflammatory cytokine expressions in the lung tissue. Korean J Pediatr. 2013;56:151–8.
- 45. Xu J, Zhu YT, Wang GZ, Han D, Wu YY, Zhang DX, Liu Y, Zhang YH, Xie XM, Li SJ, et al. The PPARγ agonist, rosiglitazone, attenuates airway inflammation and remodeling via heme oxygenase-1 in murine model of asthma. Acta Pharmacol Sin. 2015;36:171–8.
- 46. Ferguson HE, Thatcher TH, Olsen KC, Garcia-Bates TM, Baglole CJ, Kottmann RM, Strong ER, Phipps RP, Sime PJ. Peroxisome proliferatoractivated receptor-gamma ligands induce heme oxygenase-1 in lung fibroblasts by a PPARgamma-independent, glutathione-dependent mechanism. Am J Physiol Lung Cell Mol Physiol. 2009;297:L912–9.
- Krönke G, Kadl A, Ikonomu E, Blüml S, Fürnkranz A, Sarembock IJ, Bochkov VN, Exner M, Binder BR, Leitinger N. Expression of heme oxygenase-1 in human vascular cells is regulated by peroxisome proliferatoractivated receptors. Arterioscler Thromb Vasc Biol. 2007;27:1276–82.
- Zhang D, Wang G, Han D, Zhang Y, Xu J, Lu J, Li S, Xie X, Liu L, Dong L, Li M. Activation of PPAR-y ameliorates pulmonary arterial hypertension via inducing heme oxygenase-1 and p21(WAF1): an in vivo study in rats. Life Sci. 2014;98:39–43.
- 49. Kamata H, Hirata H. Redox regulation of cellular signalling. Cell Signal. 1999;11:1–14.
- Loboda A, Damulewicz M, Pyza E, Jozkowicz A, Dulak J. Role of Nrf2/HO-1 system in development, oxidative stress response and diseases: an evolutionarily conserved mechanism. Cell Mol Life Sci. 2016;73:3221–47.
- Seo WY, Goh AR, Ju SM, Song HY, Kwon DJ, Jun JG, Kim BC, Choi SY, Park J. Celastrol induces expression of heme oxygenase-1 through ROS/ Nrf2/ARE signaling in the HaCaT cells. Biochem Biophys Res Commun. 2011;407:535–40.
- Rushworth SA, Chen XL, Mackman N, Ogborne RM, O'Connell MA. Lipopolysaccharide-induced heme oxygenase-1 expression in human monocytic cells is mediated via Nrf2 and protein kinase C. J Immunol. 2005;175:4408–15.
- Huang J, Shen XD, Yue S, Zhu J, Gao F, Zhai Y, Busuttil RW, Ke B, Kupiec-Weglinski JW. Adoptive transfer of heme oxygenase-1 (HO-1)-modified macrophages rescues the nuclear factor erythroid 2-related factor (Nrf2) antiinflammatory phenotype in liver ischemia/reperfusion injury. Mol Med. 2014;20:448–55.
- Jamal Uddin M, Joe Y, Kim SK, Oh Jeong S, Ryter SW, Pae HO, Chung HT. IRG1 induced by heme oxygenase-1/carbon monoxide inhibits LPSmediated sepsis and pro-inflammatory cytokine production. Cell Mol Immunol. 2016;13:170–9.
- 55. Srisook K, Han SS, Choi HS, Li MH, Ueda H, Kim C, Cha YN. CO from enhanced HO activity or from CORM-2 inhibits both O2- and NO production and downregulates HO-1 expression in LPS-stimulated macrophages. Biochem Pharmacol. 2006;71:307–18.
- Polvani S, Tarocchi M, Galli A. PPARγ and Oxidative Stress: Con(β) Catenating NRF2 and FOXO. PPAR Res. 2012;2012:641087.

- Gardner OS, Dewar BJ, Graves LM. Activation of mitogen-activated protein kinases by peroxisome proliferator-activated receptor ligands: an example of nongenomic signaling. Mol Pharmacol. 2005;68:933–41.
- Koyani CN, Kitz K, Rossmann C, Bernhart E, Huber E, Trummer C, Windischhofer W, Sattler W, Malle E. Activation of the MAPK/Akt/Nrf2-Egr1/ HO-1-GCLc axis protects MG-63 osteosarcoma cells against 15d-PGJ2mediated cell death. Biochem Pharmacol. 2016;104:29–41.
- Donovan C, Tan X, Bourke JE. PPARY Ligands Regulate Noncontractile and Contractile Functions of Airway Smooth Muscle: Implications for Asthma Therapy. PPAR Res. 2012;2012:809164.
