REVIEW

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Polyamines: their significance for maintaining health and contributing to diseases

Mengjuan Xuan^{1†}, Xinyu Gu^{2†}, Juan Li^{1†}, Di Huang³, Chen Xue^{1*} and Yuting He^{4*}



Polyamines are essential for the growth and proliferation of mammalian cells and are intimately involved in biological mechanisms such as DNA replication, RNA transcription, protein synthesis, and post-translational modification. These mechanisms regulate cellular proliferation, differentiation, programmed cell death, and the formation of tumors. Several studies have confirmed the positive effect of polyamines on the maintenance of health, while others have demonstrated that their activity may promote the occurrence and progression of diseases. This review examines a variety of topics, such as polyamine source and metabolism, including metabolism, transport, and the potential impact of polyamines on health and disease. In addition, a brief summary of the effects of oncogenes and signaling pathways on tumor polyamine metabolism is provided.

Keywords Polyamine metabolism, Disease, Cancer, Oncogene, Mechanism

Introduction

Polyamines, which are small polycationic compounds with a positive charge, are found in all types of living cells, including those of mammals, plants, and prokaryotes [1]. It is well known that there are three primary types of polyamines present in mammalian cells: putrescine (Put),

[†]Mengjuan Xuan, Xinyu Gu and Juan Li contributed equally.

*Correspondence: Chen Xue xchenzzu@163.com Yuting He fccheyt1@zzu.edu.cn ¹ Department of Infectious Disease, The First Affiliated Hospital of Zhengzhou University, No. 1 Jianshe East Road, Erqi District, Zhengzhou 450052, Henan, China ² Department of Oncology, College of Clinical Medicine, The First Affiliated Hospital, Henan University of Science and Technology, Luoyang 471000, Henan, China

³ Department of Child Health Care, The Third Affiliated Hospital

of Zhengzhou University, Zhengzhou 450000, Henan, China

⁴ Department of Hepatobiliary and Pancreatic Surgery, The First Affiliated Hospital of Zhengzhou University, Zhengzhou 450052, Henan, China spermidine (Spd), and spermine (Spm). The discovery of polyamines, specifically spermidine and spermine, dates back to 1678 when Antonie van Leeuwenhoek made the initial observation [2]. Throughout the twentieth century, their structure was characterized and their biosynthetic pathway was determined as a result of ongoing research [3]. Although all eukaryotic cells are capable of synthesizing the three major polyamines, polyamines can also be obtained through diet and other alternative sources [4], including intestinal bacteria. In addition, polyamines are essential for the growth and proliferation of mammalian cells and are intricately linked to biological processes, such as replication, transcription, translation, and posttranslational modification, which modulate cellular proliferation, differentiation, apoptosis, and tumorigenesis [4, 5]. Briefly, polyamines within mammalian cells are irreplaceable, as the depletion of polyamines will completely halt cell growth and proliferation.

Numerous empirical studies support the claim that spermidine, the most abundant polyamine in mammals, acts as an elixir of life [6, 7]. Previous studies have



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demonstrated that polyamine concentrations tend to decrease with age, demonstrating a negative correlation with advancing age [8, 9]. One study, for instance, elucidated the relationship between polyamine levels and age, revealing that individuals aged 31-56 years and over 90 years have significantly lower total polyamine concentrations than those aged 60-80 years and older. Surprisingly, however, the concentrations of spermidine and spermine in the aforementioned two age groups are greater than in those aged 60 to 80 years, and individuals aged over 90 years have the highest relative proportion of spermine among these three age groups [10]. In this experiment, the relationship between polyamine concentration and age differed from the above statement. There were two primary factors that contributed to the observed results, namely, a small sample size of only 78 individuals, and the discrepant levels of polyamines in plasma as compared with those found in tissue [11]. Additionally, the administration of Spm has been demonstrated to markedly increase the lifespan of several model organisms (such as yeast, fruit flies, worms, and etc.), which suggests that spermidine may be a promising agent for enhancing health and longevity [8, 12].

It has been experimentally demonstrated that polyamines exert a crucial function in ensuring the survival of cells and promoting their growth [13, 14]. Consequently, the regulation of polyamine concentration and activity is of the utmost importance and is achieved via a complex network of pathways. The regulation of intracellular polyamine levels and activities is governed by pathways involved in polyamine synthesis, degradation, and transport, which have been extensively studied in bacteria, yeast, and higher organisms [15]. Consequently, fluctuations in polyamine concentrations and disruptions in polyamine metabolism will inevitably have negative effects on overall health, possibly even initiating the occurrence and progression of diseases, such as acute kidney injury, Alzheimer's disease, diabetes, etc. [16–18]. Furthermore, it is essential to acknowledge that numerous studies have elucidated notable alterations in the quantities and functions of polyamines in cancer [19, 20].

The primary objective of the present review is to examine the source and metabolic mechanism of polyamines in humans, investigate the functions and mechanisms of polyamines in both non-neoplastic and neoplastic diseases, and ultimately provide potential insights for the treatment of said ailments.

The source and metabolism of polyamine

Polyamine biosynthesis

The biosynthesis pathway of mammalian polyamine is depicted in Fig. 1 as a series of enzymatic reactions that lead to the production of these important intracellular components. Numerous microorganisms and higher plants have the ability to synthesize putrescine from agmatine via decarboxylation of arginine (ADC) [21, 22]. Despite the presence of evidence for ADC in mammals, Coleman et al. observed that ADC within mitochondrial extracts lack catalytic activity [23]. However, Wang et al. has indicated that the deficiency of ornithine decarboxylase (ODC) in ovine conceptus trophectoderm would ultimately result in an elevation of the activity of ADC [24]. Accordingly, the prevailing pathway for the de novo biosynthesis of putrescine in these species is via the enzyme ODC [25]. ODC is a pyridoxal phosphate-

dependent enzyme and is frequently thought to be the

rate-limiting factor in modulating the concentration of

Put [26]. Ornithine, which is obtainable from the plasma

and can also be generated intracellularly through argin-

ase (Arg 1) activity, serves as the substrate for the synthe-

sis of Put. Thus, it is possible that arginase, which is more

widely distributed than other urea cycle enzymes, is pre-

sent in extrahepatic tissues in order to ensure the availa-

bility of ornithine for polyamine synthesis. Arginase can,

therefore, be regarded as the first step in the polyamine

biosynthetic pathway [27]. A group of aminopropyl-transferases, specifically spermidine synthase (SRM) and spermine synthase (SMS), catalyzes the conversion of putrescine to spermidine and spermine during the synthesis of higher polyamines [28]. The aforementioned aminopropyl group originates from methionine and undergoes a two-step process consisting of conversion to S-adenosylmethionine and decarboxylation [27]. Subsequently, the decarboxylated S-adenosylmethionine (dcAdoMet) serves as an aminopropyl donor in a manner comparable to the use of S-adenosylmethionine as a methyl donor. Despite a close similarity in their responses, human SRM and SMS are discrete enzymes and exhibit stringent selectivity for their respective substrates [29]. Human SRM, a dimer comprised of two identical subunits, encompasses an active site within its C-terminal domain that shares a structural similarity with that of SMS [30]. Although the configurations of their active sites are comparable, they differ primarily in the constricted spatial allowance for the amine substrate within the framework of SRM, which guarantees putrescine is the only substrate for SRM [5].

During the synthesis of spermidine and spermine, the aminopropyl donor dcAdoMet produces the secondary metabolite 5'-methylthioadenosine (5'-MTA) [19]. In the methionine salvage pathway, MTA is initially phosphorylated by 5'-methylthioadenosine-phosphorylase (MTAP), after which it is converted into adenine (one of the primary components of ATP) and 5-methylthior-ibose-1-phosphate. Methionine metabolism involves the conversion of 5-methylthioribose-1-phosphate into a

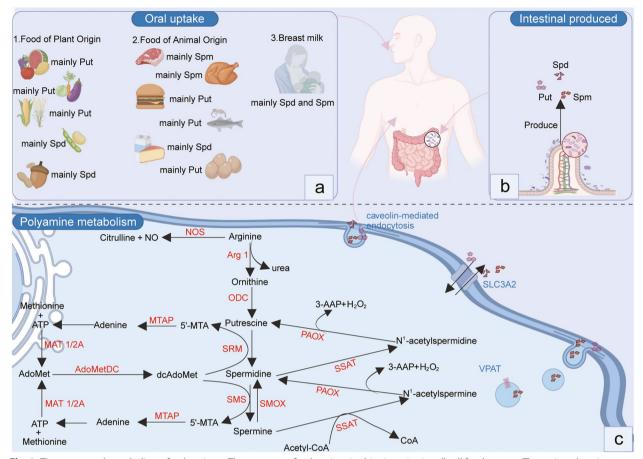


Fig. 1 The source and metabolism of polyamine. **a** The presence of polyamines is ubiquitous in virtually all food sources. The main polyamines in breast milk are spermidine and spermidine, and the content and distribution of polyamines in plant-derived and animal-derived foods differ based on their respective categorizations. **b** The intestinal microbiota has the capability to produce polyamines. **c** The biosynthesis of polyamines commences with the conversion of L-ornithine into putrescine via the action of ornithine decarboxylase (ODC). Subsequently, the addition of an aminopropyl group, which is contributed by dcAdoMet, results in the formation of spermidine and spermine. In the process of polyamine catabolism, the enzyme SSAT are responsible for the acetylation of spermine and spermidine, resulting in the production of N1-acetylspermine and N1-acetylspermidine. SMOX can oxidize spermine directly to spermidine These metabolites are either secreted from cells or undergo reconversion back into spermidine and putrescine via the enzyme PAOX. Created with BioRender.com

metabolite that can serve as a substrate for AdoMet synthesis through a cascade of enzymatic reactions. This is facilitated by methionine adenosyl-transferase 2 (MAT2), which catalyzes the final step of AdoMet synthesis by facilitating the ATP-dependent transfer of an adenosyl group to methionine. MTAP deficiency results in the inability of endogenous MTA to salvage methionine or adenine, resulting in impaired polyamine biosynthesis and the accumulation of dcAdoMet and 5'-MTA [31].

Polyamine catabolism

It is generally accepted that the biochemical reactions catalyzed by spermidine synthase and spermine synthase are effectively irreversible. However, it has been established for many years that the transformation of spermine into spermidine and spermidine into putrescine can occur in vivo [28, 32, 33]. Spermine oxidase (SMOX) catalyzes the conversion of spermine to spermidine, 3-aminopropanaldehyde, and H_2O_2 , with negligible effect on spermidine [33]. On the other hand, the enzyme acetylpolyamine oxidase (PAOX) catalyzes the transformation of N1-acetylspermidine into putrescine and N-acetyl-3-aminopropanaldehyde [34–37]. PAOX is also highly efficient at converting N1-acetylspermine to spermidine. The substrates required for PAOX-mediated reactions are produced through the catalytic activity of spermidine/spermine-N1-acetyltransferase (SSAT) [38]. The enzyme SSAT, encoded by the gene SAT1, typically exists at minimal concentrations but can be easily stimulated by increased levels of free polyamines [38]. SSAT facilitates the formation of N1-acetylspermine and N1-acetylspermidine, which may be extruded from the cellular environment or converted into 3-acetylaminopropanal, H_2O_2 , and either Spd or Put, through the action of PAOX. The products generated via the reactions SMOX and PAOX, as well as acrolein, a toxic byproduct of spontaneous deamination of 3-aminopropanal [39] and Spm oxidation produced following cellular injury by copper-dependent oxidases present extracellularly [40], have been associated with renal insufficiency [41], ischemia–reperfusion [42] and injury cerebral infarction [43].

Polyamine uptake

Aside from intracellular biosynthesis, the systemic availability of polyamines also depends on two other essential sources: extrinsic (oral) absorption via dietary intake and synthesis by intestinal microflora (Fig. 1) [44]. Surprisingly, it is believed that food consumption and endogenous production by intestinal microbiota are the primary sources of polyamines in healthy adult humans [45-47]. Following absorption by the gastrointestinal tract, polyamines are rapidly assimilated [48], resulting in a rapid elevation of portal vein concentrations and subsequent dispersion throughout the entire spectrum of organs and tissues [49]. Polyamines are ubiquitously present in all food types, such as shellfish with elevated polyamine concentrations per unit of caloric intake, albeit with substantial variation in their concentrations [50]. Given the abundance of polyamine sources, it appears that any effort to reduce the concentration of polyamines in order to promote health or alleviate disease must involve the elimination of all polyamine sources. Otherwise, the residual sources of polyamines could compensate for the maintenance of optimal polyamine levels [45].

Polyamine transport

Due to the protonation of the primary and secondary amino groups of polyamines at the physiological pH of the extracellular environment [51], polyamine is conceptually incapable of diffusing passively through cellular membranes. While there exists a certain degree of comprehension regarding the polyamine transport, further researches are required to substantiate the precise transport mechanism and the transportation carriers involved. There have been three postulated models regarding the transport of polyamine, which include glypican-mediated endocytosis, plasma transport and vesicular sequestration, and caveolin-mediated endocytosis [52]. The glypican-1 molecule possesses a side chain consisting of heparan sulfate that can bind with Spm and facilitate its transport into the cytoplasm [53]. It has been hypothesized that polyamine transport may also involve transporters on the plasma membrane, which are similar to those found in yeast and bacteria [54]. Solute carrier (SLC) transporters and ATP transporters (such as ATP13A2 and ATP13A3) serve as a mediator for the transport of polyamine in mammalian cells [52, 55-57]. For instance, SLC family 3 member 2 (SLC3A2) facilitates the export of acetylated polyamines through a reaction involving the exchange of polyamines with arginine [58]. In addition, the finding of a vesicular polyamine transporter (VPAT, also known as SLC18B1) in astrocytes [59] and mast cells [60] highlights the potential neuro- and immune-modulatory effects of this transport mechanism [60, 61]. Moreover, the activated K-RAS modulates polyamine transport in colon cancer cells by regulating the expression of caveolin-1, which exerts a suppressive effect on caveolar endocytosis [62]. Therefore, the accurate delineation of the polyamine transport system will generate novel perspectives regarding the management and prognosis of certain diseases.

The effect and mechanism of polyamine on disease

Growing evidence suggests that polyamines exert a substantial effect on the pathogenesis and progression of numerous diseases. In this regard, it has also been established that polyamines have diagnostic value in certain conditions, including cardiovascular disease [63, 64] and metabolic syndromes [65, 66]. Furthermore, their prognostic potential in predicting disease outcomes has also been demonstrated. Tables 1 and 2 provide extensive data on the properties and potential biological functions of polyamines in both neoplastic and non-neoplastic diseases.

Non-neoplastic diseases

Polyamines in cardiovascular protection

Polyamines are essential to the angiogenesis process, which occurs in response to tissue damage or tumor growth [4]. It has been demonstrated that the inhibition of polyamine synthesis inhibits angiogenesis in both gastric ulceration models [119] and tumor models [100, 120].

The process of aging induces two critical alterations in arteries, which markedly elevate the susceptibility to cardiovascular diseases (CVD): 1. The rigidity of the major elastic arteries (viz. the aorta and carotid arteries) and 2. The emergence of vascular endothelial dysfunction [121, 122]. Arterial stiffening results from age-related changes in the arterial wall, including increased collagen deposition, decreased elastin content, and cross-linking of these and other structural proteins via the formation of advanced glycation end products (AGEs) [123]. Agerelated vascular endothelial dysfunction is primarily the result of decreased nitric oxide (NO) bioavailability, as indicated by impaired endothelium-dependent dilation

Neoplasm type	Target gene	Regulation	Expression	Associated protein	Inhibitor	Concentration/ Activity	Polyamine level	Function	Refs
OC	MYC	positive	High	ODC	DFMO	Ļ	Ļ	inducing apoptosis through regula- tion of AP-1 by DFMO	[67]
	/	/	/	ODC	DEMO + aza	Ļ	Ļ	Increasing anti-tumor M1 macrophages through DFMO and aza	[68]
	/	/	/	ODC	DFMO	Ļ	Ļ	enhancing the cytotoxicity of PARP inhibi- tors via DFMO	[69]
	/	/	/	ODC	SBP-101	\downarrow	\downarrow	suppressing OC via SBP-101	[70]
HCC	MAT1A	positive	Low	MATI/III	/	↓	\downarrow	increasing HCC cell proliferation	[71–73]
	MAT2A	positive	High	MATII	/	↑	↑	activating MAT2A in turn	[74]
	/	/	/	GSK-3β	SSAT	Ļ	Ļ	suppressing tumor progres- sion and metas- tasis	[75]
	/	/	/	HDAC4	Spd	\downarrow	↑	enhancing autophagy flux	[76]
GC	SMO	positive	High	SMO	/	Ŷ	Ļ	promotes H. pylori-induced carcinogenesis	[77]
	/	/	/	SMO	/	Ŷ	Ļ	increasing gastric cancer risk by H. pylori-induced overexpression of SMO	[78]
	CagA	positive	High	SMO	/	î	Ļ	increasing malignant transformation through CagA	[79]
	AMD1	positive	High	AMD1	SAM486A	Ļ	Ļ	suppressing tumor growth by inhibiting AMD1	[80]
CRC	MAT2A	positive	High	MATII	/	1	↑	activating MAT2A in turn	[74]
	MYC	positive	High	ODC	/	î	↑	activating ODC through mutant APC	[81–83]
	APC	positive	High	ODC	DFMO	Ļ	Ļ	suppressing colorectal carcinogenesis by DFMO	[83]
	/	/	/	GSK-3β	SSAT	Ļ	Ļ	suppressing tumor progres- sion and metas- tasis	[75]
	K-RAS	negative	High	Caveolin-1	/	Ļ	Ŷ	boosting poly- amine uptake via caveolin-1 phosphorylation	[84]

Table 1 The regulation and function of polyamine in carcinogenesis

Neoplasm type	Target gene	Regulation	Expression	Associated protein	Inhibitor	Concentration/ Activity	Polyamine level	Function	Refs
PDAC	/	/	/	EP300	Spd	Ļ	↑	inhibiting EP300 via Spd	[85–87]
	MTAP	negative	Low	ODC	/	î	↑	increasing ODC through the loss of MTAP	[88, 89]
PCa	/	/	/	MTAP; SSAT	MTDIA + BENSpm	↓;↑	Ļ	Delaying or pre- venting PCa recurrence	[90]
	PTEN	negative	Low	AdoMetDC	/	↑	↑	promoting tumor growth	[91]
	MYC	positive	Low	ODC	PGC1-a	\downarrow	\downarrow	suppressing PCa	[92]
Neuroblas- toma	MYCN	positive	High	ODC	DFMO	Ļ	Î	inhibiting MYCN-induced Neuroblas- toma initiation and progression by DFMO	[93–96]
	MYCN	positive	High	SLC3A2	/	Ŷ	↑	increasing poly- amine synthesis by regulating SLC3A2	[97]
	MYC	positive	High	ODC	DFMO + AMXT- 1501	Ļ	Ļ	emphasizing the necessity of polyamines in Neuroblas- toma	[98]
	MYC	positive	Low	ODC	/	Ļ	Ļ	inhibiting poly- amine metabo- lism by glucose deprivation	[99]

(EDD) mediated by NO [124, 125]. LaRocca et al., suggested that spermidine has a remarkable anti-aging effect on arteries, which is due to its ability to increase the bioavailability of nitric oxide, reduce oxidative stress, alter structural factors, and enhance autophagy [101]. In addition, Eisenberg et al., concurred that spermidine administration promoted myocardial autophagy, mitophagy, and mitochondrial respiration, while simultaneously enhancing the mechanical and elastic properties of cardiomyocytes in vivo, which was accompanied by elevated titin phosphorylation and the suppression of subclinical inflammation [63]. Following the experiments conducted by Eisenberg et al., numerous researchers have endeavored to determine the precise mechanism by which spermidine promotes cardiac protection. As evident from the studies conducted by Yan et al., increasing autophagy via Spd-mediated targeting of the AMPK/ mTOR signaling pathway improves cardiac dysfunction after myocardial infarction [104]. Notably, the aforementioned conclusions were all derived from in vivo experiments conducted on mice. Holbert et al. definitively highlighted that bovine serum amine oxidase in vitro experimentation can catalyze the oxidation of exogenous polyamines and subsequently trigger the release of reactive oxygen species (ROS), leading to the induction of autophagy [126]. Therefore, it is challenging to establish the particular mechanism of polyamines, particularly Spd, in facilitating cardiac protection. In 2020, Wang et al., conducted a study proving that Spd stimulates mitochondrial biogenesis [105]. The protective effect of Spd on the cardiovascular system has been confirmed in quite a few studies [102, 103]. This activation occurred as a result of SIRT1-mediated deacetylation of PGC-1 α , which subsequently led to the alleviation of cardiac aging. Multiple other studies have verified the cardioprotective properties of spermidine on the cardiovascular system [127–129]. Therefore, spermidine is a potentially effective nutraceutical intervention for the mitigation of arterial aging and the prevention of age-related cardiovascular diseases (Fig. 2).

