REVIEW

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Abstract

Background Chronic obstructive pulmonary disease (COPD) is characterized by persistent respiratory symptoms and airflow limitation due to airway and/or alveolar remodeling. Although the abnormalities are primarily prompted by chronic exposure to inhaled irritants, maladjusted and self-reinforcing immune responses are significant contributors to the development and progression of the disease. The p38 isoforms are regarded as pivotal hub proteins that regulate immune and inflammatory responses in both healthy and disease states. As a result, their inhibition has been the subject of numerous recent studies exploring their therapeutic potential in COPD.

Main body We performed a systematic search based on the PRISMA guidelines to find relevant studies about P38 signaling in COPD patients. We searched the PubMed and Google Scholar databases and used "P38" AND "COPD" Mesh Terms. We applied the following inclusion criteria: (1) human, animal, ex vivo and in vitro studies; (2) original research articles; (3) published in English; and (4) focused on P38 signaling in COPD pathogenesis, progression, or treatment. We screened the titles and abstracts of the retrieved studies and assessed the full texts of the eligible studies for quality and relevance. We extracted the following data from each study: authors, year, country, sample size, study design, cell type, intervention, outcome, and main findings. We classified the studies according to the role of different cells and treatments in P38 signaling in COPD.

Conclusion While targeting p38 MAPK has demonstrated some therapeutic potential in COPD, its efficacy is limited. Nevertheless, combining p38 MAPK inhibitors with other anti-inflammatory steroids appears to be a promising treatment choice. Clinical trials testing various p38 MAPK inhibitors have produced mixed results, with some showing improvement in lung function and reduction in exacerbations in COPD patients. Despite these mixed results, research on p38 MAPK inhibitors is still a major area of study to develop new and more effective therapies for COPD. As our understanding of COPD evolves, we may gain a better understanding of how to utilize p38 MAPK inhibitors to treat this disease.

Keywords Chronic Obstructive Pulmonary Disease, COPD, COPD pharmacotherapy, p38 MAPK inhibitors, p38 MAPK signaling, Systematic review

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Plain English summary

We wanted to determine what studies have been done on how a protein called p38 affects a lung disease called COPD. COPD is a condition that makes it hard to breathe and can cause coughing, wheezing, and chest infections. p38 is a protein that helps cells to respond to stress and inflammation, but it may also play a role in causing or worsening COPD. We searched two main online databases for studies that met our criteria. We looked for studies that involved humans, studies that used animals or cells in the lab, studies that reported new findings, studies that were written in English, and studies that focused on p38 and COPD. We found 361 studies that matched our criteria. We read the titles and summaries of these studies and checked the full texts for quality and relevance. We collected information from each study, such as who did it, when and where it was done, how many people were involved, what type of cells were studies based on the type of cells and type of treatment they studied. We found that different types of cells (such as lung cells, immune cells, and blood cells) and different types of treatment can affect how p38 works in COPD.

Background

According to the Global Initiative for Chronic Obstructive Lung Disease (GOLD) description, "Chronic obstructive pulmonary disease (COPD) is a heterogeneous lung condition characterized by chronic respiratory symptoms (dyspnea, cough, sputum production, exacerbations) due to abnormalities of the airways (bronchitis, bronchiolitis) and/or alveoli (emphysema) that cause persistent, often progressive airflow obstruction." [1]. COPD is among the top third causes of death, according to the World Health Organization (WHO) [2]. The disease is characterized by small airway complications such as obstructive bronchiolitis, fibrosis, airway remodeling, and emphysema [3]. The clinical manifestations of COPD vary among different patients [4], and an accelerated decline in lung function is observed in approximately half of patients [5]. This suggests the critical impact of personal distinctive inflammatory signatures and genetic/epigenetic makeup on disease severity [6]. Currently, there is no established cure for COPD, and the current medical interventions mostly include alleviating disease complications with bronchodilators such as inhaled long-acting \u03b32-agonists (LABAs) and long-acting muscarinic antagonists (LAMAs) [7].