- Tickner J, Fan LM, Du J, Meijles D, Li JM. Nox2-derived ROS in PPARy signaling and cell-cycle progression of lung alveolar epithelial cells. Free Radic Biol Med. 2011;51:763–72.
- Lam GY, Huang J, Brumell JH. The many roles of NOX2 NADPH oxidasederived ROS in immunity. Semin Immunopathol. 2010;32:415–30.
- Cho RL, Yang CC, Tseng HC, Hsiao LD, Lin CC, Yang CM. Haem oxygenase-1 up-regulation by rosiglitazone via ROS-dependent Nrf2-antioxidant response elements axis or PPARy attenuates LPS-mediated lung inflammation. Br J Pharmacol. 2018;175:3928–46.
- Barbieri SS, Cavalca V, Eligini S, Brambilla M, Caiani A, Tremoli E, Colli S. Apocynin prevents cyclooxygenase 2 expression in human monocytes through NADPH oxidase and glutathione redox-dependent mechanisms. Free Radic Biol Med. 2004;37:156–65.
- Ke K, Sul OJ, Choi EK, Safdar AM, Kim ES, Choi HS. Reactive oxygen species induce the association of SHP-1 with c-Src and the oxidation of both to enhance osteoclast survival. Am J Physiol Endocrinol Metab. 2014;307:E61–70.
- Peng G, Huang J, Boyd M, Kleinberg ME. Properties of phagocyte NADPH oxidase p47-phox mutants with unmasked SH3 (Src homology 3) domains: full reconstitution of oxidase activity in a semi-recombinant cell-free system lacking arachidonic acid. Biochem J. 2003;373:221–9.
- Dewar BJ, Gardner OS, Chen CS, Earp HS, Samet JM, Graves LM. Capacitative calcium entry contributes to the differential transactivation of the epidermal growth factor receptor in response to thiazolidinediones. Mol Pharmacol. 2007;72:1146–56.
- 67. Chi PL, Lin CC, Chen YW, Hsiao LD, Yang CM. CO Induces Nrf2-Dependent Heme Oxygenase-1 Transcription by Cooperating with Sp1 and c-Jun in Rat Brain Astrocytes. Mol Neurobiol. 2015;52:277–92.
- Han Z, Varadharaj S, Giedt RJ, Zweier JL, Szeto HH, Alevriadou BR. Mitochondria-derived reactive oxygen species mediate heme oxygenase-1 expression in sheared endothelial cells. J Pharmacol Exp Ther. 2009;329:94–101.
- Yang CM, Lin CC, Lee IT, Hsu CK, Tai YC, Hsieh HL, Chi PL, Hsiao LD. c-Src-dependent transactivation of EGFR mediates CORM-2-induced HO-1 expression in human tracheal smooth muscle cells. J Cell Physiol. 2015;230:2351–61.
- Cho RL, Lin WN, Wang CY, Yang CC, Hsiao LD, Lin CC, Yang CM. Heme oxygenase-1 induction by rosiglitazone via PKCα/AMPKα/p38 MAPKα/ SIRT1/PPARγ pathway suppresses lipopolysaccharide-mediated pulmonary inflammation. Biochem Pharmacol. 2018;148:222–37.
- Papaiahgari S, Zhang Q, Kleeberger SR, Cho HY, Reddy SP. Hyperoxia stimulates an Nrf2-ARE transcriptional response via ROS-EGFR-PI3K-Akt/ ERK MAP kinase signaling in pulmonary epithelial cells. Antioxid Redox Signal. 2006;8:43–52.
- Fan HC, Fernández-Hernando C, Lai JH. Protein kinase C isoforms in atherosclerosis: pro- or anti-inflammatory? Biochem Pharmacol. 2014;88:139–49.
- Passos GF, Medeiros R, Marcon R, Nascimento AF, Calixto JB, Pianowski LF. The role of PKC/ERK1/2 signaling in the anti-inflammatory effect of tetracyclic triterpene euphol on TPA-induced skin inflammation in mice. Eur J Pharmacol. 2013;698:413–20.
- 74. Isakov N, Altman A. Regulation of immune system cell functions by protein kinase C. Front Immunol. 2013;4:384.
- Gould CM, Newton AC. The life and death of protein kinase C. Curr Drug Targets. 2008;9:614–25.
- Mylroie H, Dumont O, Bauer A, Thornton CC, Mackey J, Calay D, Hamdulay SS, Choo JR, Boyle JJ, Samarel AM, et al. PKCe-CREB-Nrf2 signalling induces HO-1 in the vascular endothelium and enhances resistance to inflammation and apoptosis. Cardiovasc Res. 2015;106:509–19.