Polyamines in alzheimer's disease

Alzheimer's disease (AD) is recognized as the leading cause of cognitive dysfunction or dementia in individuals

Non-neoplastic Diseases	Organism	Tissue	Associated protein/ cytokine	Function	Treatment	Concentration/ Activity	Effect	Ref(s)
Circulatory system	mice	vessel in gastric cancer	ODC	biosynthesis of polyamine	DFMO	Ļ	decreasing the vessel density	[100]
	mice	artery	ODC	biosynthesis of polyamine	extracellular spermidine	Ļ	increasing NO bioavail- ability	[101, 102]
	mice	heart	TNF-a	inflammation of cardiomyo- cytes	spermidine	Ļ	decreasing the passive stiff- ness of cardio- myocytes	[63, 103]
	rats	heart	АМРК	activation of autophagy	/	Ŷ	promoting autophagy by Spd-acti- vated AMPK	[104]
	rats	heart	SIRT1	biogenesis of mitochon- drion	/	Ŷ	improving mitochondrial biogenesis by Spd	[105]
Alzheimer's disease	mice	cerebrum	Arg 1; ODC	biosynthesis of polyamine	/	↑	inducing memory loss	[106, 107]
Asthma	mice	lung	SSAT; SMO	catabolism of polyamine	MDL72.527	\downarrow	airway epithelial injury	[108]
	mice	lung	ODC	biosynthesis of polyamine	/	↑	airway hyperre- sponsiveness	[109]
Obesity	mice	WAT	SSAT	catabolism of polyamine	/	↑	increasing lipid oxidation	[110–112]
	drosophila	abdomen	SPDSY; SPMSY	biosynthesis of polyamine	/	/	regulating triglyceride storage	[113]
	mice	WAT	SSAT	catabolism of polyamine	/	↑	decreasing fatty acid synthesis	[114]
Pancreatitis	rats	pancreas	SSAT	catabolism of polyamine	/	↑	increasing trypsinogen activation	[115–117]
Psoriasis	mice	skin	Arg 1	biosynthesis of polyamine	nor-NOHA	Ļ	inhibiting self- RNA sensation within keratino- cytes	[118]

Table 2 The regulation and Function of polyamine in non-neoplastic diseases

aged 65 and older and is rapidly emerging as one of the most expensive, lethal, and burdensome diseases of the twenty-first century [130, 131]. Alzheimer's disease is characterized by the deposition of beta-amyloid (A β) plaques and subsequent memory loss [132, 133]. The initial neuronal impairment in Alzheimer's disease patients typically occurs in regions of the brain responsible for cognitive processes such as memory, language, and thought. This, in turn, results in the emergence of early symptoms that typically involve difficulties in these particular cognitive domains [134, 135].

Considering the close association between the genes expressed by glial cells and AD, genetic studies suggest that AD research should shift its focus from neurons to glial cells and neuroinflammation [136]. Evidence from prior studies suggests that Put has the ability to produce GABA via the enzyme MAO-B (monoamine oxidase B) in astrocytes, which may pose a risk for memory impairment [137, 138]. In addition, a recent study demonstrates that the administration of β -amyloid induces a noteworthy upregulation of Put and GABA expression within astrocytes through the activation of specific intracellular signaling cascades [137]. Arg 1 and ODC are enzymes that catalyze the putrescine synthesis process, and inhibiting their activity would lead to a significant reduction in the production of putrescine. Therefore, inhibition of specific enzymes represents a potential therapeutic avenue for ameliorating cognitive

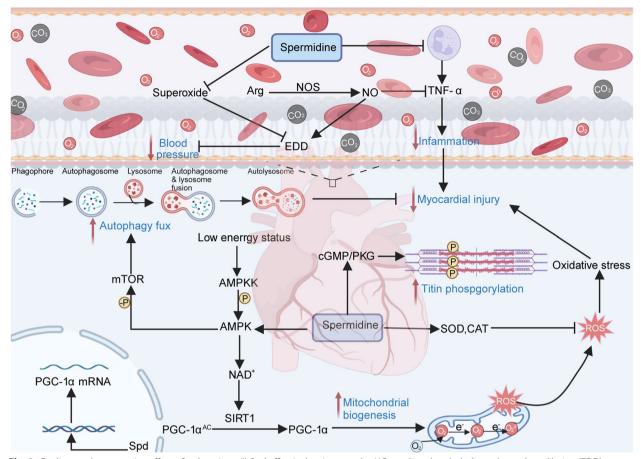


Fig. 2 Cardiovascular protective effect of polyamines. (I) Spd effectively reinstates the NO-mediated endothelium-dependent dilation (EDD) by augmenting the level of NO bioavailability and abating oxidative stress, therefore reducing blood pressure. (II). Spd increases autophagy flux through the AMPK-mTOR signaling pathway, contributing to the stimulation of cardiomyocyte protective autophagy. (III). The structure and function of cardiomyocytes can be improved through Titin phosphorylation and mitochondria formation. And Spd can activate mitochondrial biogenesis through SIRT1-mediated PGC-1α deacetylation. (IV). Spd effectively suppresses the release of TNF-α by immune cells, consequently lowering levels of subclinical chronic inflammation and ultimately preventing the onset of myocardial injury. Created with BioRender.com

impairments and potentially reducing β -amyloid plaque burdens [106, 107].

Polyamines in asthma

Asthma is a respiratory disorder characterized by intermittent bronchospasm, which results in dyspnea and wheezing [139, 140]. It is characterized by airway inflammation, airway hyperresponsiveness, and mucus hypersecretion, all of which contribute to variable airflow obstruction. The heterogeneous underlying inflammatory mechanisms in asthma further add complexity to the disease [140]. In this regard, elevated polyamine concentrations have been observed in the circulation of individuals experiencing asthma attacks, in addition to significantly elevated polyamine concentrations in bronchoalveolar lavage fluid collected from asthmatic patients [109, 141, 142].

The pathologic characteristics of asthma are intimately connected to the pathologic reactions and morphologic alterations of inflamed and structural cells. Moreover, the interaction between polyamines and immune and structural cells in the context of asthmatic pathophysiology implicates the polyamine pathway as having a beneficial effect on asthmatic responses [143]. SSAT and SMOX play significant roles in the process of polyamine catabolism [33, 38]. Knocking-down of the genes of these enzymes results in a decrease in polyamine catabolism, ultimately leading to an increase in the concentration of spermidine and spermine which will result in the induction of stress and apoptosis in bronchial epithelial cells [108]. In addition, research has shown that upregulation of Arg1 expression results in an increase in polyamine synthesis, ultimately leading to the development of airway hyperresponsiveness, which may be partially attributed to the inhibiting effect of Arg1 overexpression

on NO synthesis [109]. Inflammatory cells can release a number of inflammatory factors that result in trachea spasms and severe asthma attacks [144]. Furthermore, it has been discovered that polyamines enhance the disease-causing capacity of cells involved in inflammatory responses, specifically mast cells and granulocytes such as eosinophils and neutrophils, by stimulating the liberation of their pro-inflammatory agents or extending their lifespan [143, 145, 146]. The underlying mechanisms by which polyamine contributes to the onset and progression of asthma have the potential to facilitate the development of therapeutic interventions.

Polyamines in obesity

Obesity constitutes a hazard in relation to a multitude of ailments encompassing metabolic morbidities (such as type 2 diabetes and hepatic steatosis), cardiovascular infirmities (such as coronary heart disease and cerebrovascular accidents), neoplastic disorders (such as hepatocellular and renal cell carcinoma), depressive illnesses, and other debilitating conditions, all of which have a negative effect on the health and lifespan of individuals [147–149]. The majority of patients may not benefit from traditional and invasive therapeutic approaches [150]. Polyamine exerts an extensive role in energy metabolism, particularly in regulating lipid metabolism, consequently representing a pivotal area of focus in the exploration of obesity and its associated ailments.Numerous studies have investigated the relationship between polyamines and energy metabolism, as well as the effects of variations in the enzymes involved in polyamine metabolism on body mass. The mechanism by which Spm exerts its anti-obesity effects was elucidated in the present study by Nakatani et al. [151]. In particular, Spm promotes β -oxidation [110–112] and suppresses pre-adipocyte differentiation, thereby reducing lipid accumulation [151]. The article by Sadasivan et al., demonstrated the significant efficacy of intraperitoneal administration of Spm in mice fed a high-fat diet, as evidenced by a 24% decrease in body weight and a 57% decrease in white adipose tissue (WAT) in comparison to the untreated control group [112]. A significant negative association between spermidine consumption and obesity was demonstrated by the findings of an epidemiological study [152]. Additionally, it was discovered that the administration of spermidine may lead to varying degrees of improvement in the weight status of mice whose obesity was induced by a high-fat diet [153, 154]. In addition, enzymes involved in the metabolism of polyamines regulate the storage site of triglycerides [113] and inhibit fatty acid synthesis [114]. The discoveries presented herein offer fresh perspectives on the pharmacodynamics of polyamine, and its suitability as a curative modality for obesity management.

Polyamines in pancreatitis

Acute pancreatitis is a prevalent inflammatory disorder of the exocrine gland, the pancreas, which manifests as severe abdominal pain and multi-organ dysfunction [155]. It can precipitate pancreatic tissue death [156] and persistent systemic dysfunction, resulting in a 1–5% mortality rate [155]. Due to the severe complications and increased mortality associated with pancreatitis, as well as the fact that spermidine is found in the highest concentration in pancreatic tissue [157] and polyamines are involved in the processes of cell proliferation and apoptosis [13, 14], it seems plausible that the deficiency of polyamines in individuals may serve as a catalyst for the onset and progression of pancreatitis.

It is essential to emphasize that SSAT is a key enzyme in polyamine catabolism, facilitating the acetylation of spermidine and spermine by transferring acetyl groups from CoA to produce acetylpolyamines [38]. In the previous two experiments, zinc administration was used to induce pancreatitis in transgenic rats genetically modified to overexpress the SSAT gene under the control of the inducible mouse metallothionein I promoter [115, 116]. Both of the aforementioned studies came to the conclusion that the catabolism of polyamines caused pancreatitis. The disagreement between the two groups of researchers relates to their divergent perspectives regarding the precise mechanism of pancreatitis. The former group hypothesized that polyamines function as protease inhibitors and demonstrated that SSAT activated trypsinogen while treatment with polyamine analogs prevented trypsinogen activation [115]. In contrast, the latter group hypothesized that H_2O_2 (a metabolite of acetylated polyamine) caused pancreatitis, but they disproved this theory by administering PAOX inhibitors, which claimed the inflammatory process was unrelated to the production of H_2O_2 [116]. The administration of polyamines has been shown to prevent pancreatitis, and the supplementation of polyamine analogs has also been observed to alleviate complications associated with severe pancreatitis [115–117, 158].

Polyamines on psoriasis

Psoriasis, a persistent and chronic inflammatory condition, has been discovered to exhibit a state of heightened arginine metabolism and exaggerated production of polyamines [118]. In particular, the absence of protein phosphatase 6 (PP6) hinders the maintenance of skin homeostasis, and this phenomenon has been shown to be associated with increased transcription of ARG1 via phosphorylation and activation of the C/EBP- β transcription factor, as well as an increased production of polyamines [118]. It is believed that the addition of cationic polyamines, which have a propensity for binding with nucleic acids, accelerates the internalization of self-RNA by psoriatic keratinocytes and the recognition of self-RNA by myeloid dendritic cells [118]. Significantly, the improvement of this inflammatory condition has been demonstrated through the restoration of the urea cycle with the nor-NOHA arginase inhibitor or administering DFMO as a treatment, offering a novel therapeutic approach for psoriasis [17].

Neoplastic diseases

Ovarian cancer

According to ovarian cancer statistics in the United States in 2022, ovarian carcinoma (OC) comprises the leading cause of death among reproductive-related malignancies, with a prevalence of ~ 20,000 documented cases and ~ 13,000 fatalities per year [159]. More than 70% of OC patients are diagnosed at advanced stages (III–IV), placing ovarian cancer as the fifth leading cause of cancer-related deaths among women in developed nations [160]. Thus, the involvement of polyamines in the timely detection and subsequent management of ovarian cancer demonstrates great promise and potential.

Elevated levels of putrescine, spermidine, and spermine have been found in the urine of individuals with OC [161, 162]. Correspondingly, they have been utilized as biomarkers for the early detection of ovarian cancer [163, 164]. In addition, Fahrmann et al., presented findings of a distinct pattern of polyamines identified in blood samples, which holds significant potential for the early detection of OC [165]. In addition, this polyamine signature serves as a valuable complement to CA125 (human mucin 16, known as the most optimal marker for the early detection of OC) in identifying a greater number of ovarian cancer cases that would have been missed by CA125 alone [165, 166]. Given the propensity of polyamine to promote the proliferation and invasion of tumors, reducing its activity and concentration would be a crucial therapeutic intervention. As an inhibitor of the ratelimiting enzyme ODC in polyamine biosynthesis, DFMO (also known as α -difluoromethylornithine) is reported to induce apoptosis by regulating AP-1 signaling through JNK phosphorylation [67]. In addition, DFMO has demonstrated the ability to enhance the anticancer potential of co-administered pharmaceutical agents, such as 5-azacytidine [68] and poly (ADP-ribose) polymerase (PARP) inhibitor [69]. The co-administration of DFMO and 5-azacytidine upregulates M1 macrophage, resulting in a marked improvement in the survival rate of the combined therapeutic modality. In addition, the concomitant utilization of DFMO and inhibitors of PARP confers not only enhanced susceptibility of cancer cells to PARP inhibitors, but also increased cytotoxicity of the cisplatin. Moreover, the polyamine analog SBP-101(diethyl dihydroxyhomospermine) inhibits tumor cells by decreasing the activity of ODC [70]. In a word, the comprehensive investigation into the level and metabolism of polyamine will bring about new insights in diagnosis and treatment of OC.

Hepatocellular carcinoma

Primary hepatic carcinoid tumor, which predominantly results from hepatocellular carcinoma (HCC), imposes a severe economic and health burden on society, especially in China and other Asian countries/regions due to the high prevalence of chronic hepatitis B virus (HBV) infection [167]. HCC is a highly prevalent malignancy that ranks as the fourth leading cause of cancer-related mortality and the second leading cause of years of life lost due to cancer worldwide [168–170]. However, there is a lack of consensus regarding the function and mechanism of polyamines within the hepatic tissue and adjacent normal tissues of HCC-afflicted individuals. Thus, further investigation is necessary to delve more comprehensively in greater depth.

Polyamines exert regulatory control over the progression of the cell cycle and inhibit polyamine biosynthesis, resulting in cell cycle arrest and apoptosis [171], as previously described. A sudden increase in polyamine biosynthesis could potentially promote the rapid proliferation of preneoplastic and neoplastic hepatocytes [71-73]. During the progression of hepatocarcinogenesis in rats, there is a gradual increase in the expression of the ODC gene, as well as a concomitant elevation in ODC activity and polyamine synthesis [172, 173]. A similar upregulation of genes involved in polyamine synthesis is also observed in human hepatocellular carcinoma [174]. Frau et al., supported that SAM, acting as the principal supplier of methyl groups and a precursor for the synthesis of polyamines, interferes with polyamine synthesis via inhibition of ODC activity, which is partially dependent on the accumulation of 5'-MTA [71]. However, Lauda et al., demonstrated that increased expression of MAT2A, which encodes for the enzymatic component of MAT that catalyzes the formation of SAM [175], leads to more SAM for polyamine biosynthesis. The elevation in polyamine, specifically putrescine in this instance, resulted in a upregulation of mRNA expression levels for MAT2A and ODC, consequently augmenting the proliferative potential of the neoplastic cell [74]. Strikingly, in mice exposed to a potent chemical carcinogen, the administration of spermidine has been found to inhibit the development of HCC [76]. The selective aggregation of truncated microtubule-associated protein 1S (MAP1S) in response to mitotic arrest results in the disruption of mitochondrial function on the mitotic spindle, ultimately leading in mitotic cell death [176]. Accordingly, MAP1S has been found to interact with HDAC4 via a designated domain, and inhibiting HDAC4 results in increased acetylation and enhanced stability of MAP1S [177]. Spermidine enhances the acetylation and stability of MAP1S while also inducing autophagy flux via the depletion of cytosolic HDAC4, thereby reducing the association between MAP1S and HDAC4 [76]. Thus, the characterization of the autophagic flux induced by MAPS1 in mitigating liver cancer in murine models is likely to provide a novel strategy for harnessing MAP1S-triggered autophagic flux for the prevention, delay, or treatment of HCC [76]. In addition, Wang et al. discovered that the β-catenin pathway, which facilitated HCC cells growth and metastasis, can be inhibited through the upregulation of SSAT. This upregulation was strongly associated with a decrease in polyamines, leading to the inhibition of β-catenin translocation into the nucleus [75].