In COPD patients, there is impaired crosstalk between lung circulatory and resident immune cells, resulting in the increased recruitment of neutrophils, macrophages, and T- and B-lymphocytes inside the airway lumen of COPD patients. Inhaled irritants provoke surface macrophages and airway epithelial cells to release chemokines, which recruit inflammatory cells to the lungs. However, this inflammation persists even after quitting smoking, possibly due to the selfsustaining feedback loop that hinders the alleviation of the inflammatory responses [8]. This unbalanced and dysregulated regulation leads to chronic inflammation, fibrosis, cell death, and extracellular matrix (ECM) destruction (i.e., airway remodeling) in the lungs of COPD patients.

Signal transduction regulatory mechanisms, including posttranslational phosphorylation of proteins, play a crucial role in COPD pathogenesis. Mitogen-activated protein kinases (MAPKs) are a diverse group of these kinases involved in many signal transduction pathways [9]. There are four well-characterized subfamilies of this group in mammalian cells, including ERK1/2 (Extracellular signal-regulated kinase 1 and 2), ERK5, JNKs (c-Jun N-terminal kinases), and p38s (Protein 38) [10]. As shown in earlier studies, p38 MAPKs are triggered via inflammatory mediators and oxidative stress, which activate many diverse downstream signaling pathways in different cells. External factors such as viral and bacterial infections and internal factors are involved in regulating the p38 MAPK signaling pathway [11].

However, given the role of chronic inflammation in COPD pathogenesis, the development of anti-inflammatory treatments for COPD has gained more attention in recent years. In this review, we conducted a comprehensive search to evaluate the significance of the p38 signaling pathway in the pathogenesis of COPD and the efficacy of available p38 inhibitors for COPD treatment.

Our approach and method

In July 2023, we conducted a comprehensive electronic search of two prominent databases, PubMed and Google Scholar, to identify studies published in English that explored the relationship between COPD and p38 with no temporal restrictions. We searched all fields of the articles using predefined search terms "COPD" AND "p38 using EndNote online search and utilized truncation to capture different variations of the terms such as "chronic obstructive pulmonary disease", "emphysema", and "AECOPD". To ensure the accuracy of our search strategy, we cross-checked the truncation terms with available MeSH terms.

To ensure the quality of our review, we conducted a meticulous screening of the titles and abstracts of the studies retrieved from our search. We selected only those studies that investigated the role of p38 MAPK signaling in COPD and its association with various cells, treatments, and clinical trials. We excluded studies that did not meet the research question's criteria, such as those that examined the role of p38 MAPK signaling in other respiratory diseases and review articles, to ensure the relevance of the included studies in our review.

To identify more relevant studies, we expanded our search to include additional search terms, such as "epithelial", "macrophage", "fibroblast", "neutrophil", "smooth muscle", "lymphocyte", and "endothelial", which we categorized separately. We also categorized treatments and interventions in in vivo (human and animal studies), in vitro, and ex vivo studies and subcategorized herbal interventions. After obtaining the full text of the selected studies, we conducted a comprehensive review to extract relevant data and assess evidence quality, including study design, sample size, patient characteristics, interventions, outcomes, statistical methods and findings, biological samples, organism, and key findings. We synthesized the selected studies' findings in a narrative review and grouped them by study design, patient characteristics, and measured outcomes. The systematic search strategy is illustrated following the PRISMA standard in Fig. 1 [12].

p38 signaling pathway

The p38 signaling pathway is a well-characterized intracellular cascade that plays a crucial role in regulating cellular responses to diverse extracellular stimuli, such as inflammatory cytokines, oxidative stress, osmotic shock, ultraviolet radiation, and growth factors. The



Fig. 1 Flow Diagram Illustrating the Selection Process of Studies for Inclusion in the PRISMA Systematic Review

primary mediator of the p38 pathway is a member of the mitogen-activated protein kinase (MAPK) family, which is a serine/threonine kinase family that phosphorylates a vast array of substrates in the cytoplasm and nucleus. The p38 signaling pathway is activated through a series of phosphorylation events that culminate in the activation of downstream effectors, including transcription factors, kinases, and phosphatases. This pathway is involved in numerous cellular processes, including inflammation, stress responses, cell cycle regulation, and differentiation, which underscores its importance in maintaining cellular homeostasis. Moreover, dysregulation of the p38 signaling pathway has been implicated in the pathogenesis of various diseases, such as cancer, autoimmune disorders, and neurodegenerative diseases, highlighting the need to gain a comprehensive understanding of the molecular mechanisms underlying p38 signaling.