- Dempsey EC, Cool CD, Littler CM. Lung disease and PKCs. Pharmacol Res. 2007;55:545–59.

- Rojo AI, Salina M, Salazar M, Takahashi S, Suske G, Calvo V, de Sagarra MR, Cuadrado A. Regulation of heme oxygenase-1 gene expression through the phosphatidylinositol 3-kinase/PKC-zeta pathway and Sp1. Free Radic Biol Med. 2006;41:247–61.
- Nakashima S. Protein kinase C alpha (PKC alpha): regulation and biological function. J Biochem. 2002;132:669–75.
- Paumelle R, Blanquart C, Briand O, Barbier O, Duhem C, Woerly G, Percevault F, Fruchart JC, Dombrowicz D, Glineur C, Staels B. Acute antiinflammatory properties of statins involve peroxisome proliferatoractivated receptor-alpha via inhibition of the protein kinase C signaling pathway. Circ Res. 2006;98:361–9.
- Hsu HY, Chu LC, Hua KF, Chao LK. Heme oxygenase-1 mediates the anti-inflammatory effect of Curcumin within LPS-stimulated human monocytes. J Cell Physiol. 2008;215:603–12.
- Zhang Z, Cui W, Li G, Yuan S, Xu D, Hoi MP, Lin Z, Dou J, Han Y, Lee SM. Baicalein protects against 6-OHDA-induced neurotoxicity through activation of Keap1/Nrf2/HO-1 and involving PKCα and PI3K/AKT signaling pathways. J Agric Food Chem. 2012;60:8171–82.
- Lee WH, Kim SG. AMPK-Dependent Metabolic Regulation by PPAR Agonists. PPAR Res. 2010;2010:549101.
- Liu XM, Peyton KJ, Shebib AR, Wang H, Korthuis RJ, Durante W. Activation of AMPK stimulates heme oxygenase-1 gene expression and human endothelial cell survival. Am J Physiol Heart Circ Physiol. 2011;300:H84–93.
- Sag D, Carling D, Stout RD, Suttles J. Adenosine 5<sup>'</sup>-monophosphateactivated protein kinase promotes macrophage polarization to an antiinflammatory functional phenotype. J Immunol. 2008;181:8633–41.
- Salminen A, Hyttinen JM, Kaarniranta K. AMP-activated protein kinase inhibits NF-κB signaling and inflammation: impact on healthspan and lifespan. J Mol Med (Berl). 2011;89:667–76.
- Lee JS, Park SJ, Cho YS, Huh JW, Oh YM, Lee SD. Role of AMP-Activated Protein Kinase (AMPK) in Smoking-Induced Lung Inflammation and Emphysema. Tuberc Respir Dis (Seoul). 2015;78:8–17.
- Gejjalagere Honnappa C, Mazhuvancherry Kesavan U. A concise review on advances in development of small molecule anti-inflammatory therapeutics emphasising AMPK: An emerging target. Int J Immunopathol Pharmacol. 2016;29:562–71.
- Cheng XY, Li YY, Huang C, Li J, Yao HW. AMP-activated protein kinase reduces inflammatory responses and cellular senescence in pulmonary emphysema. Oncotarget. 2017;8:22513–23.
- Li J, Miller EJ, Ninomiya-Tsuji J, Russell RR III, Young LH. AMP-activated protein kinase activates p38 mitogen-activated protein kinase by increasing recruitment of p38 MAPK to TAB1 in the ischemic heart. Circ Res. 2005;97:872–9.
- Kim JH, Park JM, Kim EK, Lee JO, Lee SK, Jung JH, You GY, Park SH, Suh PG, Kim HS. Curcumin stimulates glucose uptake through AMPK-p38 MAPK pathways in L6 myotube cells. J Cell Physiol. 2010;223:771–8.
- 92. Alam J, Cook JL. How many transcription factors does it take to turn on the heme oxygenase-1 gene? Am J Respir Cell Mol Biol. 2007;36:166–74.
- McNally SJ, Harrison EM, Ross JA, Garden OJ, Wigmore SJ. Curcumin induces heme oxygenase 1 through generation of reactive oxygen species, p38 activation and phosphatase inhibition. Int J Mol Med. 2007;19:165–72.
- Feinstein DL, Spagnolo A, Akar C, Weinberg G, Murphy P, Gavrilyuk V, Dello Russo C. Receptor-independent actions of PPAR thiazolidinedione agonists: is mitochondrial function the key? Biochem Pharmacol. 2005;70:177–88.