Gastric cancer

According to current statistics, gastric cancer (GC) stands as the fifth most prevalent form of cancer worldwide and the fourth primary mortality-inducing neoplasm [178, 179]. Significant regional disparities exist in the prevalence of GC, with notable occurrences in East Asian countries such as Japan and Northern Europe [179]. In addition, there is growing concern regarding the escalating incidence of GC among those under 50 years old [180]. Prior studies have established a link between polyamine metabolism and the infection of gastric epithelial cells, as well as the increased risk of GC mediated by *Helicobacter pylori* (Hp).

Spermine Oxidase (SMOX) is a crucial enzyme in the metabolism of polyamines in the gastrointestinal tract due to its efficient regulation of spermidine and spermine concentration, which is necessary for maintaining proper physiological function [181]. Numerous studies have elucidated that SMOX can trigger the occurrence of GC by instigating DNA damage [78, 182, 183]. The precise mechanism underlying DNA damage caused by SMOX in Hp-associated gastric cancer requires further elucidation in scholarly literature. The activation of the β -catenin oncogenic signaling pathway in gastric epithelial cells stimulated by Hp is facilitated by the mediator SMOX, as reported by Sierra et al. [77]. As indicated in 2011, there is also the opinion that the pathogenesis of Hp-induced gastric carcinogenesis is associated with the oncoprotein cytotoxin-associated gene A (CagA) of microbiological origin [79]. AMD1, also known as S-adenosylmethionine decarboxylase (AdoMetDC), is one of the key enzymes of polyamine biosynthesis that has a tumorigenic effect on human gastric cancer and affects patient prognosis [80]. In conclusion, it is evident that polyamines play a complex and sophisticated regulatory role in the occurrence and mechanism of Hp-induced GC. The investigation of the regulatory pathway of polyamines in the pathogenesis of gastric cancer and the search for novel polyamine inhibitors have the potential to provide a ray of hope for patients afflicted with this malignancy.

Colorectal cancer

In the United States, colorectal cancer (CRC) is the second leading cause of cancer-related deaths and the third most prevalent malignancy worldwide [184]. Notably, approximately a quarter of individuals diagnosed with CRC exhibit synchronous liver metastases, and between 50 and 75% of patients experience liver metastases within a three-year timeframe following initial colon surgery [185]. Despite some advancements in early detection and the use of modern surgical techniques in conjunction with radiotherapy and chemotherapy, the prognosis for CRC patients remains dismal to this day [186]. Therefore, it is crucial to identify the putative molecular mechanisms that drive the progression and metastasis of CRC. The WNT signaling pathway has been identified as an ODC gene regulator [81]. The WNT signaling pathway, when activated, upregulates MYC [82], which acts as a transcriptional activator of ODC [187]. While the WNT cascade is normally suppressed in adult intestinal tissues, it can be disrupted by mutations. In particular, the APC tumor suppressor gene, a key component of the WNT cascade, has been identified as the cause of familial adenomatous polyposis (FAP) [188]. Over 80% of sporadic colorectal cancers are also attributable to APC mutations [189]. These mutations and dysregulated WNT signaling ultimately lead to an increase in MYC activity, ODC expression, and polyamine pools [83]. K-RAS's mutations are deemed to exert a central influence in both the initial stages of malignant progression of colorectal cells and the advanced metastatic ailment [190]. Rial et al., observed that mutant K-RAS upregulates ODC transcription and downregulates SSAT transcription, leading to an increase in polyamine biosynthesis and a decrease in polyamine catabolism (Fig. 3) [83]. Moreover, polyamines can be internalized by cells via the caveolin-mediated endocytosis pathway. The activation of K-RAS results in the phosphorylation of caveolin-1, which effectively enhances the endocytic process and subsequently increases the cellular intake of polyamines [84]. As early as 1986, it has reported that patients exhibiting elevated levels of polyamines in either their blood or urine tend to present with more advanced stages of disease and a poorer prognosis, especially the type of CRC [191, 192]. Elevated levels of polyamines can facilitate the hypusination of eukaryotic initiation factor 5A (eIF5A), thereby triggering the biosynthesis of MYC and initiating a positive feedback loop that consistently promotes the progression of CRC [193,

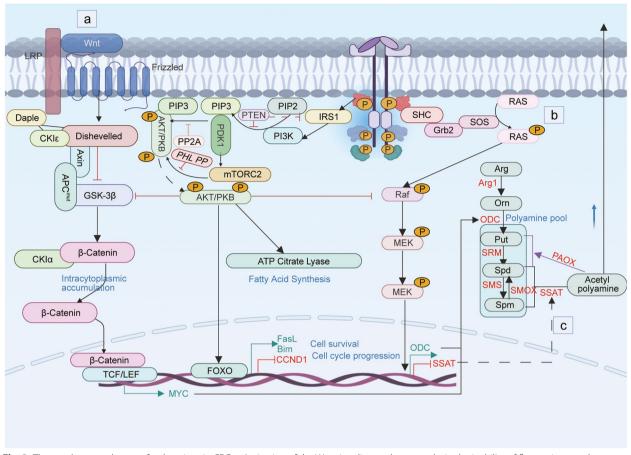


Fig. 3 The regulatory pathways of polyamines in CRC. **a** Activation of the Wnt signaling pathway results in the inability of β -catenin to undergo phosphorylation by GSK-3 β , ultimately leading to its accumulation within the cell and subsequent nuclear translocation. Upon entering the nucleus, MYC expression is upregulated through binding to TCF/LEF, which will increase the expression of ODC. **B** Excessive stimulation of the RAS signaling pathway results in the stimulation of ODC expression while suppressing the expression of SSAT. **c.** PI3K/AKT pathway can regulat the Wnt and Ras signaling pathways. Elevating the expression of ODC and reducing the expression of SSAT will result in an augmented intracellular pool of polyamines. The elevation in polyamine concentration frequently correlates with an unfavorable prognosis. Created with BioRender.com

194]. Disrupting either link within this loop efficiently inhibits CRC development.

Pancreatic cancer

Pancreatic cancer (PC) is a malignant neoplasm with an extraordinarily high mortality rate. Pancreatic ductal adenocarcinoma (PDAC) represents the predominant form of PC, comprising over 85% of all diagnosed cases [160]. PDAC is a type of cancer that is primarily driven by inflammation and is characterized by a substantial fibroinflammatory microenvironment [195]. Notably, PDAC's reliance on polyamine confers a number of survival advantages. First, the elevated concentrations of spermidine within the intracellular environment serve as a potential reservoir for the biosynthesis and secretion of substantial amounts of immunosuppressive spermine [196]. Second, it has been observed that spermidine can initiate autophagy by inhibiting EP-300 [85], a process that may promote the survival of PDAC cells in light of their increased metabolic rate, elevated levels of ROS and genomic instability [86, 87]. Elevated levels of spermidine synthesis are linked to increased expression of ODC. The upregulation of ODC is a consequence of the MTAP loss, a frequently observed occurrence in PC [88, 89]. Thirdly, the secretion of polyamines can furnish a plethora of polyamine substrates that can drive desmoplasia through extracellular transglutaminase 2 activity [196]. Desmoplasia is observable in both primary PDAC tumors and their metastatic counterparts [197]. Briefly stating, PDAC proliferation, microenvironment, desmoplasia, and immune function are just a few of the many functions of polyamines that have been elucidated in a large number of published articles. Combinatorial therapies that target polyamine depletion have demonstrated efficacy in inhibiting tumor growth and boosting immune response [198-200]. SBP-101, a Spm analogue that effectively reduces the activity of ODC and slightly promotes the catabolism of polyamine, has currently been evaluated in Phase 2/3 clinical trials for the treatment of PDAC (NCT05254171) [70]. Eventually, this field represents a promising avenue for drug discovery, and the successful development of polyamine-targeted therapies could have far-reaching implications for a variety of human diseases that hinge on polyamines for cellular growth and intercellular communication [201, 202].

Prostate cancers

Prostate cancer (PCa) is the most prevalent non-cutaneous neoplasm affecting the male population on a global scale, with an expected incidence of approximately 1.6 million cases and 366,000 deaths per year [203]. Despite advances in therapies targeting the androgen axis, the emergence of castration-resistant prostate cancer (CRPC) among patients is unavoidable [204]. Therefore, there continues to be an urgent need for more effective PCa treatments.

Luminal epithelial cells of the prostate gland exude amounts of acetylated polyamines into the prostatic lumen, rendering them vulnerable to disruptions in associated metabolic pathways [90]. Affronti et. al., have demonstrated that the rapid rate of flux through polyamine biosynthesis in PCa can be utilized in three distinct ways [90]. First, models of both androgen-stimulated and castration-recurrent PCa have revealed that dietary folate depletion is an efficacious means of treatment [205, 206]. Secondly, and rogen-sensitive models have shown that the inhibition of the methionine salvage pathway is an effective approach [207]. The current investigation has established that a combination of methionine salvage pathway inhibition and activation of polyamine catabolism is synergistic and highly effective in treating both androgensensitive and androgen-independent models of PCa [90]. In addition, a study demonstrates that PGC1- α is a vital regulator in the modulation of PCa cell aggressiveness and that the depletion of PGC1- α induces a dependence of prostate cancer cells on the polyamine pathway, which promotes metastasis [92]. PGC1-αreduces the expression of MYC and ODC, thereby significantly decreasing intracellular polyamine levels. In other words, PGC1-α effectively suppresses the metastatic properties of prostate cancer cells by downregulating the polyamine biosynthesis pathway. However, Li et al. have reported that the exogenous addition of Spm can impede the proliferation of CRPC [208]. Thus, further investigations are necessary to elucidate the exact function and specific molecular mechanisms underlying the influence of polyamine concentration on the growth and proliferation of prostate cancer cells.

Neuroblastoma

Neuroblastoma is a common childhood tumor that originates in the peripheral nervous system. The amplification of the MYCN proto-oncogene has a strong correlation with advanced disease and is the genetic characteristic most consistently associated with ineffective treatment [209, 210]. The oncogene ODC1, which encodes the rate-limiting enzyme in polyamine synthesis and serves as a compelling target [211], is a compelling mediator of MYC effects [187]. Due to the dysregulation of MYC, the activity of ODC is typically upregulated in cancer, resulting in an increase in polyamine levels that promote the rapid proliferation of tumor cells [99]. The administration of DFMO, a selective inhibitor of ODC, has been shown to be an effective strategy for the inhibition of both the occurrence and progression of MYC-associated Neuroblastoma [93-96, 98]. Currently, DFMO has progressed to the Phase 2 stage of clinical trials conducted in neuroblastoma (NCT02395666). Furthermore, it has been demonstrated that probenecid can exert inhibitory effects on the renal clearance of DFMO [212]. Therefore, Schultz et al. also indicated that the addition of probenecid as a supplementary medication during the course of neuroblastoma therapy enhanced the therapeutic outcome of DFMO. In this regard, Michael et al., provided evidence that the expansion of polyamines via the broad deregulation of regulatory enzymes, including ODC1, was a defining characteristic of neuroblastomas exhibiting MYCN amplification, and they identified a strong correlation between elevated levels of ODC1 and unfavorable clinical outcomes in a large patient cohort, including those without MYCN amplification [93]. Furthermore, MYC positively regulates the transcriptional activation of SLC3A2, leading to an increase in cellular uptake of polyamines, in addition to its role in upregulating ODC activity [97]. However, Lodeserto et al. argued that when the level of polyamine in cancer cells surpassed a certain threshold, it triggered the generation of oxidative metabolites which ultimately causes cell death [213]. Thus, they elicited cell death in neuroblastoma cell lines by elevating the polyamine concentration of cancer cells through the utilization of nanospermidine.

3. The effect and mechanism of oncogenes and signaling pathway on polyamine

Changes in polyamine concentrations may cause DNA damage [13], which has a close relationship with oncogene dysregulation and the development of cancer. Polyamines are integral components of the downstream target and signaling pathways of numerous oncogenes [75, 165, 214], and the content and activity of polyamines in tumors frequently undergo dramatic alterations [215, **216**]. Consequently, a comprehensive examination of the significance of the interaction between polyamines, oncogenes, and signaling pathways in the development and invasion of tumors would yield novel insights into the prevention and treatment of tumors (Fig. 4).

MYC

The MYC gene cluster, which consists of the c-MYC (MYCC), MYCN, and MYCL genes, has been intensively studied in cancer and developmental biology [217]. MYC activation has been observed in numerous types of

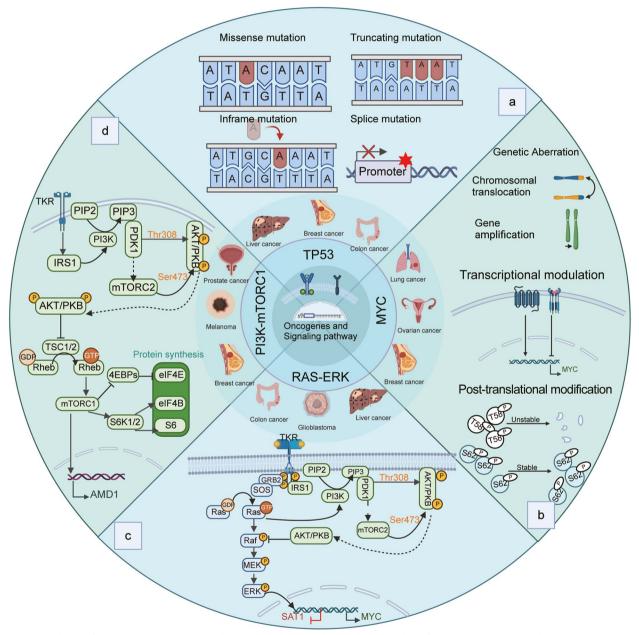


Fig. 4 Regulation of polyamine by oncogenes and signaling pathways. **a** The translation product of TP53, known as p53, regulates the expression of ODC, a key enzyme in polyamine synthesis. Mutations in the TP53 gene will have a significant impact on the metabolism and function of polyamines. The tumor suppressor gene TP53 can be inactivated in four ways: Missense mutation; Truncating mutation; Frameshift mutation; Splice mutation. **b** Activated MYC exerts regulatory effects on several crucial enzymes involved in polyamine metabolism, including ODC, SRM and SMS. Genetic aberrations, transcriptional regulation, and protein instability can activate MYC. Increasing phospho-serine 62 (P-S62)-MYC and decreasing phospho-threonine 58 (P-T58)-MYC stabilize MYC protein. **c.** RAS-RAF-MEK-ERK signaling pathway upregulates the expression of ODC and downregulates the expression of SSAT. **d** PI3K-mTORC1 signaling pathway increases the expression of dcAdoMet, thus regulating the biosynthesis of polyamine. Created with BioRender.com

cancer, which results from genetic mutations, transcriptional regulation, and protein stabilization.

The first indication of direct interaction between polyamine metabolism and oncogene was the demonstration that the MYC oncogene targeted the transcription of ornithine decarboxylase (ODC) [187]. The stimulation of growth, which results in the elevated expression of MYC, induces an increase in ODC1 mRNA, ODC protein, and ODC activity, thereby supplying the cells with the polyamines required for proliferation [187, 218]. Furthermore, Forshell et al. observed in their research that the stimulation of MYC directly led to the induction of SRM [219]. They also demonstrated that the chemical prevention of B-cell lymphoma via the suppression of SRM represented a promising therapeutic strategy. In addition, the expression of MYC may potentially influence the concentration of Spm through the direct stimulation of SMS [220]. Thus, the MYC signaling pathway is deemed as being among the principal culprits responsible for the dysregulation of polyamine metabolism in cancer [19]. As MYC family members are frequently amplified or overexpressed in cancer, the regulation of polyamine biosynthesis by MYC family genes emerges as a significant contributory factor in various cancer types, such as myeloproliferative neoplasms (MPNs) [221, 222], neuroblastoma [93, 223], CRC [224] and breast cancers [225, 226].

TP53

The TP53 gene is responsible for encoding the essential tumor suppressor protein p53, which is necessary for the maintenance of normal cellular growth and the prevention of tumorigenesis [227]. The p53 inhibits the expression of key enzymes involved in the urea cycle, resulting in a decrease in the overall rate of urea synthesis and the accumulation of excess ammonia [228]. The excessive accumulation of ammonia can induce a notable decrease in the expression levels of ODC mRNA [229], the ratelimiting factor in polyamine biosynthesis. As a result, the concentration of polyamines within the cell decreases. Moreover, spermine has the ability to induce autophagy by stimulating p53 [230], contributing to the suppression of tumorigenesis. Given the vital role polyamines play in cell proliferation and survival [13, 14], p53 can potentially exert anticancer effects by regulating the urea cycle and suppressing polyamine synthesis [228, 229]. Unfortunately, TP53 mutations are common in the vast majority of human cancers, resulting in impaired anti-tumor activity of the p53 protein and the protein's endowment with oncogenic properties [231, 232].

PI3K-mTOR complex 1 (mTORC1) signaling pathway

A fundamental signaling molecule that integrates metabolic and growth pathways is the mechanistic target of rapamycin (mTOR) [233, 234], an evolutionarily conserved serine/threonine kinase classified as a member of the phosphoinositide 3-kinase-related kinase (PI3K) family [235]. The central function of mTOR in cellular proliferation has been ascribed to mTORC1 [236], which, upon activation stimulates protein and lipid synthesis while inhibiting autophagy and lysosome formation [235]. In part, these responses are triggered by the phosphorylation of mTORC1 substrates, such as ribosomal S6 kinase 1 (S6K1), eukaryotic translation initiation factor 4E (eIF4E)-binding proteins 1 and 2 (4EBP1/2), and p70S6K1 [236, 237].