Positive regulation of p38 signaling

p38 MAP kinases are activated by a three-tiered mechanism, where they phosphorylate p38 on the Thr180 and Tyr182 residues. The activation of p38 is initiated by diverse types of cellular receptors, such as cytokine receptors, growth factor receptors, G protein-coupled receptors, and pattern recognition receptors [13], which involve upstream kinases such as MKK3, MKK4, MKK6, and MKK7 [13, 14]. For instance, TNF α and IL-1 activate p38 via the NF- κ B pathway, insulin activates p38 using the PI3K/Akt pathway, and LPS activates p38 through the TLR4/MyD88 pathway [13]. Therefore, the cellular receptors involved in the activation of p38 may vary depending on the cell type and stimulus.

There are molecules, such as TNF- α , IL-1 β , IL-6, and MCP-1, mediate immune and inflammatory responses. Cytokines can activate p38 through different receptors and signaling pathways, such as TNFR1-TRAF2-TAB1-TAK1-MKK3/6, IL-1R-TI-RAP-MAL-TRAF6-TAK1-MKK3/6, IL-6R-JAK2-TYK2-MKK3/6, and CCR2-MCPIP-MKK3/6 [15, 16]. Cytokines are increased in the airways and circulation of COPD patients and contribute to chronic inflammation and tissue damage. The proinflammatory cytokines interleukin-1 beta (IL-1 β) and tumor necrosis factor-alpha $(TNF-\alpha)$ have been shown to activate p38 signaling and promote inflammation in COPD patients [17]. Additionally, oxidative stress, which is increased in COPD due to the reduced antioxidant capacity of the lungs, can activate p38 signaling and promote inflammation [18]. Additionally, UV radiation activates the p38 pathway by causing DNA damage via the upstream kinase MKK3 [19]. Likewise, heat shock and osmotic stress induce the activation of the p38 pathway through the upstream kinases MKK6 and MKK3, respectively [20].

Oxidative stress occurs when the production of reactive oxygen species (ROS) exceeds the antioxidant capacity of the cells. ROS are involved in the activation of p38 and other stress kinases, as well as redox-sensitive transcription factors, such as NF- κ B and AP-1, which regulate inflammation and tissue remodeling in COPD. Oxidative stress can activate p38 through various upstream kinases, such as ASK1, TAK1, MLK3, and MEKK1 [16]. Oxidative stress is induced by cigarette smoke and other environmental pollutants that are risk factors for COPD [21].

Moreover, there are components of the outer membrane of gram-negative bacteria that can trigger innate immune responses. LPS can activate p38 through Toll-like receptor 4 (TLR4) and its adaptor proteins, such as MyD88, TRIF, TRAM, and TRAF6, which can recruit TAK1-MKK3/6 to the receptor complex [16]. LPS can also induce oxidative stress and cytokine production that can further activate p38. LPS is present in the airways of COPD patients due to bacterial colonization or infection and can exacerbate inflammation and airflow limitation [21].

Negative regulation of p38 signaling

The upstream regulation of p38 and other MAPKs is a crucial aspect of MAPK signaling. Phosphatases, such as protein phosphatase 2A/C (PP2A/C), wild-type p53-induced phosphatase 1 (Wip1), MAP kinase phosphatases (MKPs), and dual-specificity phosphatases (DUSPs), naturally regulate the activity of p38 and other MAPKs by dephosphorylating them and leading to their inactivation [22]. MKPs function as negative feedback regulators of MAPK signaling and can modulate inflammation, apoptosis, and oxidative stress in lung cells in response to various stimuli, such as cytokines, growth factors, and oxidative stress [23].