- Wang X, Wang Z, Liu JZ, Hu JX, Chen HL, Li WL, Hai CX. Double antioxidant activities of rosiglitazone against high glucose-induced oxidative stress in hepatocyte. Toxicol In Vitro. 2011;25:839–47.
- Kietzmann T, Samoylenko A, Immenschuh S. Transcriptional regulation of heme oxygenase-1 gene expression by MAP kinases of the JNK and p38 pathways in primary cultures of rat hepatocytes. J Biol Chem. 2003;278:17927–36.
- Shan Y, Pepe J, Lu TH, Elbirt KK, Lambrecht RW, Bonkovsky HL. Induction of the heme oxygenase-1 gene by metalloporphyrins. Arch Biochem Biophys. 2000;380:219–27.
- Ohlmann A, Giffhorn-Katz S, Becker I, Katz N, Immenschuh S. Regulation of heme oxygenase-1 gene expression by anoxia and reoxygenation in primary rat hepatocyte cultures. Exp Biol Med (Maywood). 2003;228:584–9.

- Balogun E, Hoque M, Gong P, Killeen E, Green CJ, Foresti R, Alam J, Motterlini R. Curcumin activates the haem oxygenase-1 gene via regulation of Nrf2 and the antioxidant-responsive element. Biochem J. 2003;371:887–95.
- Lim HJ, Lee KS, Lee S, Park JH, Choi HE, Go SH, Kwak HJ, Park HY. 15d-PGJ2 stimulates HO-1 expression through p38 MAP kinase and Nrf-2 pathway in rat vascular smooth muscle cells. Toxicol Appl Pharmacol. 2007;223:20–7.
- 101. Koyasu S. The role of PI3K in immune cells. Nat Immunol. 2003;4:313–9.
- Ghigo A, Damilano F, Braccini L, Hirsch E. PI3K inhibition in inflammation: Toward tailored therapies for specific diseases. Bioessays. 2010;32:185–96.
- 103. Martin D, Rojo AI, Salinas M, Diaz R, Gallardo G, Alam J, De Galarreta CM, Cuadrado A. Regulation of heme oxygenase-1 expression through the phosphatidylinositol 3-kinase/Akt pathway and the Nrf2 transcription factor in response to the antioxidant phytochemical carnosol. J Biol Chem. 2004;279:8919–29.
- Ryter SW, Alam J, Choi AM. Heme oxygenase-1/carbon monoxide: from basic science to therapeutic applications. Physiol Rev. 2006;86:583–650.
- 105. Alvarez-Maqueda M, El Bekay R, Alba G, Monteseirín J, Chacón P, Vega A, Martín-Nieto J, Bedoya FJ, Pintado E, Sobrino F. 15-deoxy-delta 12,14-prostaglandin J2 induces heme oxygenase-1 gene expression in a reactive oxygen species-dependent manner in human lymphocytes. J Biol Chem. 2004;279(21929–21):937.
- 106. Hamdulay SS, Wang B, Birdsey GM, Ali F, Dumont O, Evans PC, Haskard DO, Wheeler-Jones CP, Mason JC. Celecoxib activates PI-3 K/Akt and mitochondrial redox signaling to enhance heme oxygenase-1-mediated anti-inflammatory activity in vascular endothelium. Free Radic Biol Med. 2010;48:1013–23.
- Deng X, Rui W, Zhang F, Ding W. PM2.5 induces Nrf2-mediated defense mechanisms against oxidative stress by activating PIK3/AKT signaling pathway in human lung alveolar epithelial A549 cells. Cell Biol Toxicol. 2013;29:143–57.
- Xu X, Li H, Hou X, Li D, He S, Wan C, Yin P, Liu M, Liu F, Xu J. Punicalagin Induces Nrf2/HO-1 Expression via Upregulation of PI3K/AKT Pathway and Inhibits LPS-Induced Oxidative Stress in RAW264.7 Macrophages. Mediators Inflamm. 2015;2015:380218.
- 109. Ricchetti GA, Williams LM, Foxwell BM. Heme oxygenase 1 expression induced by IL-10 requires STAT-3 and phosphoinositol-3 kinase and is inhibited by lipopolysaccharide. J Leukoc Biol. 2004;76:719–26.
- Zhang X, Shan P, Jiang G, Zhang SS, Otterbein LE, Fu XY, Lee PJ. Endothelial STAT3 is essential for the protective effects of HO-1 in oxidant-induced lung injury. Faseb J. 2006;20:2156–8.