Akinyele et al., have confirmed that the modulation of polyamine levels results in significant changes in the phosphorylation levels of 4EBP1 and p70S6K, thereby affecting translation initiation in breast cancer cells, and that the knockdown of the mTOR gene inhibits cell proliferation while simultaneously causing a decrease in putrescine and spermidine content [214]. In addition, it has been shown that the PI3K-mTOR complex 1 (mTORC1) signaling pathway is associated with polyamine metabolism in PCa via the induction of AMD1 expression [91, 238, 239]. Current studies demonstrate that mTORC1 is required for the upregulation of AdoMetDC activity and levels of dcAdoMet, and the suppression of mTOR results in a significant decrease in AdoMetDC activity and intracellular polyamine levels [19, 91]. Initiation of polyamine catabolism also induces a shift in the positioning of mTOR in glioma cells, which has a negative effect on mTOR-related protein synthesis and ultimately leads to apoptosis [240].

RAS-RAF-MEK-ERK signaling pathway

The RAS/RAF/MEK/ERK signal transduction pathway is essential for cellular communication both within and between cells, regulating fundamental cellular processes such as growth, survival, differentiation, and oncogenesis [241]. As a substrate of RAS, ODC is composed of four distinct splice variants, each of which contains unique intronic sequences. Notably, all splice variants can be translated by means of IRES (internal ribosome entry site) mechanisms [242]. When the RAS signaling pathway is suppressed in non-cancerous cells, the IRES activity of ornithine decarboxylase (ODC) is reduced; however, in cells that have undergone RAS transformation, there is a noticeable rise in the activity of ODC IRES, leading to heightened levels of ODC activity [243]. Notably, RAS is highly prone to mutations in various types of human cancers, resulting in ODC activity [242]. Activated KRAS substantially enhances the uptake of polyamines, thereby altering the localization of the urokinase-type plasminogen activator receptor (uPAR) ligand and resulting in the stimulation of SRC [243]. When stimulated, the SRC enzyme catalyzes the addition of phosphate groups onto caveolin-1, an inhibitor of caveolar endocytosis [84], which has been shown to be associated with increased polyamine uptake in CRC cells [84]. Moreover, the impact of KRAS on the inhibition of SSAT has been proven through its disruption of the peroxisome proliferator-activated receptor- γ (PPAR γ) mediated transactivation of SAT1. This interference enables transformed cells to maintain enhanced polyamine levels [19, 244].

Summary

Our work focuses primarily on summarizing the function and mechanism of polyamines in maintaining health and contributing to disease. A brief summary of the origin and metabolism of polyamine is also provided. Ultimately, the manuscript describes the influence of oncogenes and signaling pathways on polyamine metabolism in a variety of cancer types via enzyme modulation. We hope that this review will provide some insights for the further elucidation and empirical study of polyamines.

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Authors' contributions

The idea for the article originated from Yuting He and Chen Xue. The literature search was performed by Mengjuan Xuan, Xinyu Gu and Juan Li. The first draft of the manuscript was written by Mengjuan Xuan, and critically revised by Yuting He and Chen Xue. Di Huang created the figures using Biorender.com. All authors read and approved the final manuscript.

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Availability of data and materials

No original datasets were generated for this review. All data supporting the information given here can be found in the references cited within the paper.

Declarations

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References

 Thomas T, Thomas TJ. Polyamines in cell growth and cell death: molecular mechanisms and therapeutic applications. Cell Mol Life Sci. 2001;58:244–58. https://doi.org/10.1007/pl00000852.

- 2. Wallace HM. The polyamines: past, present and future. Essays Biochem. 2009;46:1–9. https://doi.org/10.1042/bse0460001.
- Wallace HM, Fraser AV, Hughes A. A perspective of polyamine metabolism. Biochem J. 2003;376:1–14. https://doi.org/10.1042/bj20031327.
- Gerner EW, Meyskens FL Jr. Polyamines and cancer: old molecules, new understanding. Nat Rev Cancer. 2004;4:781–92. https://doi.org/10.1038/ nrc1454.
- 5. Pegg AE. Mammalian polyamine metabolism and function. IUBMB Life. 2009;61:880–94. https://doi.org/10.1002/iub.230.
- Đorđievski S, Vukašinović EL, Čelić TV, Pihler I, Kebert M, Kojić D, Purać J. Spermidine dietary supplementation and polyamines level in reference to survival and lifespan of honey bees. Sci Rep. 2023;13:4329. https:// doi.org/10.1038/s41598-023-31456-4.
- Madeo F, Eisenberg T, Büttner S, Ruckenstuhl C, Kroemer G. Spermidine: a novel autophagy inducer and longevity elixir. Autophagy. 2010;6:160– 2. https://doi.org/10.4161/auto.6.1.10600.
- Minois N, Carmona-Gutierrez D, Madeo F. Polyamines in aging and disease. Aging (Albany NY). 2011;3:716–32. https://doi.org/10.18632/ aging.100361.
- Handa AK, Fatima T, Mattoo AK. Polyamines: Bio-Molecules with Diverse Functions in Plant and Human Health and Disease. Front Chem. 2018;6:10. https://doi.org/10.3389/fchem.2018.00010.
- Pucciarelli S, Moreschini B, Micozzi D, De Fronzo GS, Carpi FM, Polzonetti V, Vincenzetti S, Mignini F, Napolioni V. Spermidine and spermine are enriched in whole blood of nona/centenarians. Rejuvenation Res. 2012;15:590–5. https://doi.org/10.1089/rej.2012.1349.
- Levêque J, Foucher F, Bansard JY, Havouis R, Grall JY, Moulinoux JP. Polyamine profiles in tumor, normal tissue of the homologous breast, blood, and urine of breast cancer sufferers. Breast Cancer Res Treat. 2000;60(2):99–105. https://doi.org/10.1023/a:1006319818530.
- Eisenberg T, Knauer H, Schauer A, Büttner S, Ruckenstuhl C, Carmona-Gutierrez D, Ring J, Schroeder S, Magnes C, Antonacci L, Fussi H, Deszcz L, Hartl R, Schraml E, Criollo A, Megalou E, Weiskopf D, Laun P, Heeren G, Breitenbach M, Grubeck-Loebenstein B, Herker E, Fahrenkrog B, Fröhlich KU, Sinner F, Tavernarakis N, Minois N, Kroemer G, Madeo F. Induction of autophagy by spermidine promotes longevity. Nat Cell Biol. 2009;11:1305–14. https://doi.org/10.1038/ncb1975.
- Pegg AE. Toxicity of polyamines and their metabolic products. Chem Res Toxicol. 2013;26:1782–800. https://doi.org/10.1021/tx400316s.
- Igarashi K, Kashiwagi K. Modulation of cellular function by polyamines. Int J Biochem Cell Biol. 2010;42:39–51. https://doi.org/10.1016/j.biocel. 2009.07.009.
- Miller-Fleming L, Olin-Sandoval V, Campbell K, Ralser M. Remaining Mysteries of Molecular Biology: The Role of Polyamines in the Cell. J Mol Biol. 2015;427:3389–406. https://doi.org/10.1016/j.jmb.2015.06.020.
- Casero RA, Pegg AE. Polyamine catabolism and disease. Biochem J. 2009;421:323–38. https://doi.org/10.1042/bj20090598.
- Nakanishi S, Cleveland JL (2021) Polyamine Homeostasis in Development and Disease. Med Sci (Basel). 9. https://doi.org/10.3390/medsc i9020028.
- Zahedi K, Barone S, Soleimani M (2022) Polyamines and Their Metabolism: From the Maintenance of Physiological Homeostasis to the Mediation of Disease. Med Sci (Basel). 10. https://doi.org/10.3390/medsci1003 0038.
- Casero RA Jr, Murray Stewart T, Pegg AE. Polyamine metabolism and cancer: treatments, challenges and opportunities. Nat Rev Cancer. 2018;18:681–95. https://doi.org/10.1038/s41568-018-0050-3.
- Holbert CE, Cullen MT, Casero RA Jr, Stewart TM. Polyamines in cancer: integrating organismal metabolism and antitumour immunity. Nat Rev Cancer. 2022;22:467–80. https://doi.org/10.1038/s41568-022-00473-2.
- Wu H, Fu B, Sun P, Xiao C, Liu JH. A NAC Transcription Factor Represses Putrescine Biosynthesis and Affects Drought Tolerance. Plant Physiol. 2016;172:1532–47. https://doi.org/10.1104/pp.16.01096.
- Thongbhubate K, Irie K, Sakai Y, Itoh A, Suzuki H. Improvement of putrescine production through the arginine decarboxylase pathway in Escherichia coli K-12. AMB Express. 2021;11:168. https://doi.org/10. 1186/s13568-021-01330-5.
- Coleman CS, Hu G, Pegg AE. Putrescine biosynthesis in mammalian tissues. Biochem J. 2004;379(Pt 3):849–55. https://doi.org/10.1042/BJ200 40035.

- Wang X, Ying W, Dunlap KA, Lin G, Satterfield MC, Burghardt RC, Wu G, Bazer FW. Arginine decarboxylase and agmatinase: an alternative pathway for de novo biosynthesis of polyamines for development of mammalian conceptuses. Biol Reprod. 2014;90:84. https://doi.org/10. 1095/biolreprod.113.114637.
- Pegg AE. Regulation of ornithine decarboxylase. J Biol Chem. 2006;281:14529–32. https://doi.org/10.1074/jbc.R500031200.
- Lee J, Michael AJ, Martynowski D, Goldsmith EJ, Phillips MA. Phylogenetic diversity and the structural basis of substrate specificity in the beta/alpha-barrel fold basic amino acid decarboxylases. J Biol Chem. 2007;282:27115–25. https://doi.org/10.1074/jbc.M704066200.
- 27. Pegg AE, McCann PP. Polyamine metabolism and function. Am J Physiol. 1982;243:C212–21. https://doi.org/10.1152/ajpcell.1982.243.5. C212.
- Ikeguchi Y, Bewley MC, Pegg AE. Aminopropyltransferases: function, structure and genetics. J Biochem. 2006;139:1–9. https://doi.org/10. 1093/jb/mvj019.
- 29. Wu H, Min J, Ikeguchi Y, Zeng H, Dong A, Loppnau P, Pegg AE, Plotnikov AN. Structure and mechanism of spermidine synthases. Biochemistry. 2007;46:8331–9. https://doi.org/10.1021/bi602498k.
- Wu H, Min J, Zeng H, McCloskey DE, Ikeguchi Y, Loppnau P, Michael AJ, Pegg AE, Plotnikov AN. Crystal structure of human spermine synthase: implications of substrate binding and catalytic mechanism. J Biol Chem. 2008;283:16135–46. https://doi.org/10.1074/jbc.M710323200.
- Tang B, Lee HO, Gupta S, Wang L, Kurimchak AM, Duncan JS, Kruger WD. Extracellular 5'-methylthioadenosine inhibits intracellular symmetric dimethylarginine protein methylation of FUSE-binding proteins. J Biol Chem. 2022;298:102367. https://doi.org/10.1016/j.jbc.2022.102367.
- 32. Seiler N. Catabolism of polyamines. Amino Acids. 2004;26:217–33. https://doi.org/10.1007/s00726-004-0070-z.
- Wang Y, Casero RA Jr. Mammalian polyamine catabolism: a therapeutic target, a pathological problem, or both? J Biochem. 2006;139:17–25. https://doi.org/10.1093/jb/mvj021.
- Wang Y, Hacker A, Murray-Stewart T, Frydman B, Valasinas A, Fraser AV, Woster PM, Casero RA Jr. Properties of recombinant human N1-acetylpolyamine oxidase (hPAO): potential role in determining drug sensitivity. Cancer Chemother Pharmacol. 2005;56:83–90. https://doi.org/10. 1007/s00280-004-0936-5.
- Wu T, Ling KQ, Sayre LM, McIntire WS. Inhibition of murine N1-acetylated polyamine oxidase by an acetylenic amine and the allenic amine, MDL 72527. Biochem Biophys Res Commun. 2005;326:483–90. https:// doi.org/10.1016/j.bbrc.2004.11.054.
- Takao K, Shibata S, Ozawa T, Wada M, Sugitia Y, Samejima K, Shirahata A. A conceptual model of the polyamine binding site of N1-acetylpolyamine oxidase developed from a study of polyamine derivatives. Amino Acids. 2009;37:401–5. https://doi.org/10.1007/s00726-008-0168-9.
- Henderson Pozzi M, Gawandi V, Fitzpatrick PF. pH dependence of a mammalian polyamine oxidase: insights into substrate specificity and the role of lysine 315. Biochemistry. 2009;48:1508–16. https://doi.org/ 10.1021/bi802227m.
- Pegg AE. Spermidine/spermine-N(1)-acetyltransferase: a key metabolic regulator. Am J Physiol Endocrinol Metab. 2008;294:E995–1010. https:// doi.org/10.1152/ajpendo.90217.2008.
- Uemura T, Akasaka Y, Ikegaya H. Correlation of polyamines, acroleinconjugated lysine and polyamine metabolic enzyme levels with age in human liver. Heliyon. 2020;6:e05031. https://doi.org/10.1016/j.heliyon. 2020.e05031.
- Yoshida M, Tomitori H, Machi Y, Hagihara M, Higashi K, Goda H, Ohya T, Niitsu M, Kashiwagi K, Igarashi K. Acrolein toxicity: Comparison with reactive oxygen species. Biochem Biophys Res Commun. 2009;378:313– 8. https://doi.org/10.1016/j.bbrc.2008.11.054.
- Igarashi K, Ueda S, Yoshida K, Kashiwagi K. Polyamines in renal failure. Amino Acids. 2006;31:477–83. https://doi.org/10.1007/ s00726-006-0264-7.
- Masuko T, Takao K, Samejima K, Shirahata A, Igarashi K, Casero RA Jr, Kizawa Y, Sugita Y. N(1)-Nonyl-1,4-diaminobutane ameliorates brain infarction size in photochemically induced thrombosis model mice. Neurosci Lett. 2018;672:118–22. https://doi.org/10.1016/j.neulet.2018. 01.054.
- Tomitori H, Usui T, Saeki N, Ueda S, Kase H, Nishimura K, Kashiwagi K, Igarashi K. Polyamine oxidase and acrolein as novel biochemical

markers for diagnosis of cerebral stroke. Stroke. 2005;36:2609–13. https://doi.org/10.1161/01.STR.0000190004.36793.2d.

- Madeo F, Eisenberg T, Pietrocola F, Kroemer G (2018) Spermidine in health and disease. Science. 359. https://doi.org/10.1126/science.aan27 88.
- 45. Seiler N. Polyamine metabolism. Digestion. 1990;46(Suppl 2):319–30. https://doi.org/10.1159/000200405.
- 46. Sarhan S, Knodgen B, Seiler N. The gastrointestinal tract as polyamine source for tumor growth. Anticancer Res. 1989;9:215–23.
- Cipolla B, Guillí F, Moulinoux JP. Polyamine-reduced diet in metastatic hormone-refractory prostate cancer (HRPC) patients. Biochem Soc Trans. 2003;31:384–7. https://doi.org/10.1042/bst0310384.
- Bardocz S, Brown DS, Grant G, Pusztai A. Luminal and basolateral polyamine uptake by rat small intestine stimulated to grow by Phaseolus vulgaris lectin phytohaemagglutinin in vivo. Biochim Biophys Acta. 1990;1034:46–52. https://doi.org/10.1016/0304-4165(90)90151-l.
- Uda K, Tsujikawa T, Fujiyama Y, Bamba T. Rapid absorption of luminal polyamines in a rat small intestine ex vivo model. J Gastroenterol Hepatol. 2003;18:554–9. https://doi.org/10.1046/j.1440-1746.2003.03020.x.
- Cipolla BG, Havouis R, Moulinoux JP. Polyamine reduced diet (PRD) nutrition therapy in hormone refractory prostate cancer patients. Biomed Pharmacother. 2010;64:363–8. https://doi.org/10.1016/j.biopha. 2009.09.022.
- Mandal S, Mandal A, Johansson HE, Orjalo AV, Park MH. Depletion of cellular polyamines, spermidine and spermine, causes a total arrest in translation and growth in mammalian cells. Proc Natl Acad Sci U S A. 2013;110:2169–74. https://doi.org/10.1073/pnas.1219002110.
- Abdulhussein AA, Wallace HM. Polyamines and membrane transporters. Amino Acids. 2014;46:655–60. https://doi.org/10.1007/ s00726-013-1553-6.
- Belting M, Mani K, Jonsson M, Cheng F, Sandgren S, Jonsson S, Ding K, Delcros JG, Fransson LA. Glypican-1 is a vehicle for polyamine uptake in mammalian cells: a pivital role for nitrosothiol-derived nitric oxide. J Biol Chem. 2003;278:47181–9. https://doi.org/10.1074/jbc.M308325200.
- Nowotarski SL, Woster PM, Casero RA Jr. Polyamines and cancer: implications for chemotherapy and chemoprevention. Expert Rev Mol Med. 2013;15:e3. https://doi.org/10.1017/erm.2013.3.
- van Veen S, Martin S, Van den Haute C, Benoy V, Lyons J, Vanhoutte R, Kahler JP, Decuypere JP, Gelders G, Lambie E, Zielich J, Swinnen JV, Annaert W, Agostinis P, Ghesquiere B, Verhelst S, Baekelandt V, Eggermont J, Vangheluwe P. ATP13A2 deficiency disrupts lysosomal polyamine export. Nature. 2020;578:419–24. https://doi.org/10.1038/ s41586-020-1968-7.
- Sekhar V, Andl T, Ot P. ATP13A3 facilitates polyamine transport in human pancreatic cancer cells. Sci Rep. 2022;12:4045. https://doi.org/10.1038/ s41598-022-07712-4.
- Hamouda NN, Van den Haute C, Vanhoutte R, Sannerud R, Azfar M, Mayer R, Cortes Calabuig A, Swinnen JV, Agostinis P, Baekelandt V, Annaert W, Impens F, Verhelst SHL, Eggermont J, Martin S, Vangheluwe P. ATP13A3 is a major component of the enigmatic mammalian polyamine transport system. J Biol Chem. 2021;296:100182. https://doi.org/ 10.1074/jbc.RA120.013908.
- Uemura T, Yerushalmi HF, Tsaprailis G, Stringer DE, Pastorian KE, Hawel L 3rd, Byus CV, Gerner EW. Identification and characterization of a diamine exporter in colon epithelial cells. J Biol Chem. 2008;283:26428– 35. https://doi.org/10.1074/jbc.M804714200.
- Hiasa M, Miyaji T, Haruna Y, Takeuchi T, Harada Y, Moriyama S, Yamamoto A, Omote H, Moriyama Y. Identification of a mammalian vesicular polyamine transporter. Sci Rep. 2014;4:6836. https://doi.org/10.1038/srep0 6836.
- Takeuchi T, Harada Y, Moriyama S, Furuta K, Tanaka S, Miyaji T, Omote H, Moriyama Y, Hiasa M. Vesicular Polyamine Transporter Mediates Vesicular Storage and Release of Polyamine from Mast Cells. J Biol Chem. 2017;292:3909–18. https://doi.org/10.1074/jbc.M116.756197.
- Wendth AJ Jr, Garlick WB, Pantoja GE, Shamoun J. Transcatheter biopsy of renal carcinoma invading the inferior vena cava. J Urol. 1976;115:331–2. https://doi.org/10.1016/s0022-5347(17)59193-4.
- Basu Roy UK, Henkhaus RS, Loupakis F, Cremolini C, Gerner EW, Ignatenko NA. Caveolin-1 is a novel regulator of K-RAS-dependent migration in colon carcinogenesis. Int J Cancer. 2013;133:43–57. https:// doi.org/10.1002/ijc.28001.