Conversely, protein kinases can phosphorylate and activate or inhibit p38 and other MAPKs. MAP kinaseinteracting serine/threonine protein kinases (MNKs) are examples of protein kinases that inhibit p38 activation by phosphorylating and inhibiting MKK3, which is an upstream activator of p38. MNKs can also regulate the translation of several proteins involved in inflammation and fibrosis, such as TNF- α , IL-6, and collagen [15]. Additionally, MAP3Ks, which respond to different types of cellular stress, such as oxidative stress, DNA damage, cytokines, and TLR agonists, regulate p38 signaling. Apoptosis signal-regulating kinase 1 (ASK1, also known as MAP3K5), TGF-β-activated kinase 1 (TAK1, also known as MAP3K7), mixed lineage kinase 3 (MLK3, also known as MAP3K11), and mitogen-activated protein kinase kinase kinase 1 (MAP3K1, also known as MEKK1)

are some examples of MAP3Ks that regulate p38 signaling [24].

Physiologically, p38 can be inhibited either directly or indirectly by anti-inflammatory agents, antioxidants, and anticancer drugs. Antioxidants act by scavenging reactive oxygen species (ROS) or by preventing their formation. ROS are involved in the activation of p38 and other stress kinases and redox-sensitive transcription factors, such as NF-KB and AP-1, which regulate inflammation and tissue remodeling in COPD. Antioxidants can attenuate p38-mediated inflammatory responses in lung cells [13]. The phosphodiesterase-4 (PDE4) inhibitor roflumilast inhibits p38 activation and reduces inflammation in COPD patients [15]. It is crucial to achieve a comprehensive understanding of the negative regulators of p38 signaling in COPD to develop innovative therapeutic strategies for COPD and identify potential therapeutic targets.

The versatility of the p38 MAPK family

The p38 MAPK family consists of four isoforms: p38 α , p38 β , p38 γ , and p38 δ . These isoforms share a high degree of amino acid sequence homology and are activated by a variety of extracellular stimuli, including inflammatory cytokines, stress, and growth factors. p38 α is the most widely studied isoform of p38 and is ubiquitously expressed in mammalian tissues. It plays a critical role in regulating cellular responses to stress and inflammation, including cell cycle progression, apoptosis, and immune responses [20].

p38β is closely related to p38α, sharing 75% sequence identity. It is expressed at lower levels than $p38\alpha$ and is primarily found in the heart and skeletal muscle, where it has been shown to play a role in regulating cardiac hypertrophy and myogenesis [25]. p38y and p388 are more recently found isoforms of p38 that have distinct tissue distributions and functions. p38y is primarily expressed in the brain and has been implicated in regulating synaptic plasticity and memory formation. p388, on the other hand, is highly expressed in adipose tissue and has been shown to play a role in regulating adipocyte differentiation and insulin signaling [20]. Overall, the different isoforms of p38 have distinct tissue distributions and functions but share a similar protein structure and function as stress-activated protein kinases that play critical roles in regulating cellular responses to stress and inflammation.

The p38MAPK family includes highly conserved serine-threonine protein kinases with four isoforms, namely, α , β , γ , and δ . Although these proteins are activated downstream of various signals, they differ in terms of tissue distribution and activators/substrates [26]. A wide variety of stimuli, such as oxidative stress, inflammatory cytokines, and growth factors, stimulate several MAPK kinases (MEKK/MAP3K), which activate downstream kinases [26]. Alternatively, p38 α and p38 β can be activated in T cells in response to TCR (T-cell receptor) signaling, which is increased by T-cell costimulation, downstream of ζ -chain-associated protein kinase of 70 kilo Daltons (ZAP70) and spleen tyrosine kinase (SYK) protein (Fig. 2) [27].