- 111. Tron K, Samoylenko A, Musikowski G, Kobe F, Immenschuh S, Schaper F, Ramadori G, Kietzmann T. Regulation of rat heme oxygenase-1 expression by interleukin-6 via the Jak/STAT pathway in hepatocytes. J Hepatol. 2006;45:72–80.
- 112. Weis N, Weigert A, von Knethen A, Brüne B. Heme oxygenase-1 contributes to an alternative macrophage activation profile induced by apoptotic cell supernatants. Mol Biol Cell. 2009;20:1280–8.
- Alam J, Camhi S, Choi AM. Identification of a second region upstream of the mouse heme oxygenase-1 gene that functions as a basal level and inducer-dependent transcription enhancer. J Biol Chem. 1995;270(11977–11):984.
- 114. Immenschuh S, Ramadori G. Gene regulation of heme oxygenase-1 as a therapeutic target. Biochem Pharmacol. 2000;60:1121–8.
- 115. Hondares E, Mora O, Yubero P, Rodriguez de la Concepción M, Iglesias R, Giralt M, Villarroya F. Thiazolidinediones and rexinoids induce peroxisome proliferator-activated receptor-coactivator (PGC)-1alpha gene transcription: an autoregulatory loop controls PGC-1alpha expression in adipocytes via peroxisome proliferator-activated receptor-gamma coactivation. Endocrinology. 2006;147:2829–38.
- 116. Nguyen T, Sherratt PJ, Pickett CB. Regulatory mechanisms controlling gene expression mediated by the antioxidant response element. Annu Rev Pharmacol Toxicol. 2003;43:233–60.
- Kensler TW, Wakabayashi N, Biswal S. Cell survival responses to environmental stresses via the Keap1-Nrf2-ARE pathway. Annu Rev Pharmacol Toxicol. 2007;47:89–116.

- Yamada N, Yamaya M, Okinaga S, Nakayama K, Sekizawa K, Shibahara S, Sasaki H. Microsatellite polymorphism in the heme oxygenase-1 gene promoter is associated with susceptibility to emphysema. Am J Hum Genet. 2000;66:187–95.
- 119. Exner M, Minar E, Wagner O, Schillinger M. The role of heme oxygenase-1 promoter polymorphisms in human disease. Free Radic Biol Med. 2004;37:1097–104.
- Itoh K, Wakabayashi N, Katoh Y, Ishii T, Igarashi K, Engel JD, Yamamoto M. Keap1 represses nuclear activation of antioxidant responsive elements by Nrf2 through binding to the amino-terminal Neh2 domain. Genes Dev. 1999;13:76–86.
- 121. Foresti R, Bains SK, Pitchumony TS, de Castro Brás LE, Drago F, Dubois-Randé JL, Bucolo C, Motterlini R. Small molecule activators of the Nrf2-HO-1 antioxidant axis modulate heme metabolism and inflammation in BV2 microglia cells. Pharmacol Res. 2013;76:132–48.
- Baird L, Llères D, Swift S, Dinkova-Kostova AT. Regulatory flexibility in the Nrf2-mediated stress response is conferred by conformational cycling of the Keap1-Nrf2 protein complex. Proc Natl Acad Sci U S A. 2013;110:15259–64.
- 123. Hira S, Tomita T, Matsui T, Igarashi K, Ikeda-Saito M. Bach1, a hemedependent transcription factor, reveals presence of multiple heme binding sites with distinct coordination structure. IUBMB Life. 2007;59:542–51.
- Espinosa-Diez C, Miguel V, Mennerich D, Kietzmann T, Sánchez-Pérez P, Cadenas S, Lamas S. Antioxidant responses and cellular adjustments to oxidative stress. Redox Biol. 2015;6:183–97.
- 125. Yoshizaki T, Schenk S, Imamura T, Babendure JL, Sonoda N, Bae EJ, Oh DY, Lu M, Milne JC, Westphal C, et al. SIRT1 inhibits inflammatory pathways in macrophages and modulates insulin sensitivity. Am J Physiol Endocrinol Metab. 2010;298:E419–28.
- Gao R, Ma Z, Hu Y, Chen J, Shetty S, Fu J. Sirt1 restrains lung inflammasome activation in a murine model of sepsis. Am J Physiol Lung Cell Mol Physiol. 2015;308:L847–53.
- 127. Lee BW, Chun SW, Kim SH, Lee Y, Kang ES, Cha BS, Lee HC. Lithospermic acid B protects  $\beta$ -cells from cytokine-induced apoptosis by alleviating apoptotic pathways and activating anti-apoptotic pathways of Nrf2-HO-1 and Sirt1. Toxicol Appl Pharmacol. 2011;252:47–54.