- 63. Eisenberg T, Abdellatif M, Schroeder S, Primessnig U, Stekovic S, Pendl T, Harger A, Schipke J, Zimmermann A, Schmidt A, Tong M, Ruckenstuhl C, Dammbrueck C, Gross AS, Herbst V, Magnes C, Trausinger G, Narath S, Meinitzer A, Hu Z, Kirsch A, Eller K, Carmona-Gutierrez D, Büttner S, Pietrocola F, Knittelfelder O, Schrepfer E, Rockenfeller P, Simonini C, Rahn A, Horsch M, Moreth K, Beckers J, Fuchs H, Gailus-Durner V, Neff F, Janik D, Rathkolb B, Rozman J, de Angelis MH, Moustafa T, Haemmerle G, Mayr M, Willeit P, von Frieling-Salewsky M, Pieske B, Scorrano L, Pieber T, Pechlaner R, Willeit J, Sigrist SJ, Linke WA, Mühlfeld C, Sadoshima J, Dengjel J, Kiechl S, Kroemer G, Sedej S, Madeo F. Cardioprotection and lifespan extension by the natural polyamine spermidine. Nat Med. 2016;22:1428–38. https://doi.org/10.1038/nm.4222.
- Soda K, Kano Y, Chiba F. Food polyamine and cardiovascular disease–an epidemiological study. Glob J Health Sci. 2012;4:170–8. https://doi.org/ 10.5539/gjhs.v4n6p170.
- Bonhoure N, Byrnes A, Moir RD, Hodroj W, Preitner F, Praz V, Marcelin G, Chua SC Jr, Martinez-Lopez N, Singh R, Moullan N, Auwerx J, Willemin G, Shah H, Hartil K, Vaitheesvaran B, Kurland I, Hernandez N, Willis IM. Loss of the RNA polymerase III repressor MAF1 confers obesity resistance. Genes Dev. 2015;29:934–47. https://doi.org/10.1101/gad.258350.115.
- 66. Fernández ÁF, Bárcena C, Martínez-García GG, Tamargo-Gómez I, Suárez MF, Pietrocola F, Castoldi F, Esteban L, Sierra-Filardi E, Boya P, López-Otín C, Kroemer G, Mariño G. Autophagy couteracts weight gain, lipotoxicity and pancreatic β-cell death upon hypercaloric pro-diabetic regimens. Cell Death Dis. 2017;8:e2970. https://doi.org/10.1038/cddis.2017.373.
- Hwang WY, Park WH, Suh DH, Kim K, Kim YB, No JH (2021) Difluoromethylornithine Induces Apoptosis through Regulation of AP-1 Signaling via JNK Phosphorylation in Epithelial Ovarian Cancer. Int J Mol Sci. 22. https://doi.org/10.3390/ijms221910255.
- Travers M, Brown SM, Dunworth M, Holbert CE, Wiehagen KR, Bachman KE, Foley JR, Stone ML, Baylin SB, Casero RA Jr, Zahnow CA. DFMO and 5-Azacytidine Increase M1 Macrophages in the Tumor Microenvironment of Murine Ovarian Cancer. Cancer Res. 2019;79:3445–54. https:// doi.org/10.1158/0008-5472.Can-18-4018.
- El Naggar O, Doyle B, Mariner K, Gilmour SK (2022) Difluoromethylornithine (DFMO) Enhances the Cytotoxicity of PARP Inhibition in Ovarian Cancer Cells. Med Sci (Basel). 10. https://doi.org/10.3390/medsci1002 0028.
- Holbert CE, Foley JR, Murray Stewart T, Casero RA, Jr. (2022) Expanded Potential of the Polyamine Analogue SBP-101 (Diethyl Dihydroxyhomospermine) as a Modulator of Polyamine Metabolism and Cancer Therapeutic. Int J Mol Sci. 23. https://doi.org/10.3390/ijms23126798.
- Frau M, Feo F, Pascale RM. Pleiotropic effects of methionine adenosyltransferases deregulation as determinants of liver cancer progression and prognosis. J Hepatol. 2013;59:830–41. https://doi.org/10.1016/j. jhep.2013.04.031.
- Feo F, Garcea R, Pascale R, Pirisi L, Daino L, Donaera A. The variations of S-adenosyl-L-methionine content modulate hepatocyte growth during phenobarbital promotion of diethylnitrosamine-induced rat liver carcinogenesis. Toxicol Pathol. 1987;15:109–14. https://doi.org/10.1177/ 019262338701500117.
- Heby O. Role of polyamines in the control of cell proliferation and differentiation. Differentiation. 1981;19:1–20. https://doi.org/10.1111/j. 1432-0436.1981.tb01123.x.
- Tomasi ML, Ryoo M, Skay A, Tomasi I, Giordano P, Mato JM, Lu SC. Polyamine and methionine adenosyltransferase 2A crosstalk in human colon and liver cancer. Exp Cell Res. 2013;319:1902–11. https://doi.org/ 10.1016/j.yexcr.2013.04.005.
- 75. Wang C, Ruan P, Zhao Y, Li X, Wang J, Wu X, Liu T, Wang S, Hou J, Li W, Li Q, Li J, Dai F, Fang D, Wang C, Xie S. Spermidine/spermine N1-acetyltransferase regulates cell growth and metastasis via AKT/β-catenin signaling pathways in hepatocellular and colorectal carcinoma cells. Oncotarget. 2017;8:1092–109. https://doi.org/10.18632/oncotarget. 13582.
- Yue F, Li W, Zou J, Jiang X, Xu G, Huang H, Liu L. Spermidine Prolongs Lifespan and Prevents Liver Fibrosis and Hepatocellular Carcinoma by Activating MAP1S-Mediated Autophagy. Cancer Res. 2017;77:2938–51. https://doi.org/10.1158/0008-5472.Can-16-3462.
- 77. Sierra JC, Piazuelo MB, Luis PB, Barry DP, Allaman MM, Asim M, Sebrell TA, Finley JL, Rose KL, Hill S, Holshouser SL, Casero RA, Cleveland JL, Woster PM, Schey KL, Bimczok D, Schneider C, Gobert AP, Wilson

KT. Spermine oxidase mediates Helicobacter pylori-induced gastric inflammation, DNA damage, and carcinogenic signaling. Oncogene. 2020;39:4465–74. https://doi.org/10.1038/s41388-020-1304-6.

- 78. Chaturvedi R, de Sablet T, Asim M, Piazuelo MB, Barry DP, Verriere TG, Sierra JC, Hardbower DM, Delgado AG, Schneider BG, Israel DA, Romero-Gallo J, Nagy TA, Morgan DR, Murray-Stewart T, Bravo LE, Peek RM Jr, Fox JG, Woster PM, Casero RA Jr, Correa P, Wilson KT. Increased Helicobacter pylori-associated gastric cancer risk in the Andean region of Colombia is mediated by spermine oxidase. Oncogene. 2015;34:3429–40. https://doi.org/10.1038/onc.2014.273.
- Chaturvedi R, Asim M, Romero-Gallo J, Barry DP, Hoge S, de Sablet T, Delgado AG, Wroblewski LE, Piazuelo MB, Yan F, Israel DA, Casero RA Jr, Correa P, Gobert AP, Polk DB, Peek RM Jr, Wilson KT. Spermine oxidase mediates the gastric cancer risk associated with Helicobacter pylori CagA. Gastroenterology. 2011;141:1696–708.e1-2. https://doi.org/10. 1053/j.gastro.2011.07.045.
- Xu L, You X, Cao Q, Huang M, Hong LL, Chen XL, Lei L, Ling ZQ, Chen Y. Polyamine synthesis enzyme AMD1 is closely associated with tumorigenesis and prognosis of human gastric cancers. Carcinogenesis. 2020;41:214–22. https://doi.org/10.1093/carcin/bgz098.
- Fultz KE, Gerner EW. APC-dependent regulation of ornithine decarboxylase in human colon tumor cells. Mol Carcinog. 2002;34:10–8. https:// doi.org/10.1002/mc.10043.
- He TC, Sparks AB, Rago C, Hermeking H, Zawel L, da Costa LT, Morin PJ, Vogelstein B, Kinzler KW. Identification of c-MYC as a target of the APC pathway. Science. 1998;281:1509–12. https://doi.org/10.1126/science. 281.5382.1509.
- Rial NS, Meyskens FL, Gerner EW. Polyamines as mediators of APCdependent intestinal carcinogenesis and cancer chemoprevention. Essays Biochem. 2009;46:111–24. https://doi.org/10.1042/bse0460008.
- Roy UK, Rial NS, Kachel KL, Gerner EW. Activated K-RAS increases polyamine uptake in human colon cancer cells through modulation of caveolar endocytosis. Mol Carcinog. 2008;47:538–53. https://doi.org/10. 1002/mc.20414.
- Pietrocola F, Lachkar S, Enot DP, Niso-Santano M, Bravo-San Pedro JM, Sica V, Izzo V, Maiuri MC, Madeo F, Mariño G, Kroemer G. Spermidine induces autophagy by inhibiting the acetyltransferase EP300. Cell Death Differ. 2015;22:509–16. https://doi.org/10.1038/cdd.2014.215.
- Yang S, Kimmelman AC. A critical role for autophagy in pancreatic cancer. Autophagy. 2011;7:912–3. https://doi.org/10.4161/auto.7.8.15762.
- Yang S, Wang X, Contino G, Liesa M, Sahin E, Ying H, Bause A, Li Y, Stommel JM, Dell'antonio G, Mautner J, Tonon G, Haigis M, Shirihai OS, Doglioni C, Bardeesy N, Kimmelman AC. Pancreatic cancers require autophagy for tumor growth. Genes Dev. 2011;25:717–29. https://doi. org/10.1101/gad.2016111.
- Subhi AL, Tang B, Balsara BR, Altomare DA, Testa JR, Cooper HS, Hoffman JP, Meropol NJ, Kruger WD. Loss of methylthioadenosine phosphorylase and elevated ornithine decarboxylase is common in pancreatic cancer. Clin Cancer Res. 2004;10:7290–6. https://doi.org/10.1158/1078-0432. Ccr-04-0972.
- Subhi AL, Diegelman P, Porter CW, Tang B, Lu ZJ, Markham GD, Kruger WD. Methylthioadenosine phosphorylase regulates ornithine decarboxylase by production of downstream metabolites. J Biol Chem. 2003;278:49868–73. https://doi.org/10.1074/jbc.M308451200.
- Affronti HC, Rowsam AM, Pellerite AJ, Rosario SR, Long MD, Jacobi JJ, Bianchi-Smiraglia A, Boerlin CS, Gillard BM, Karasik E, Foster BA, Moser M, Wilton JH, Attwood K, Nikiforov MA, Azabdaftari G, Pili R, Phillips JG, Casero RA Jr, Smiraglia DJ. Pharmacological polyamine catabolism upregulation with methionine salvage pathway inhibition as an effective prostate cancer therapy. Nat Commun. 2020;11:52. https://doi.org/ 10.1038/s41467-019-13950-4.
- 91. Zabala-Letona A, Arruabarrena-Aristorena A, Martín-Martín N, Fernandez-Ruiz S, Sutherland JD, Clasquin M, Tomas-Cortazar J, Jimenez J, Torres I, Quang P, Ximenez-Embun P, Bago R, Ugalde-Olano A, Loizaga-Iriarte A, Lacasa-Viscasillas I, Unda M, Torrano V, Cabrera D, van Liempd SM, Cendon Y, Castro E, Murray S, Revandkar A, Alimonti A, Zhang Y, Barnett A, Lein G, Pirman D, Cortazar AR, Arreal L, Prudkin L, Astobiza I, Valcarcel-Jimenez L, Zuñiga-García P, Fernandez-Dominguez I, Piva M, Caro-Maldonado A, Sánchez-Mosquera P, Castillo-Martín M, Serra V, Beraza N, Gentilella A, Thomas G, Azkargorta M, Elortza F, Farràs R, Olmos D, Efeyan A, Anguita J, Muñoz J, Falcón-Pérez JM, Barrio R,

Macarulla T, Mato JM, Martinez-Chantar ML, Cordon-Cardo C, Aransay AM, Marks K, Baselga J, Tabernero J, Nuciforo P, Manning BD, Marjon K, Carracedo A. mTORC1-dependent AMD1 regulation sustains polyamine metabolism in prostate cancer. Nature. 2017;547:109–13. https://doi.org/10.1038/nature22964.

- Kaminski L, Torrino S, Dufies M, Djabari Z, Haider R, Roustan FR, Jaune E, Laurent K, Nottet N, Michiels JF, Gesson M, Rocchi S, Mazure NM, Durand M, Tanti JF, Ambrosetti D, Clavel S, Ben-Sahra I, Bost F. PGC1α Inhibits Polyamine Synthesis to Suppress Prostate Cancer Aggressiveness. Cancer Res. 2019;79:3268–80. https://doi.org/10.1158/0008-5472. Can-18-2043.
- Hogarty MD, Norris MD, Davis K, Liu X, Evageliou NF, Hayes CS, Pawel B, Guo R, Zhao H, Sekyere E, Keating J, Thomas W, Cheng NC, Murray J, Smith J, Sutton R, Venn N, London WB, Buxton A, Gilmour SK, Marshall GM, Haber M. ODC1 is a critical determinant of MYCN oncogenesis and a therapeutic target in neuroblastoma. Cancer Res. 2008;68:9735–45. https://doi.org/10.1158/0008-5472.Can-07-6866.
- Evageliou NF, Haber M, Vu A, Laetsch TW, Murray J, Gamble LD, Cheng NC, Liu K, Reese M, Corrigan KA, Ziegler DS, Webber H, Hayes CS, Pawel B, Marshall GM, Zhao H, Gilmour SK, Norris MD, Hogarty MD. Polyamine Antagonist Therapies Inhibit Neuroblastoma Initiation and Progression. Clin Cancer Res. 2016;22:4391–404. https://doi.org/10.1158/1078-0432. Ccr-15-2539.
- Rounbehler RJ, Li W, Hall MA, Yang C, Fallahi M, Cleveland JL. Targeting ornithine decarboxylase impairs development of MYCN-amplified neuroblastoma. Cancer Res. 2009;69:547–53. https://doi.org/10.1158/ 0008-5472.Can-08-2968.
- Bassiri H, Benavides A, Haber M, Gilmour SK, Norris MD, Hogarty MD. Translational development of difluoromethylornithine (DFMO) for the treatment of neuroblastoma. Transl Pediatr. 2015;4:226–38. https://doi. org/10.3978/j.issn.2224-4336.2015.04.06.
- 97. Gamble LD, Purgato S, Murray J, Xiao L, Yu DMT, Hanssen KM, Giorgi FM, Carter DR, Gifford AJ, Valli E, Milazzo G, Kamili A, Mayoh C, Liu B, Eden G, Sarraf S, Allan S, Di Giacomo S, Flemming CL, Russell AJ, Cheung BB, Oberthuer A, London WB, Fischer M, Trahair TN, Fletcher JJ, Marshall GM, Ziegler DS, Hogarty MD, Burns MR, Perini G, Norris MD, Haber M (2019) Inhibition of polyamine synthesis and uptake reduces tumor progression and prolongs survival in mouse models of neuroblastoma. Sci Transl Med. 11. https://doi.org/10.1126/scitranslmed.aau1099.
- Samal K, Zhao P, Kendzicky A, Yco LP, McClung H, Gerner E, Burns M, Bachmann AS, Sholler G. AMXT-1501, a novel polyamine transport inhibitor, synergizes with DFMO in inhibiting neuroblastoma cell proliferation by targeting both ornithine decarboxylase and polyamine transport. Int J Cancer. 2013;133:1323–33. https://doi.org/10.1002/ijc. 28139.
- Ruiz-Pérez MV, Medina M, Urdiales JL, Keinänen TA, Sánchez-Jiménez F. Polyamine metabolism is sensitive to glycolysis inhibition in human neuroblastoma cells. J Biol Chem. 2015;290:6106–19. https://doi.org/10. 1074/jbc.M114.619197.
- Takahashi Y, Mai M, Nishioka K. alpha-difluoromethylornithine induces apoptosis as well as anti-angiogenesis in the inhibition of tumor growth and metastasis in a human gastric cancer model. Int J Cancer. 2000;85:243–7.
- LaRocca TJ, Gioscia-Ryan RA, Hearon CM Jr, Seals DR. The autophagy enhancer spermidine reverses arterial aging. Mech Ageing Dev. 2013;134:314–20. https://doi.org/10.1016/j.mad.2013.04.004.
- Soda K. Polyamine intake, dietary pattern, and cardiovascular disease. Med Hypotheses. 2010;75:299–301. https://doi.org/10.1016/j.mehy. 2010.03.008.
- 103. de Cabo R, Navas P. Spermidine to the rescue for an aging heart. Nat Med. 2016;22:1389–90. https://doi.org/10.1038/nm.4243.
- 104. Yan J, Yan JY, Wang YX, Ling YN, Song XD, Wang SY, Liu HQ, Liu QC, Zhang Y, Yang PZ, Wang XB, Chen AH. Spermidine-enhanced autophagic flux improves cardiac dysfunction following myocardial infarction by targeting the AMPK/mTOR signalling pathway. Br J Pharmacol. 2019;176:3126–42. https://doi.org/10.1111/bph.14706.
- 105. Wang J, Li S, Wang J, Wu F, Chen Y, Zhang H, Guo Y, Lin Y, Li L, Yu X, Liu T, Zhao Y. Spermidine alleviates cardiac aging by improving mitochondrial biogenesis and function. Aging (Albany NY). 2020;12:650–71. https:// doi.org/10.18632/aging.102647.