- 128. Kim SR, Lee KS, Park SJ, Min KH, Choe YH, Moon H, Yoo WH, Chae HJ, Han MK, Lee YC. Involvement of sirtuin 1 in airway inflammation and hyperresponsiveness of allergic airway disease. J Allergy Clin Immunol. 2010;125:449–460.e414.
- 129. Yang SR, Wright J, Bauter M, Seweryniak K, Kode A, Rahman I. Sirtuin regulates cigarette smoke-induced proinflammatory mediator release via ReIA/p65 NF-kappaB in macrophages in vitro and in rat lungs in vivo: implications for chronic inflammation and aging. Am J Physiol Lung Cell Mol Physiol. 2007;292:L567–76.
- Ruderman NB, Xu XJ, Nelson L, Cacicedo JM, Saha AK, Lan F, Ido Y. AMPK and SIRT1: a long-standing partnership? Am J Physiol Endocrinol Metab. 2010;298:E751–60.
- Jia N, Sun Q, Su Q, Chen G. SIRT1-mediated deacetylation of PGC1α attributes to the protection of curcumin against glutamate excitotoxicity in cortical neurons. Biochem Biophys Res Commun. 2016;478:1376–81.
- 132. Ali F, Ali NS, Bauer A, Boyle JJ, Hamdulay SS, Haskard DO, Randi AM, Mason JC. PPARdelta and PGC1alpha act cooperatively to induce haem oxygenase-1 and enhance vascular endothelial cell resistance to stress. Cardiovasc Res. 2010;85:701–10.
- 133. Ghisletti S, Huang W, Jepsen K, Benner C, Hardiman G, Rosenfeld MG, Glass CK. Cooperative NCoR/SMRT interactions establish a corepressorbased strategy for integration of inflammatory and anti-inflammatory signaling pathways. Genes Dev. 2009;23:681–93.
- Rodgers JT, Lerin C, Haas W, Gygi SP, Spiegelman BM, Puigserver P. Nutrient control of glucose homeostasis through a complex of PGC-1alpha and SIRT1. Nature. 2005;434:113–8.
- 135. Rai A, Tripathi S, Kushwaha R, Singh P, Srivastava P, Sanyal S, Bandyopadhyay S. CDK5-induced p-PPARγ(Ser 112) downregulates GFAP via PPREs in developing rat brain: effect of metal mixture and troglitazone in astrocytes. Cell Death Dis. 2014;5:e1033.
- Burns KA, Vanden Heuvel JP. Modulation of PPAR activity via phosphorylation. Biochim Biophys Acta. 2007;1771:952–60.

- Lin CF, Young KC, Bai CH, Yu BC, Ma CT, Chien YC, Chiang CL, Liao CS, Lai HW, Tsao CW. Rosiglitazone regulates anti-inflammation and growth inhibition via PTEN. Biomed Res Int. 2014;2014:787924.
- Lv H, Yu Z, Zheng Y, Wang L, Qin X, Cheng G, Ci X. Isovitexin Exerts Anti-Inflammatory and Anti-Oxidant Activities on Lipopolysaccharide-Induced Acute Lung Injury by Inhibiting MAPK and NF-κB and Activating HO-1/Nrf2 Pathways. Int J Biol Sci. 2016;12:72–86.
- Ryu H, Lee J, Zaman K, Kubilis J, Ferrante RJ, Ross BD, Neve R, Ratan RR. Sp1 and Sp3 are oxidative stress-inducible, antideath transcription factors in cortical neurons. J Neurosci. 2003;23:3597–606.
- 140. Yeh SH, Yang WB, Gean PW, Hsu CY, Tseng JT, Su TP, Chang WC, Hung JJ. Translational and transcriptional control of Sp1 against ischaemia through a hydrogen peroxide-activated internal ribosomal entry site pathway. Nucleic Acids Res. 2011;39:5412–23.
- 141. Tan NY, Khachigian LM. Sp1 phosphorylation and its regulation of gene transcription. Mol Cell Biol. 2009;29:2483–8.
- 142. Dunah AW, Jeong H, Griffin A, Kim YM, Standaert DG, Hersch SM, Mouradian MM, Young AB, Tanese N, Krainc D. Sp1 and TAFII130 transcriptional activity disrupted in early Huntington's disease. Science. 2002;296:2238–43.
- 143. Liu YW, Tseng HP, Chen LC, Chen BK, Chang WC. Functional cooperation of simian virus 40 promoter factor 1 and CCAAT/enhancer-binding protein beta and delta in lipopolysaccharide-induced gene activation of IL-10 in mouse macrophages. J Immunol. 2003;171:821–8.
- Fridmacher V, Kaltschmidt B, Goudeau B, Ndiaye D, Rossi FM, Pfeiffer J, Kaltschmidt C, Israël A, Mémet S. Forebrain-specific neuronal inhibition of nuclear factor-kappaB activity leads to loss of neuroprotection. J Neurosci. 2003;23:9403–8.
- Meissner M, Stein M, Urbich C, Reisinger K, Suske G, Staels B, Kaufmann R, Gille J. PPARalpha activators inhibit vascular endothelial growth factor receptor-2 expression by repressing Sp1-dependent DNA binding and transactivation. Circ Res. 2004;94:324–32.
- 146. Chu S, Ferro TJ. Sp1: regulation of gene expression by phosphorylation. Gene. 2005;348:1–11.
- 147. Karin M. The regulation of AP-1 activity by mitogen-activated protein kinases. J Biol Chem. 1995;270:16483–6.
- 148. Hess J, Angel P, Schorpp-Kistner M. AP-1 subunits: quarrel and harmony among siblings. J Cell Sci. 2004;117:5965–73.
- Alam J, Den Z. Distal AP-1 binding sites mediate basal level enhancement and TPA induction of the mouse heme oxygenase-1 gene. J Biol Chem. 1992;267:21894–900.
- Wright MM, Kim J, HockTD, Leitinger N, Freeman BA, Agarwal A. Human haem oxygenase-1 induction by nitro-linoleic acid is mediated by cAMP, AP-1 and E-box response element interactions. Biochem J. 2009;422:353–61.
- Deshane J, Kim J, Bolisetty S, Hock TD, Hill-Kapturczak N, Agarwal A. Sp1 regulates chromatin looping between an intronic enhancer and distal promoter of the human heme oxygenase-1 gene in renal cells. J Biol Chem. 2010;285:16476–86.
- 152. Rinne ST, Liu CF, Feemster LC, Collins BF, Bryson CL, O'Riordan TG, Au DH. Thiazolidinediones are associated with a reduced risk of COPD exacerbations. Int J Chron Obstruct Pulmon Dis. 2015;10:1591–7.
- 153. Rinne ST, Feemster LC, Collins BF, Au DH, Perkins M, Bryson CL, O'Riordan TG, Liu CF. Thiazolidinediones and the risk of asthma exacerbation among patients with diabetes: a cohort study. Allergy Asthma Clin Immunol. 2014;10:34.
- 154. Wang MT, Lai JH, Huang YL, Kuo FC, Wang YH, Tsai CL, Tu MY. Use of antidiabetic medications and risk of chronic obstructive pulmonary disease exacerbation requiring hospitalization: a disease risk scorematched nested case-control study. Respir Res. 2020;21:319.
- 155. Chen KY, Wu SM, Tseng CH, Lee KY, Lin YH, Liu HY, Chien LN. Combination therapies with thiazolidinediones are associated with a lower risk of acute exacerbations in new-onset COPD patients with advanced diabetic mellitus: a cohort-based case-control study. BMC Pulm Med. 2021;21:141.
- 156. Kobayashi M, Thomassen MJ, Rambasek T, Bonfield TL, Raychaudhuri B, Malur A, Winkler AR, Barna BP, Goldman SJ, Kavuru MS. An inverse relationship between peroxisome proliferator-activated receptor gamma and allergic airway inflammation in an allergen challenge model. Ann Allergy Asthma Immunol. 2005;95:468–73.
- 157. Spears M, Donnelly I, Jolly L, Brannigan M, Ito K, McSharry C, Lafferty J, Chaudhuri R, Braganza G, Bareille P, et al. Bronchodilatory effect of the PPAR-gamma agonist rosiglitazone in smokers with asthma. Clin Pharmacol Ther. 2009;86:49–53.

- 158. Richards DB, Bareille P, Lindo EL, Quinn D, Farrow SN. Treatment with a peroxisomal proliferator activated receptor gamma agonist has a modest effect in the allergen challenge model in asthma: a randomised controlled trial. Respir Med. 2010;104:668–74.
- 159. Park SJ, Lee KS, Kim SR, Min KH, Choe YH, Moon H, Chae HJ, Yoo WH, Lee YC. Peroxisome proliferator-activated receptor gamma agonist down-regulates IL-17 expression in a murine model of allergic airway inflammation. J Immunol. 2009;183:3259–67.