- 106. Wong W. Pathogenic putrescine. Sci Signal. 2022;15:eade8161. https:// doi.org/10.1126/scisignal.ade8161.
- 107. Ju YH, Bhalla M, Hyeon SJ, Oh JE, Yoo S, Chae U, Kwon J, Koh W, Lim J, Park YM, Lee J, Cho IJ, Lee H, Ryu H, Lee CJ. Astrocytic urea cycle detoxifies Aβ-derived ammonia while impairing memory in Alzheimer's disease. Cell Metab. 2022;34:1104–20.e8. https://doi.org/10.1016/j.cmet. 2022.05.011.
- Jain V, Raina S, Gheware AP, Singh R, Rehman R, Negi V, Murray Stewart T, Mabalirajan U, Mishra AK, Casero RA Jr, Agrawal A, Ghosh B. Reduction in polyamine catabolism leads to spermine-mediated airway epithelial injury and induces asthma features. Allergy. 2018;73:2033–45. https://doi.org/10.1111/all.13472.
- North ML, Grasemann H, Khanna N, Inman MD, Gauvreau GM, Scott JA. Increased ornithine-derived polyamines cause airway hyperresponsiveness in a mouse model of asthma. Am J Respir Cell Mol Biol. 2013;48:694–702. https://doi.org/10.1165/rcmb.2012-0323OC.
- 110. Pirinen E, Kuulasmaa T, Pietilä M, Heikkinen S, Tusa M, Itkonen P, Boman S, Skommer J, Virkamäki A, Hohtola E, Kettunen M, Fatrai S, Kansanen E, Koota S, Niiranen K, Parkkinen J, Levonen AL, Ylä-Herttuala S, Hiltunen JK, Alhonen L, Smith U, Jänne J, Laakso M. Enhanced polyamine catabolism alters homeostatic control of white adipose tissue mass, energy expenditure, and glucose metabolism. Mol Cell Biol. 2007;27:4953–67. https://doi.org/10.1128/mcb.02034-06.
- 111. Jell J, Merali S, Hensen ML, Mazurchuk R, Spernyak JA, Diegelman P, Kisiel ND, Barrero C, Deeb KK, Alhonen L, Patel MS, Porter CW. Genetically altered expression of spermidine/spermine N1-acetyltransferase affects fat metabolism in mice via acetyl-CoA. J Biol Chem. 2007;282:8404–13. https://doi.org/10.1074/jbc.M610265200.
- 112. Sadasivan SK, Vasamsetti B, Singh J, Marikunte VV, Oommen AM, Jagannath MR, Pralhada Rao R. Exogenous administration of spermine improves glucose utilization and decreases bodyweight in mice. Eur J Pharmacol. 2014;729:94–9. https://doi.org/10.1016/j.ejphar.2014.01.073.
- Morales TS, Avis EC, Paskowski EK, Shabar H, Nowotarski SL, DiAngelo JR (2021) The Role of Spermidine Synthase (SpdS) and Spermine Synthase (Sms) in Regulating Triglyceride Storage in Drosophila. Med Sci (Basel).
 https://doi.org/10.3390/medsci9020027.
- Liu C, Perez-Leal O, Barrero C, Zahedi K, Soleimani M, Porter C, Merali S. Modulation of polyamine metabolic flux in adipose tissue alters the accumulation of body fat by affecting glucose homeostasis. Amino Acids. 2014;46:701–15. https://doi.org/10.1007/s00726-013-1548-3.
- 115. Hyvönen MT, Herzig KH, Sinervirta R, Albrecht E, Nordback I, Sand J, Keinänen TA, Vepsäläinen J, Grigorenko N, Khomutov AR, Krüger B, Jänne J, Alhonen L. activated polyamine catabolism in acute pancreatitis: alpha-methylated polyamine analogues prevent trypsinogen activation and pancreatitis-associated mortality. Am J Pathol. 2006;168:115– 22. https://doi.org/10.2353/ajpath.2006.050518.
- Alhonen L, Parkkinen JJ, Keinanen T, Sinervirta R, Herzig KH, Jänne J. Activation of polyamine catabolism in transgenic rats induces acute pancreatitis. Proc Natl Acad Sci U S A. 2000;97:8290–5. https://doi.org/ 10.1073/pnas.140122097.
- Hyvönen MT, Merentie M, Uimari A, Keinänen TA, Jänne J, Alhonen L. Mechanisms of polyamine catabolism-induced acute pancreatitis. Biochem Soc Trans. 2007;35:326–30. https://doi.org/10.1042/bst0350326.
- 118. Lou F, Sun Y, Xu Z, Niu L, Wang Z, Deng S, Liu Z, Zhou H, Bai J, Yin Q, Cai X, Sun L, Wang H, Li Q, Wu Z, Chen X, Gu J, Shi YL, Tao W, Ginhoux F, Wang H. Excessive Polyamine Generation in Keratinocytes Promotes Self-RNA Sensing by Dendritic Cells in Psoriasis. Immunity. 2020;53:204– 16.e10. https://doi.org/10.1016/j.immuni.2020.06.004.
- Brzozowski T, Konturek SJ, Drozdowicz D, Dembiński A, Stachura J. Healing of chronic gastric ulcerations by L-arginine. Role of nitric oxide, prostaglandins, gastrin and polyamines. Digestion. 1995;56:463–71. https://doi.org/10.1159/000201277.
- 120. Takigawa M, Enomoto M, Nishida Y, Pan HO, Kinoshita A, Suzuki F. Tumor angiogenesis and polyamines: alpha-difluoromethylornithine, an irreversible inhibitor of ornithine decarboxylase, inhibits B16 melanoma-induced angiogenesis in ovo and the proliferation of vascular endothelial cells in vitro. Cancer Res. 1990;50:4131–8.
- Lakatta EG, Levy D. Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises: Part I: aging arteries: a "set up" for vascular disease. Circulation. 2003;107:139–46. https://doi.org/10.1161/ 01.cir.0000048892.83521.58.

- 122. North BJ, Sinclair DA. The intersection between aging and cardiovascular disease. Circ Res. 2012;110:1097–108. https://doi.org/10.1161/circr esaha.111.246876.
- O'Rourke MF, Hashimoto J. Mechanical factors in arterial aging: a clinical perspective. J Am Coll Cardiol. 2007;50:1–13. https://doi.org/10.1016/j. jacc.2006.12.050.
- 124. Brandes RP, Fleming I, Busse R. Endothelial aging. Cardiovasc Res. 2005;66:286–94. https://doi.org/10.1016/j.cardiores.2004.12.027.
- 125. Lakatta EG. Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises: Part III: cellular and molecular clues to heart and arterial aging. Circulation. 2003;107:490–7. https://doi.org/10.1161/ 01.cir.0000048894.99865.02.
- Holbert CE, Dunworth M, Foley JR, Dunston TT, Stewart TM, Casero RA Jr. Autophagy induction by exogenous polyamines is an artifact of bovine serum amine oxidase activity in culture serum. J Biol Chem. 2020;295:9061–8. https://doi.org/10.1074/jbc.RA120.013867.
- 127. Sun J, Xu J, Liu Y, Lin Y, Wang F, Han Y, Zhang S, Gao X, Xu C, Yuan H. Exogenous spermidine alleviates diabetic cardiomyopathy via suppressing reactive oxygen species, endoplasmic reticulum stress, and Pannexin-1-mediated ferroptosis. Biomol Biomed. 2023;23:825–37. https://doi.org/10.17305/bb.2022.8846.
- 128. Martinelli O, Peruzzi M, Bartimoccia S, D'Amico A, Marchitti S, Rubattu S, Chiariello GA, D'Ambrosio L, Schiavon S, Miraldi F, Saade W, D'Abramo M, Pingitore A, Loffredo L, Nocella C, Forte M, Pignatelli P (2022) Natural Activators of Autophagy Increase Maximal Walking Distance and Reduce Oxidative Stress in Patients with Peripheral Artery Disease: A Pilot Study. Antioxidants (Basel). 11. https://doi.org/10.3390/antiox1109 1836.
- 129. Abdellatif M, Madeo F, Kroemer G, Sedej S. Spermidine overrides INSR (insulin receptor)-IGF1R (insulin-like growth factor 1 receptor)-mediated inhibition of autophagy in the aging heart. Autophagy. 2022;18:2500–2. https://doi.org/10.1080/15548627.2022.2095835.
- 130. (2023) 2023 Alzheimer's disease facts and figures. Alzheimers Dement. 19:1598–695. https://doi.org/10.1002/alz.13016.
- Atri A. The Alzheimer's Disease Clinical Spectrum: Diagnosis and Management. Med Clin North Am. 2019;103:263–93. https://doi.org/10. 1016/j.mcna.2018.10.009.
- 132. Lane CA, Hardy J, Schott JM. Alzheimer's disease. Eur J Neurol. 2018;25:59–70. https://doi.org/10.1111/ene.13439.
- Soria Lopez JA, González HM, Léger GC. Alzheimer's disease. Handb Clin Neurol. 2019;167:231–55. https://doi.org/10.1016/b978-0-12-804766-8. 00013-3.
- 134. Reiman EM, Quiroz YT, Fleisher AS, Chen K, Velez-Pardo C, Jimenez-Del-Rio M, Fagan AM, Shah AR, Alvarez S, Arbelaez A, Giraldo M, Acosta-Baena N, Sperling RA, Dickerson B, Stern CE, Tirado V, Munoz C, Reiman RA, Huentelman MJ, Alexander GE, Langbaum JB, Kosik KS, Tariot PN, Lopera F. Brain imaging and fluid biomarker analysis in young adults at genetic risk for autosomal dominant Alzheimer's disease in the presenilin 1 E280A kindred: a case-control study. Lancet Neurol. 2012;11:1048–56. https://doi.org/10.1016/s1474-4422(12)70228-4.
- Braak H, Thal DR, Ghebremedhin E, Del Tredici K. Stages of the pathologic process in Alzheimer disease: age categories from 1 to 100 years. J Neuropathol Exp Neurol. 2011;70:960–9. https://doi.org/10.1097/NEN. 0b013e318232a379.
- Arranz AM, De Strooper B. The role of astroglia in Alzheimer's disease: pathophysiology and clinical implications. Lancet Neurol. 2019;18:406– 14. https://doi.org/10.1016/s1474-4422(18)30490-3.
- 137. Jo S, Yarishkin O, Hwang YJ, Chun YE, Park M, Woo DH, Bae JY, Kim T, Lee J, Chun H, Park HJ, Lee DY, Hong J, Kim HY, Oh SJ, Park SJ, Lee H, Yoon BE, Kim Y, Jeong Y, Shim I, Bae YC, Cho J, Kowall NW, Ryu H, Hwang E, Kim D, Lee CJ. GABA from reactive astrocytes impairs memory in mouse models of Alzheimer's disease. Nat Med. 2014;20:886–96. https://doi.org/10.1038/nm.3639.
- 138. Park JH, Ju YH, Choi JW, Song HJ, Jang BK, Woo J, Chun H, Kim HJ, Shin SJ, Yarishkin O, Jo S, Park M, Yeon SK, Kim S, Kim J, Nam MH, Londhe AM, Kim J, Cho SJ, Cho S, Lee C, Hwang SY, Kim SW, Oh SJ, Cho J, Pae AN, Lee CJ, Park KD. Newly developed reversible MAO-B inhibitor circumvents the shortcomings of irreversible inhibitors in Alzheimer's disease. Sci Adv. 2019;5:eaav0316. https://doi.org/10. 1126/sciadv.aav0316.

- Stern J, Pier J, Litonjua AA. Asthma epidemiology and risk factors. Semin Immunopathol. 2020;42:5–15. https://doi.org/10.1007/ s00281-020-00785-1.
- 140. Porsbjerg C, Melén E, Lehtimäki L, Shaw D. Asthma Lancet. 2023;401:858–73. https://doi.org/10.1016/s0140-6736(22)02125-0.
- Kurosawa M, Shimizu Y, Tsukagoshi H, Ueki M. Elevated levels of peripheral-blood, naturally occurring aliphatic polyamines in bronchial asthmatic patients with active symptoms. Allergy. 1992;47:638– 43. https://doi.org/10.1111/j.1398-9995.1992.tb02388.x.
- 142. Zimmermann N, King NE, Laporte J, Yang M, Mishra A, Pope SM, Muntel EE, Witte DP, Pegg AA, Foster PS, Hamid Q, Rothenberg ME. Dissection of experimental asthma with DNA microarray analysis identifies arginase in asthma pathogenesis. J Clin Invest. 2003;111:1863–74. https://doi.org/10.1172/jci17912.
- 143. Jain V (2018) Role of Polyamines in Asthma Pathophysiology. Med Sci (Basel). 6. https://doi.org/10.3390/medsci6010004.
- 144. Barnes PJ. Immunology of asthma and chronic obstructive pulmonary disease. Nat Rev Immunol. 2008;8:183–92. https://doi.org/10. 1038/nri2254.
- Kurosawa M, Uno D, Kobayashi S. Naturally occurring aliphatic polyamines-induced histamine release from rat peritoneal mast cells. Allergy. 1991;46:349–54. https://doi.org/10.1111/j.1398-9995.1991. tb00597.x.
- 146. Ilmarinen P, Moilanen E, Erjefält JS, Kankaanranta H. The polyamine spermine promotes survival and activation of human eosinophils. J Allergy Clin Immunol. 2015;136:482–4.e11. https://doi.org/10.1016/j. jaci.2014.12.1922.
- Blüher M. Obesity: global epidemiology and pathogenesis. Nat Rev Endocrinol. 2019;15:288–98. https://doi.org/10.1038/s41574-019-0176-8.
- Fontaine KR, Redden DT, Wang C, Westfall AO, Allison DB. Years of life lost due to obesity. JAMA. 2003;289:187–93. https://doi.org/10.1001/ jama.289.2.187.
- 149. Berrington de Gonzalez A, Hartge P, Cerhan JR, Flint AJ, Hannan L, MacInnis RJ, Moore SC, Tobias GS, Anton-Culver H, Freeman LB, Beeson WL, Clipp SL, English DR, Folsom AR, Freedman DM, Giles G, Hakansson N, Henderson KD, Hoffman-Bolton J, Hoppin JA, Koenig KL, Lee IM, Linet MS, Park Y, Pocobelli G, Schatzkin A, Sesso HD, Weiderpass E, Willcox BJ, Wolk A, Zeleniuch-Jacquotte A, Willett WC, Thun MJ. Body-mass index and mortality among 1.46 million white adults. N Engl J Med. 2010;363:2211–9. https://doi.org/10.1056/NEJMoa1000367.
- Lin X, Li H. Obesity: Epidemiology, Pathophysiology, and Therapeutics. Front Endocrinol (Lausanne). 2021;12:706978. https://doi.org/10.3389/ fendo.2021.706978.
- Nakatani S, Horimoto Y, Nakabayashi N, Karasawa M, Wada M, Kobata K (2022) Spermine Suppresses Adipocyte Differentiation and Exerts Anti-Obesity Effects In Vitro and In Vivo. Int J Mol Sci. 23. https://doi.org/10. 3390/ijms231911818.
- 152. Ma L, Ni Y, Wang Z, Tu W, Ni L, Zhuge F, Zheng A, Hu L, Zhao Y, Zheng L, Fu Z. Spermidine improves gut barrier integrity and gut microbiota function in diet-induced obese mice. Gut Microbes. 2020;12:1–19. https://doi.org/10.1080/19490976.2020.1832857.
- 153. Wang D, Yin J, Zhou Z, Tao Y, Jia Y, Jie H, Zhao J, Li R, Li Y, Guo C, Zhu F, Mao H, Zhang L, Wang Q. Oral Spermidine Targets Brown Fat and Skeletal Muscle to Mitigate Diet-Induced Obesity and Metabolic Disorders. Mol Nutr Food Res. 2021;65:e2100315. https://doi.org/10.1002/mnfr. 202100315.
- Ma L, Ni Y, Hu L, Zhao Y, Zheng L, Yang S, Ni L, Fu Z. Spermidine ameliorates high-fat diet-induced hepatic steatosis and adipose tissue inflammation in preexisting obese mice. Life Sci. 2021;265:118739. https://doi. org/10.1016/j.lfs.2020.118739.
- Szatmary P, Grammatikopoulos T, Cai W, Huang W, Mukherjee R, Halloran C, Beyer G, Sutton R. Acute Pancreatitis: Diagnosis and Treatment. Drugs. 2022;82:1251–76. https://doi.org/10.1007/s40265-022-01766-4.
- Wang GJ, Gao CF, Wei D, Wang C, Ding SQ. Acute pancreatitis: etiology and common pathogenesis. World J Gastroenterol. 2009;15:1427–30. https://doi.org/10.3748/wjg.15.1427.
- Löser C, Fölsch UR, Cleffmann U, Nustede R, Creutzfeldt W. Role of ornithine decarboxylase and polyamines in camostate (Foy-305)-induced pancreatic growth in rats. Digestion. 1989;43:98–112. https://doi.org/10. 1159/000199867.