- 160. Lea S, Plumb J, Metcalfe H, Spicer D, Woodman P, Fox JC, Singh D. The effect of peroxisome proliferator-activated receptor-γ ligands on in vitro and in vivo models of COPD. Eur Respir J. 2014;43:409–20.
- Birrell MA, Patel HJ, McCluskie K, Wong S, Leonard T, Yacoub MH, Belvisi MG. PPAR-gamma agonists as therapy for diseases involving airway neutrophilia. Eur Respir J. 2004;24:18–23.
- Liu D, Zeng BX, Shang Y. Decreased expression of peroxisome proliferator-activated receptor gamma in endotoxin-induced acute lung injury. Physiol Res. 2006;55:291–9.
- 163. Cheng Y, Li S, Wang M, Cheng C, Liu R. Peroxisome Proliferator Activated Receptor gamma (PPARy) Agonist Rosiglitazone Ameliorate Airway Inflammation by Inhibiting Toll-Like Receptor 2 (TLR2)/Nod-Like Receptor with Pyrin Domain Containing 3 (NLRP3) Inflammatory Corpuscle Activation in Asthmatic Mice. Med Sci Monit. 2018;24:9045–53.
- 164. Li M, Li Z, Sun X, Yang L, Fang P, Liu Y, Li W, Xu J, Lu J, Xie M, Zhang D. Heme oxygenase-1/p21WAF1 mediates peroxisome proliferatoractivated receptor-gamma signaling inhibition of proliferation of rat pulmonary artery smooth muscle cells. FEBS J. 2010;277:1543–50.
- Liu D, Zeng BX, Zhang SH, Yao SL. Rosiglitazone, an agonist of peroxisome proliferator-activated receptor gamma, reduces pulmonary inflammatory response in a rat model of endotoxemia. Inflamm Res. 2005;54:464–70.
- 166. Croasdell A, Duffney PF, Kim N, Lacy SH, Sime PJ, Phipps RP. PPARγ and the Innate Immune System Mediate the Resolution of Inflammation. PPAR Res. 2015;2015:549691.
- Lee KS, Park SJ, Hwang PH, Yi HK, Song CH, Chai OH, Kim JS, Lee MK, Lee YC. PPAR-gamma modulates allergic inflammation through up-regulation of PTEN. FASEB J. 2005;19:1033–5.
- 168. Guyton K, Zingarelli B, Ashton S, Teti G, Tempel G, Reilly C, Gilkeson G, Halushka P, Cook J. Peroxisome proliferator-activated receptor-gamma agonists modulate macrophage activation by gram-negative and gram-positive bacterial stimuli. Shock. 2003;20:56–62.
- Wang YX. PPARs: diverse regulators in energy metabolism and metabolic diseases. Cell Res. 2010;20:124–37.
- Liu Y, Wang J, Luo S, Zhan Y, Lu Q. The roles of PPARy and its agonists in autoimmune diseases: A comprehensive review. J Autoimmun. 2020;113:102510.
- 171. Ballesteros I, Cuartero MI, Pradillo JM, de la Parra J, Pérez-Ruiz A, Corbí A, Ricote M, Hamilton JA, Sobrado M, Vivancos J, Nombela F, Lizasoain I, Moro MA. Rosiglitazone-induced CD36 up-regulation resolves inflammation by PPARy and 5-LO-dependent pathways. J Leukoc Biol. 2014;95:587–98.
- Feng H, Yin Y, Zheng R, Kang J. Rosiglitazone ameliorated airway inflammation induced by cigarette smoke via inhibiting the M1 macrophage polarization by activating PPARγ and RXRα. Int Immunopharmacol. 2021;97:107809.
- Zampetaki A, Minamino T, Mitsialis SA, Kourembanas S. Effect of heme oxygenase-1 overexpression in two models of lung inflammation. Exp Biol Med (Maywood). 2003;228:442–6.
- Drummond GS, Baum J, Greenberg M, Lewis D, Abraham NG. HO-1 overexpression and underexpression: Clinical implications. Arch Biochem Biophys. 2019;673:108073.
- Sass G, Seyfried S, Parreira Soares M, Yamashita K, Kaczmarek E, Neuhuber WL, Tiegs G. Cooperative effect of biliverdin and carbon monoxide on survival of mice in immune-mediated liver injury. Hepatology. 2004;40:1128–35.
- 176. Zhang T, Fang Q, Liu P, Wang P, Feng C, Wang J. Heme oxygenase 1 overexpression induces immune evasion of acute myeloid leukemia against natural killer cells by inhibiting CD48. J Transl Med. 2022;20:394.

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