- Rasanen TL, Alhonen L, Sinervirta R, Keinanen T, Herzig KH, Suppola S, Khomutov AR, Vepsalainen J, Janne J. A polyamine analogue prevents acute pancreatitis and restores early liver regeneration in transgenic rats with activated polyamine catabolism. J Biol Chem. 2002;277:39867–72. https://doi.org/10.1074/jbc.M205967200.
- Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics, 2022. CA Cancer J Clin. 2022;72:7–33. https://doi.org/10.3322/caac.21708.
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. CA Cancer J Clin. 2019;69:7–34. https://doi.org/10.3322/caac.21551.
- Lawton F, Griffin M, Slack J, Blackledge G. Urinary polyamine excretion patterns in patients with epithelial ovarian cancer. Gynecol Obstet Invest. 1989;28:212–4. https://doi.org/10.1159/000293581.
- Russell DH. Increased polyamine concentrations in the urine of human cancer patients. Nat New Biol. 1971;233:144–5. https://doi.org/10.1038/ newbio233144a0.
- Waalkes TP, Rosenshein NB, Shaper JH, Ettinger DS, Woo KB, Paone JF, Gehrke CW. A feasibility study in the development of biological markers for ovarian cancer. J Surg Oncol. 1982;21:207–14. https://doi.org/10. 1002/jso.2930210403.
- Suh JW, Lee SH, Chung BC, Park J. Urinary polyamine evaluation for effective diagnosis of various cancers. J Chromatogr B Biomed Sci Appl. 1997;688:179–86. https://doi.org/10.1016/s0378-4347(96)00266-6.
- 165. Fahrmann JF, Irajizad E, Kobayashi M, Vykoukal J, Dennison JB, Murage E, Wu R, Long JP, Do KA, Celestino J, Lu KH, Lu Z, Bast RC, Hanash S (2021) A MYC-Driven Plasma Polyamine Signature for Early Detection of Ovarian Cancer. Cancers (Basel). 13. https://doi.org/10.3390/cancers130 40913.
- 166. Fortner RT, Schock H, Le Cornet C, Husing A, Vitonis AF, Johnson TS, Fichorova RN, Fashemi T, Yamamoto HS, Tjonneland A, Hansen L, Overvad K, Boutron-Ruault MC, Kvaskoff M, Severi G, Boeing H, Trichopoulou A, Papatesta EM, La Vecchia C, Palli D, Sieri S, Tumino R, Sacerdote C, Mattiello A, Onland-Moret NC, Peeters PH, Bueno-de-Mesquita HBA, Weiderpass E, Quiros JR, Duell EJ, Sanchez MJ, Navarro C, Ardanaz E, Larranaga N, Nodin B, Jirstrom K, Idahl A, Lundin E, Khaw KT, Travis RC, Gunter M, Johansson M, Dossus L, Merritt MA, Riboli E, Terry KL, Cramer DW, Kaaks R. Ovarian cancer early detection by circulating CA125 in the context of anti-CA125 autoantibody levels: Results from the EPIC cohort. Int J Cancer. 2018;142:1355–60. https://doi.org/10.1002/ijc. 31164.
- Torimura T, Iwamoto H. Optimizing the management of intermediatestage hepatocellular carcinoma: Current trends and prospects. Clin Mol Hepatol. 2021;27:236–45. https://doi.org/10.3350/cmh.2020.0204.
- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68:394–424. https://doi.org/10.3322/caac.21492.
- 169. Chen Q, Zheng W, Guan J, Liu H, Dan Y, Zhu L, Song Y, Zhou Y, Zhao X, Zhang Y, Bai Y, Pan Y, Zhang J, Shao C. SOCS2-enhanced ubiquitination of SLC7A11 promotes ferroptosis and radiosensitization in hepatocellular carcinoma. Cell Death Differ. 2023;30:137–51. https://doi.org/10. 1038/s41418-022-01051-7.
- 170. Fitzmaurice C, Abate D, Abbasi N, Abbastabar H, Abd-Allah F, Abdel-Rahman O, Abdelalim A, Abdoli A, Abdollahpour I, Abdulle ASM, Abebe ND, Abraha HN, Abu-Raddad LJ, Abualhasan A, Adedeji IA, Advani SM, Afarideh M, Afshari M, Aghaali M, Agius D, Agrawal S, Ahmadi A, Ahmadian E, Ahmadpour E, Ahmed MB, Akbari ME, Akinyemiju T, Al-Aly Z, AlAbdulKader AM, Alahdab F, Alam T, Alamene GM, Alemnew BTT, Alene KA, Alinia C, Alipour V, Aljunid SM, Bakeshei FA, Almadi MAH, Almasi-Hashiani A, Alsharif U, Alsowaidi S, Alvis-Guzman N, Amini E, Amini S, Amoako YA, Anbari Z, Anber NH, Andrei CL, Anjomshoa M, Ansari F, Ansariadi A, Appiah SCY, Arab-Zozani M, Arabloo J, Arefi Z, Aremu O, Areri HA, Artaman A, Asayesh H, Asfaw ET, Ashagre AF, Assadi R, Ataeinia B, Atalay HT, Ataro Z, Atique S, Ausloos M, Avila-Burgos L, Avokpaho E, Awasthi A, Awoke N, Ayala Quintanilla BP, Ayanore MA, Ayele HT, Babaee E, Bacha U, Badawi A, Bagherzadeh M, Bagli E, Balakrishnan S, Balouchi A, Bärnighausen TW, Battista RJ, Behzadifar M, Behzadifar M, Bekele BB, Belay YB, Belayneh YM, Berfield KKS, Berhane A, Bernabe E, Beuran M, Bhakta N, Bhattacharyya K, Biadgo B, Bijani A, Bin Sayeed MS, Birungi C, Bisignano C, Bitew H, Bjørge T, Bleyer A, Bogale KA, Bojia HA, Borzì AM, Bosetti C, Bou-Orm IR, Brenner H, Brewer JD, Briko AN, Briko NI, Bustamante-Teixeira MT, Butt ZA, Carreras G,

Carrero JJ, Carvalho F, Castro C, Castro F, Catalá-López F, Cerin E, Chaiah Y, Chanie WF, Chattu VK, Chaturvedi P, Chauhan NS, Chehrazi M, Chiang PP, Chichiabellu TY, Chido-Amajuoyi OG, Chimed-Ochir O, Choi JJ, Christopher DJ, Chu DT, Constantin MM, Costa VM, Crocetti E, Crowe CS, Curado MP, Dahlawi SMA, Damiani G, Darwish AH, Daryani A, das Neves J, Demeke FM, Demis AB, Demissie BW, Demoz GT, Denova-Gutiérrez E, Derakhshani A, Deribe KS, Desai R, Desalegn BB, Desta M, Dey S, Dharmaratne SD, Dhimal M, Diaz D, Dinberu MTT, Djalalinia S, Doku DT, Drake TM, Dubey M, Dubljanin E, Duken EE, Ebrahimi H, Effiong A, Eftekhari A, El Sayed I, Zaki MES, El-Jaafary SI, El-Khatib Z, Elemineh DA, Elkout H, Ellenbogen RG, Elsharkawy A, Emamian MH, Endalew DA, Endries AY, Eshrati B, Fadhil I, Fallah Omrani V, Faramarzi M, Farhangi MA, Farioli A, Farzadfar F, Fentahun N, Fernandes E, Feyissa GT, Filip I, Fischer F, Fisher JL, Force LM, Foroutan M, Freitas M, Fukumoto T, Futran ND, Gallus S, Gankpe FG, Gayesa RT, Gebrehiwot TT, Gebremeskel GG, Gedefaw GA, Gelaw BK, Geta B, Getachew S, Gezae KE, Ghafourifard M, Ghajar A, Ghashqhaee A, Gholamian A, Gill PS, Ginindza TTG, Girmay A, Gizaw M, Gomez RS, Gopalani SV, Gorini G, Goulart BNG, Grada A, Ribeiro Guerra M, Guimaraes ALS, Gupta PC, Gupta R, Hadkhale K, Haj-Mirzaian A, Haj-Mirzaian A, Hamadeh RR, Hamidi S, Hanfore LK, Haro JM, Hasankhani M, Hasanzadeh A, Hassen HY, Hay RJ, Hay SI, Henok A, Henry NJ, Herteliu C, Hidru HD, Hoang CL, Hole MK, Hoogar P, Horita N, Hosgood HD, Hosseini M, Hosseinzadeh M, Hostiuc M, Hostiuc S, Househ M, Hussen MM, Ileanu B, Ilic MD, Innos K, Irvani SSN, Iseh KR, Islam SMS, Islami F, Jafari Balalami N, Jafarinia M, Jahangiry L, Jahani MA, Jahanmehr N, Jakovljevic M, James SL, Javanbakht M, Jayaraman S, Jee SH, Jenabi E, Jha RP, Jonas JB, Jonnagaddala J, Joo T, Jungari SB, Jürisson M, Kabir A, Kamangar F, Karch A, Karimi N, Karimian A, Kasaeian A, Kasahun GG, Kassa B, Kassa TD, Kassaw MW, Kaul A, Keiyoro PN, Kelbore AG, Kerbo AA, Khader YS, Khalilarjmandi M, Khan EA, Khan G, Khang YH, Khatab K, Khater A, Khayamzadeh M, Khazaee-Pool M, Khazaei S, Khoja AT, Khosravi MH, Khubchandani J, Kianipour N, Kim D, Kim YJ, Kisa A, Kisa S, Kissimova-Skarbek K, Komaki H, Koyanagi A, Krohn KJ, Bicer BK, Kugbey N, Kumar V, Kuupiel D, La Vecchia C, Lad DP, Lake EA, Lakew AM, Lal DK, Lami FH, Lan Q, Lasrado S, Lauriola P, Lazarus JV, Leigh J, Leshargie CT, Liao Y, Limenih MA, Listl S, Lopez AD, Lopukhov PD, Lunevicius R, Madadin M, Magdeldin S, El Razek HMA, Majeed A, Maleki A, Malekzadeh R, Manafi A, Manafi N, Manamo WA, Mansourian M, Mansournia MA, Mantovani LG, Maroufizadeh S, Martini SMS, Mashamba-Thompson TP, Massenburg BB, Maswabi MT, Mathur MR, McAlinden C, McKee M, Meheretu HAA, Mehrotra R, Mehta V, Meier T, Melaku YA, Meles GG, Meles HG, Melese A, Melku M, Memiah PTN, Mendoza W, Menezes RG, Merat S, Meretoja TJ, Mestrovic T, Miazgowski B, Miazgowski T, Mihretie KMM, Miller TR, Mills EJ, Mir SM, Mirzaei H, Mirzaei HR, Mishra R, Moazen B, Mohammad DK, Mohammad KA, Mohammad Y, Darwesh AM, Mohammadbeigi A, Mohammadi H, Mohammadi M, Mohammadian M, Mohammadian-Hafshejani A, Mohammadoo-Khorasani M, Mohammadpourhodki R, Mohammed AS, Mohammed JA, Mohammed S, Mohebi F, Mokdad AH, Monasta L, Moodley Y, Moosazadeh M, Moossavi M, Moradi G, Moradi-Joo M, Moradi-Lakeh M, Moradpour F, Morawska L, Morgado-da-Costa J, Morisaki N, Morrison SD, Mosapour A, Mousavi SM, Muche AA, Muhammed OSS, Musa J, Nabhan AF, Naderi M, Nagarajan AJ, Nagel G, Nahvijou A, Naik G, Najafi F, Naldi L, Nam HS, Nasiri N, Nazari J, Negoi I, Neupane S, Newcomb PA, Nggada HA, Ngunjiri JW, Nguyen CT, Nikniaz L, Ningrum DNA, Nirayo YL, Nixon MR, Nnaji CA, Nojomi M, Nosratnejad S, Shiadeh MN, Obsa MS, Ofori-Asenso R, Ogbo FA, Oh IH, Olagunju AT, Olagunju TO, Oluwasanu MM, Omonisi AE, Onwujekwe OE, Oommen AM, Oren E, Ortega-Altamirano DDV, Ota E, Otstavnov SS, Owolabi MO, P AM, Padubidri JR, Pakhale S, Pakpour AH, Pana A, Park EK, Parsian H, Pashaei T, Patel S, Patil ST, Pennini A, Pereira DM, Piccinelli C, Pillay JD, Pirestani M, Pishgar F, Postma MJ, Pourjafar H, Pourmalek F, Pourshams A, Prakash S, Prasad N, Qorbani M, Rabiee M, Rabiee N, Radfar A, Rafiei A, Rahim F, Rahimi M, Rahman MA, Rajati F, Rana SM, Raoofi S, Rath GK, Rawaf DL, Rawaf S, Reiner RC, Renzaho AMN, Rezaei N, Rezapour A, Ribeiro AI, Ribeiro D, Ronfani L, Roro EM, Roshandel G, Rostami A, Saad RS, Sabbagh P, Sabour S, Saddik B, Safiri S, Sahebkar A, Salahshoor MR, Salehi F, Salem H, Salem MR, Salimzadeh H, Salomon JA, Samy AM, Sanabria J, Santric Milicevic MM, Sartorius B, Sarveazad A, Sathian B, Satpathy M, Savic M, Sawhney M, Sayyah M, Schneider IJC, Schöttker B, Sekerija M, Sepanlou SG, Sepehrimanesh M, Seyedmousavi S, Shaahmadi F, Shabaninejad H, Shahbaz M, Shaikh MA,

Shamshirian A, Shamsizadeh M, Sharafi H, Sharafi Z, Sharif M, Sharifi A, Sharifi H, Sharma R, Sheikh A, Shirkoohi R, Shukla SR, Si S, Siabani S, Silva DAS, Silveira DGA, Singh A, Singh JA, Sisay S, Sitas F, Sobngwi E, Soofi M, Soriano JB, Stathopoulou V, Sufiyan MB, Tabarés-Seisdedos R, Tabuchi T, Takahashi K, Tamtaji OR, Tarawneh MR, Tassew SG, Taymoori P, Tehrani-Banihashemi A, Temsah MH, Temsah O, Tesfay BE, Tesfay FH, Teshale MY, Tessema GA, Thapa S, Tlaye KG, Topor-Madry R, Tovani-Palone MR, Traini E, Tran BX, Tran KB, Tsadik AG, Ullah I, Uthman OA, Vacante M, Vaezi M, Varona Pérez P, Veisani Y, Vidale S, Violante FS, Vlassov V, Vollset SE, Vos T, Vosoughi K, Vu GT, Vujcic IS, Wabinga H, Wachamo TM, Wagnew FS, Waheed Y, Weldegebreal F, Weldesamuel GT, Wijeratne T, Wondafrash DZ, Wonde TE, Wondmieneh AB, Workie HM, Yadav R, Yadegar A, Yadollahpour A, Yaseri M, Yazdi-Feyzabadi V, Yeshaneh A, Yimam MA Yimer EM, Yisma E, Yonemoto N, Younis MZ, Yousefi B, Yousefifard M, Yu C, Zabeh E, Zadnik V, Moghadam TZ, Zaidi Z, Zamani M, Zandian H, Zangeneh A, Zaki L, Zendehdel K, Zenebe ZM, Zewale TA, Ziapour A, Zodpey S. Murray C.I., Global, Regional, and National Cancer Incidence. Mortality, Years of Life Lost, Years Lived With Disability, and Disability-Adjusted Life-Years for 29 Cancer Groups, 1990 to 2017: A Systematic Analysis for the Global Burden of Disease Study. JAMA Oncol. 2019;5:1749-68. https://doi.org/10.1001/jamaoncol.2019.2996.

- Nitta T, Igarashi K, Yamamoto N. Polyamine depletion induces apoptosis through mitochondria-mediated pathway. Exp Cell Res. 2002;276:120– 8. https://doi.org/10.1006/excr.2002.5517.
- Pascale RM, Simile MM, Gaspa L, Daino L, Seddaiu MA, Pinna G, Carta M, Zolo P, Feo F. Alterations of ornithine decarboxylase gene during the progression of rat liver carcinogenesis. Carcinogenesis. 1993;14:1077– 80. https://doi.org/10.1093/carcin/14.5.1077.
- 173. Feo F, Garcea R, Daino L, Pascale R, Pirisi L, Frassetto S, Ruggiu ME. Early stimulation of polyamine biosynthesis during promotion by phenobarbital of diethylnitrosamine-induced rat liver carcinogenesis. The effects of variations of the S-adenosyl-L-methionine cellular pool. Carcinogenesis. 1985;6:1713–20. https://doi.org/10.1093/carcin/6.12.1713.
- 174. Pascale RM, Simile MM, De Miglio MR, Nufris A, Daino L, Seddaiu MA, Rao PM, Rajalakshmi S, Sarma DS, Feo F. Chemoprevention by S-adenosyl-L-methionine of rat liver carcinogenesis initiated by 1,2-dimethylhydrazine and promoted by orotic acid. Carcinogenesis. 1995;16:427–30. https://doi.org/10.1093/carcin/16.2.427.
- Lu SC, Mato JM. S-adenosylmethionine in liver health, injury, and cancer. Physiol Rev. 2012;92:1515–42. https://doi.org/10.1152/physrev.00047.2011.
- Liu L, Xie R, Yang C, McKeehan WL. Dual function microtubule- and mitochondria-associated proteins mediate mitotic cell death. Cell Oncol. 2009;31:393–405. https://doi.org/10.3233/clo-2009-0484.
- 177. Yue F, Li W, Zou J, Chen Q, Xu G, Huang H, Xu Z, Zhang S, Gallinari P, Wang F, McKeehan WL, Liu L. Blocking the association of HDAC4 with MAP1S accelerates autophagy clearance of mutant Huntingtin. Aging (Albany NY). 2015;7:839–53. https://doi.org/10.18632/aging.100818.
- 178. Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer Statistics, 2021. CA Cancer J Clin. 2021;71:7–33. https://doi.org/10.3322/caac.21654.
- 179. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin. 2021;71:209–49. https://doi.org/10.3322/caac.21660.
- Guan WL, He Y, Xu RH. Gastric cancer treatment: recent progress and future perspectives. J Hematol Oncol. 2023;16:57. https://doi.org/10. 1186/s13045-023-01451-3.
- 181. Gobert AP, Al-Greene NT, Singh K, Coburn LA, Sierra JC, Verriere TG, Luis PB, Schneider C, Asim M, Allaman MM, Barry DP, Cleveland JL, Destefano Shields CE, Casero RA Jr, Washington MK, Piazuelo MB, Wilson KT. Distinct Immunomodulatory Effects of Spermine Oxidase in Colitis Induced by Epithelial Injury or Infection. Front Immunol. 2018;9:1242. https://doi.org/10.3389/fimmu.2018.01242.
- 182. Xu H, Chaturvedi R, Cheng Y, Bussiere FI, Asim M, Yao MD, Potosky D, Meltzer SJ, Rhee JG, Kim SS, Moss SF, Hacker A, Wang Y, Casero RA Jr, Wilson KT. Spermine oxidation induced by Helicobacter pylori results in apoptosis and DNA damage: implications for gastric carcinogenesis. Cancer Res. 2004;64:8521–5. https://doi.org/10.1158/0008-5472. Can-04-3511.
- Chaturvedi R, de Sablet T, Peek RM, Wilson KT. Spermine oxidase, a polyamine catabolic enzyme that links Helicobacter pylori CagA and

gastric cancer risk. Gut Microbes. 2012;3:48–56. https://doi.org/10.4161/ gmic.19345.

- Xue C, Chu Q, Zheng Q, Yuan X, Su Y, Bao Z, Lu J, Li L. Current understanding of the intratumoral microbiome in various tumors. Cell Rep Med. 2023;4:100884. https://doi.org/10.1016/j.xcrm.2022.100884.
- Meriggi F, Bertocchi P, Zaniboni A. Management of potentially resectable colorectal cancer liver metastases. World J Gastrointest Surg. 2013;5:138–45. https://doi.org/10.4240/wjgs.v5.i5.138.
- 186. Yang Z, Liu S, Zhu M, Zhang H, Wang J, Xu Q, Lin K, Zhou X, Tao M, Li C, Zhu H. PS341 inhibits hepatocellular and colorectal cancer cells through the FOXO3/CTNNB1 signaling pathway. Sci Rep. 2016;6:22090. https://doi.org/10.1038/srep22090.
- Bello-Fernandez C, Packham G, Cleveland JL. The ornithine decarboxylase gene is a transcriptional target of c-Myc. Proc Natl Acad Sci U S A. 1993;90:7804–8. https://doi.org/10.1073/pnas.90.16.7804.
- Groden J, Thliveris A, Samowitz W, Carlson M, Gelbert L, Albertsen H, Joslyn G, Stevens J, Spirio L, Robertson M, et al. Identification and characterization of the familial adenomatous polyposis coli gene. Cell. 1991;66:589–600. https://doi.org/10.1016/0092-8674(81)90021-0.
- Zhang L, Shay JW (2017) Multiple Roles of APC and its Therapeutic Implications in Colorectal Cancer. J Natl Cancer Inst. 109. https://doi. org/10.1093/jnci/djw332.
- Maffeis V, Nicolè L, Cappellesso R. RAS, Cellular Plasticity, and Tumor Budding in Colorectal Cancer. Front Oncol. 2019;9:1255. https://doi.org/ 10.3389/fonc.2019.01255.
- 191. Thompson JS, Edney JA, Laughlin K. Urinary polyamines in colorectal cancer. Dis Colon Rectum. 1986;29:873–7. https://doi.org/10.1007/bf02555367.
- Löser C, Fölsch UR, Paprotny C, Creutzfeldt W. Polyamines in colorectal cancer. Evaluation of polyamine concentrations in the colon tissue, serum, and urine of 50 patients with colorectal cancer. Cancer. 1990;65:958–66. https://doi.org/10.1002/1097-0142(19900215)65:4% 3c958::aid-cncr2820650423%3e3.0.co;2-z.
- 193. Coni S, Serrao SM, Yurtsever ZN, Di Magno L, Bordone R, Bertani C, Licursi V, Ianniello Z, Infante P, Moretti M, Petroni M, Guerrieri F, Fatica A, Macone A, De Smaele E, Di Marcotullio L, Giannini G, Maroder M, Agostinelli E, Canettieri G. Blockade of EIF5A hypusination limits colorectal cancer growth by inhibiting MYC elongation. Cell Death Dis. 2020;11:1045. https://doi.org/10.1038/s41419-020-03174-6.
- 194. Coni S, Bordone R, Ivy DM, Yurtsever ZN, Di Magno L, D'Amico R, Cesaro B, Fatica A, Belardinilli F, Bufalieri F, Maroder M, De Smaele E, Di Marcotullio L, Giannini G, Agostinelli E, Canettieri G. Combined inhibition of polyamine metabolism and eIF5A hypusination suppresses colorectal cancer growth through a converging effect on MYC translation. Cancer Lett. 2023;559:216120. https://doi.org/10.1016/j.canlet.2023.216120.
- 195. Cano CE, Sandí MJ, Hamidi T, Calvo EL, Turrini O, Bartholin L, Loncle C, Secq V, Garcia S, Lomberk G, Kroemer G, Urrutia R, Iovanna JL. Homotypic cell cannibalism, a cell-death process regulated by the nuclear protein 1, opposes to metastasis in pancreatic cancer. EMBO Mol Med. 2012;4:964–79. https://doi.org/10.1002/emmm.201201255.
- Ot P. An overview of polyamine metabolism in pancreatic ductal adenocarcinoma. Int J Cancer. 2018;142:1968–76. https://doi.org/10. 1002/ijc.31155.
- Whatcott CJ, Diep CH, Jiang P, Watanabe A, LoBello J, Sima C, Hostetter G, Shepard HM, Von Hoff DD, Han H. Desmoplasia in Primary Tumors and Metastatic Lesions of Pancreatic Cancer. Clin Cancer Res. 2015;21:3561–8. https://doi.org/10.1158/1078-0432.Ccr-14-1051.
- Hayes CS, Shicora AC, Keough MP, Snook AE, Burns MR, Gilmour SK. Polyamine-blocking therapy reverses immunosuppression in the tumor microenvironment. Cancer Immunol Res. 2014;2:274–85. https://doi. org/10.1158/2326-6066.Cir-13-0120-t.
- Hayes CS, Burns MR, Gilmour SK. Polyamine blockade promotes antitumor immunity. Oncoimmunology. 2014;3:e27360. https://doi.org/10. 4161/onci.27360.
- Alexander ET, Minton A, Peters MC, Phanstiel OT, Gilmour SK. A novel polyamine blockade therapy activates an anti-tumor immune response. Oncotarget. 2017;8:84140–52. https://doi.org/10.18632/oncotarget. 20493.
- 201. Kurihara S, Sakai Y, Suzuki H, Muth A, Ot P, Rather PN. Putrescine importer PlaP contributes to swarming motility and urothelial cell

invasion in Proteus mirabilis. J Biol Chem. 2013;288:15668–76. https://doi.org/10.1074/jbc.M113.454090.

- Desforges B, Curmi PA, Bounedjah O, Nakib S, Hamon L, De Bandt JP, Pastré D. An intercellular polyamine transfer via gap junctions regulates proliferation and response to stress in epithelial cells. Mol Biol Cell. 2013;24:1529–43. https://doi.org/10.1091/mbc.E12-10-0729.
- 203. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. CA Cancer J Clin. 2015;65:87–108. https://doi. org/10.3322/caac.21262.
- Litwin MS, Tan HJ. The Diagnosis and Treatment of Prostate Cancer: A Review. JAMA. 2017;317:2532–42. https://doi.org/10.1001/jama.2017. 7248.
- Bistulfi G, Foster BA, Karasik E, Gillard B, Miecznikowski J, Dhiman VK, Smiraglia DJ. Dietary folate deficiency blocks prostate cancer progression in the TRAMP model. Cancer Prev Res (Phila). 2011;4:1825–34. https://doi.org/10.1158/1940-6207.Capr-11-0140.
- Affronti HC, Long MD, Rosario SR, Gillard BM, Karasik E, Boerlin CS, Pellerite AJ, Foster BA, Attwood K, Pili R, Wilton JH, Campbell MJ, Smiraglia DJ. Dietary folate levels alter the kinetics and molecular mechanism of prostate cancer recurrence in the CWR22 model. Oncotarget. 2017;8:103758–74. https://doi.org/10.18632/oncotarget.21911.
- Bistulfi G, Affronti HC, Foster BA, Karasik E, Gillard B, Morrison C, Mohler J, Phillips JG, Smiraglia DJ. The essential role of methylthioadenosine phosphorylase in prostate cancer. Oncotarget. 2016;7:14380–93. https://doi.org/10.18632/oncotarget.7486.
- Li X, Li F, Ye F, Guo H, Chen W, Jin J, Wang Y, Dai P, Shi H, Tao H, Dang W, Ding Y, Wang M, Jiang H, Chen K, Zhang N, Gao D, Zhang Y, Luo C. Spermine is a natural suppressor of AR signaling in castration-resistant prostate cancer. Cell Rep. 2023;42:112798. https://doi.org/10.1016/j. celrep.2023.112798.
- Brodeur GM, Seeger RC, Schwab M, Varmus HE, Bishop JM. Amplification of N-myc in untreated human neuroblastomas correlates with advanced disease stage. Science. 1984;224:1121–4. https://doi.org/10. 1126/science.6719137.
- Seeger RC, Brodeur GM, Sather H, Dalton A, Siegel SE, Wong KY, Hammond D. Association of multiple copies of the N-myc oncogene with rapid progression of neuroblastomas. N Engl J Med. 1985;313:1111–6. https://doi.org/10.1056/nejm198510313131802.
- 211. Auvinen M, Paasinen A, Andersson LC, Hölttä E. Ornithine decarboxylase activity is critical for cell transformation. Nature. 1992;360:355–8. https://doi.org/10.1038/360355a0.
- Schultz CR, Swanson MA, Dowling TC, Bachmann AS. Probenecid increases renal retention and antitumor activity of DFMO in neuroblastoma. Cancer Chemother Pharmacol. 2021;88:607–17. https://doi.org/ 10.1007/s00280-021-04309-y.
- Lodeserto P, Rossi M, Blasi P, Farruggia G, Orienti I (2022) Nanospermidine in Combination with Nanofenretinide Induces Cell Death in Neuroblastoma Cell Lines. Pharmaceutics. 14. https://doi.org/10.3390/ pharmaceutics14061215.
- 214. Akinyele O, Wallace HM (2022) Understanding the Polyamine and mTOR Pathway Interaction in Breast Cancer Cell Growth. Med Sci (Basel). 10. https://doi.org/10.3390/medsci10030051.
- 215. Thomas T, Thomas TJ. Polyamine metabolism and cancer. J Cell Mol Med. 2003;7:113–26. https://doi.org/10.1111/j.1582-4934.2003.tb00210.x.
- 216. Linsalata M, Russo F. Nutritional factors and polyamine metabolism in colorectal cancer. Nutrition. 2008;24:382–9. https://doi.org/10.1016/j. nut.2007.12.014.
- Bachmann AS, Geerts D. Polyamine synthesis as a target of MYC oncogenes. J Biol Chem. 2018;293:18757–69. https://doi.org/10.1074/jbc. TM118.003336.
- 218. Bello-Fernandez C, Cleveland JL. c-myc transactivates the ornithine decarboxylase gene. Curr Top Microbiol Immunol. 1992;182:445–52. https://doi.org/10.1007/978-3-642-77633-5_56.
- Forshell TP, Rimpi S, Nilsson JA. Chemoprevention of B-cell lymphomas by inhibition of the Myc target spermidine synthase. Cancer Prev Res (Phila). 2010;3:140–7. https://doi.org/10.1158/1940-6207.CAPR-09-0166.
- Marinkovic D, Marinkovic T, Kokai E, Barth T, Moller P, Wirth T. Identification of novel Myc target genes with a potential role in lymphomagenesis. Nucleic Acids Res. 2004;32:5368–78. https://doi.org/10.1093/nar/ gkh877.

- 221. Funakoshi-Tago M, Sumi K, Kasahara T, Tago K. Critical roles of Myc-ODC axis in the cellular transformation induced by myeloproliferative neoplasm-associated JAK2 V617F mutant. PLoS One. 2013;8:e52844. https://doi.org/10.1371/journal.pone.0052844.
- Ho K, Valdez F, Garcia R, Tirado CA. JAK2 Translocations in hematological malignancies: Review of the literature. J Assoc Genet Technol. 2010;36:107–9.
- 223. Koomoa DL, Geerts D, Lange I, Koster J, Pegg AE, Feith DJ, Bachmann AS. DFMO/eflornithine inhibits migration and invasion downstream of MYCN and involves p27Kip1 activity in neuroblastoma. Int J Oncol. 2013;42:1219–28. https://doi.org/10.3892/ijo.2013.1835.
- 224. Perez-Riesgo E, Hernando-Perez E, Feijoo V, Tajada S, Nunez L, Villalobos C (2023) Transcriptional Basis of Ca(2+) Remodeling Reversal Induced by Polyamine Synthesis Inhibition in Colorectal Cancer Cells. Cancers (Basel). 15. https://doi.org/10.3390/cancers15051600.
- Ozfiliz P, Kizilboga T, Demir S, Alkurt G, Palavan-Unsal N, Arisan ED, Dinler-Doganay G. Bag-1 promotes cell survival through c-Myc-mediated ODC upregulation that is not preferred under apoptotic stimuli in MCF-7 cells. Cell Biochem Funct. 2015;33:293–307. https://doi.org/10.1002/cbf.3114.
- 226. Fahrmann JF, Vykoukal J, Fleury A, Tripathi S, Dennison JB, Murage E, Wang P, Yu CY, Capello M, Creighton CJ, Do KA, Long JP, Irajizad E, Peterson C, Katayama H, Disis ML, Arun B, Hanash S. Association Between Plasma Diacetylspermine and Tumor Spermine Synthase With Outcome in Triple-Negative Breast Cancer. J Natl Cancer Inst. 2020;112:607–16. https://doi.org/10.1093/jnci/djz182.
- 227. Kastenhuber ER, Lowe SW. Putting p53 in Context. Cell. 2017;170:1062– 78. https://doi.org/10.1016/j.cell.2017.08.028.
- Li L, Mao Y, Zhao L, Li L, Wu J, Zhao M, Du W, Yu L, Jiang P. p53 regulation of ammonia metabolism through urea cycle controls polyamine biosynthesis. Nature. 2019;567:253–6. https://doi.org/10.1038/s41586-019-0996-7.
- 229. Li J, Meng Y, Wu X, Sun Y. Polyamines and related signaling pathways in cancer. Cancer Cell Int. 2020;20:539. https://doi.org/10.1186/s12935-020-01545-9.
- Chae YB, Kim MM. Activation of p53 by spermine mediates induction of autophagy in HT1080 cells. Int J Biol Macromol. 2014;63:56–63. https:// doi.org/10.1016/j.ijbiomac.2013.10.041.
- 231. Sabapathy K, Lane DP. Therapeutic targeting of p53: all mutants are equal, but some mutants are more equal than others. Nat Rev Clin Oncol. 2018;15:13–30. https://doi.org/10.1038/nrclinonc.2017.151.
- 232. Olivier M, Hollstein M, Hainaut P. TP53 mutations in human cancers: origins, consequences, and clinical use. Cold Spring Harb Perspect Biol. 2010;2:a001008. https://doi.org/10.1101/cshperspect.a001008.
- Laplante M, Sabatini DM. mTOR signaling in growth control and disease. Cell. 2012;149:274–93. https://doi.org/10.1016/j.cell.2012.03.017.
- Blenis J. TOR, the Gateway to Cellular Metabolism, Cell Growth, and Disease. Cell. 2017;171:10–3. https://doi.org/10.1016/j.cell.2017.08.019.
 Han J, Wang Y. mTORC1 signaling in hepatic lipid metabolism. Protein
- Cell. 2018;9:145–51. https://doi.org/10.1007/s13238-017-0409-3.
 236. Fingar DC, Richardson CJ, Tee AR, Cheatham L, Tsou C, Blenis J. mTOR
- controls cell cycle progression through its cell growth effectors S6K1 and 4E-BP1/eukaryotic translation initiation factor 4E. Mol Cell Biol. 2004;24:200–16. https://doi.org/10.1128/mcb.24.1.200-216.2004.
- Ma XM, Blenis J. Molecular mechanisms of mTOR-mediated translational control. Nat Rev Mol Cell Biol. 2009;10:307–18. https://doi.org/10. 1038/nrm2672.
- Zabala-Letona A, Arruabarrena-Aristorena A, Martín-Martín N, 238. Fernandez-Ruiz S, Sutherland JD, Clasquin M, Tomas-Cortazar J, Jimenez J, Torres I, Quang P, Ximenez-Embun P, Bago R, Ugalde-Olano A, Loizaga-Iriarte A, Lacasa-Viscasillas I, Unda M, Torrano V, Cabrera D, van Liempd SM, Cendon Y, Castro E, Murray S, Revandkar A, Alimonti A, Zhang Y, Barnett A, Lein G, Pirman D, Cortazar AR, Arreal L, Prudkin L, Astobiza I, Valcarcel-Jimenez L, Zuñiga-García P, Fernandez-Dominguez I, Piva M, Caro-Maldonado A, Sánchez-Mosquera P, Castillo-Martín M, Serra V, Beraza N, Gentilella A, Thomas G, Azkargorta M, Elortza F, Farràs R, Olmos D, Efeyan A, Anguita J, Muñoz J, Falcón-Pérez JM, Barrio R, Macarulla T, Mato JM, Martinez-Chantar ML, Cordon-Cardo C, Aransay AM, Marks K, Baselga J, Tabernero J, Nuciforo P, Manning BD, Marjon K, Carracedo A. Corrigendum: mTORC1-dependent AMD1 regulation sustains polyamine metabolism in prostate cancer. Nature. 2018;554:554. https://doi.org/10.1038/nature25470.

- 239. Gomes AP, Schild T, Blenis J. Adding Polyamine Metabolism to the mTORC1 Toolkit in Cell Growth and Cancer. Dev Cell. 2017;42:112–4. https://doi.org/10.1016/j.devcel.2017.07.004.
- Jiang R, Choi W, Hu L, Gerner EW, Hamilton SR, Zhang W. Activation of polyamine catabolism by N1, N11-diethylnorspermine alters the cellular localization of mTOR and downregulates mTOR protein level in glioblastoma cells. Cancer Biol Ther. 2007;6:1644–8. https://doi.org/10. 4161/cbt.6.10.4800.
- Degirmenci U, Wang M, Hu J (2020) Targeting Aberrant RAS/RAF/MEK/ ERK Signaling for Cancer Therapy. Cells. 9. https://doi.org/10.3390/cells 9010198.
- 242. Origanti S, Shantz LM. Ras transformation of RIE-1 cells activates capindependent translation of ornithine decarboxylase: regulation by the Raf/MEK/ERK and phosphatidylinositol 3-kinase pathways. Cancer Res. 2007;67:4834–42. https://doi.org/10.1158/0008-5472.Can-06-4627.
- Novita Sari I, Setiawan T, Seock Kim K, Toni Wijaya Y, Won Cho K, Young Kwon H. Metabolism and function of polyamines in cancer progression. Cancer Lett. 2021;519:91–104. https://doi.org/10.1016/j.canlet.2021.06.020.
- Ignatenko NA, Babbar N, Mehta D, Casero RA Jr, Gerner EW. Suppression of polyamine catabolism by activated Ki-ras in human colon cancer cells. Mol Carcinog. 2004;39:91–102. https://doi.org/10.1002/mc.10166.

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