Based on structural and sequence features as well as substrate specificities, the p38 MAPK family can be further divided into two subsets, p38 α /p38 β and p38 γ /p38 δ [10], with different effects on inflammation. For example, p38 γ can inhibit c-Jun phosphorylation, but p38 α promotes this process [28]. Additionally, p38 γ may suppress the transcription factor activating protein-1 (AP-1), suggesting an anti-inflammatory effect of this isoform [29]. Furthermore, the expression of the β isoform appears restricted to T cells, while p38 α is the most ubiquitously expressed isoform in different tissues [13, 30]. The expression levels of different p38 isoforms in COPD lung tissue have not been quantitatively studied. Therefore, p38 α is considered the main studied isoform associated with COPD pathogenesis [15, 31].

Given the importance of the p38 signaling pathway and its observed overexpression in COPD, this section will focus on the role of p38 signaling in modulating different inflammatory cells that contribute to the pathogenesis of COPD.

Dysregulation of p38 signaling

The dysregulation of the p38 signaling pathway has been linked to several diseases, including cancer, autoimmune disorders, and neurodegenerative diseases [32], highlighting the importance of comprehending the molecular mechanisms underlying p38 signaling.

In COPD, p38 signaling is upregulated in different cell types, such as bronchial epithelial cells, macrophages, and lymphocytes. This may contribute to chronic inflammation, tissue damage, and impaired repair in the lungs of COPD patients. Moreover, p38 signaling may also affect skeletal muscle structure and function in COPD, leading to sarcopenia and reduced exercise capacity. Oxidative stress has been shown to activate p38 signaling and cause muscle atrophy, fibrosis, and mitochondrial dysfunction in a mouse model of COPD [33].

The role of p38 in COPD: from cell signaling to lung damage

The role of p38 in different inflammatory cells contributing to COPD pathogenesis is crucial and will be discussed in more detail in the following section. We aim to provide a comprehensive understanding of the specific role of p38 MAPK signaling in each cell type, including



Fig. 2 General p38 Signaling Pathways. The figure illustrates the general pathways of p38 signaling. Environmental signals are transmitted through cell receptors (colored orange) to multiple MAPK kinases (MAP3K, colored pink). These MAPKK kinases phosphorylate downstream MAPK kinases (MAPKK, colored green), which subsequently activate the p38 isoforms. In noncanonical pathways, MAPKs can be activated through phosphorylation by ZAP70/SYK or through interaction with TAB1. Once activated, p38 can either remain in the cytoplasm or translocate into the nucleus, leading to the activation of various transcription factors (colored red) and kinase proteins (colored blue)

neutrophils, macrophages, T cells, and epithelial cells, which contribute to the development and progression of COPD.

Epithelial cells

Cigarette smoke and other irritants stimulate lung epithelial cells to produce inflammatory mediators, such as TNF- α (tumor necrosis factor-alpha), IL-1 β (interleukin 1 beta), IL-6, GM-CSF (granulocyte-macrophage colonystimulating factor), and CXCL8 (IL-8) [34] (Table 1 and Fig. 3), resulting in TGF- β (transforming growth factor β) production by the small airway epithelium, which triggers local fibrosis and remodeling [35]. Additionally, it stimulates epithelial cells to release chemotactic factors that attract neutrophils, T-helper (Th) 1 (Th1/ CD4+) cells, type 1 cytotoxic T cells (TC1/CD8+), and fibroblasts. Consequently, these inflammatory cells release proteases, growth factors, and proinflammatory cytokines, which promote chronic lung inflammation and structural changes [36].

In COPD patients, phospho-p38 is increased in small airway epithelia (compared to nonsmokers) [23] and in alveolar septa (compared to smokers and nonsmokers) [15]. Activation of p38 in epithelial cells of COPD patients induces local/systemic (pro) inflammatory signaling pathways and increases mucin and vascular Table 1 Genes and proteins that are affected downstream of p38 signaling within ex vivo models of COPD

Gene	Source	Treatment	Case/Control	Up/Down ^a	Ref
INFγ	CD8 cells (human)	S100+IL12+IL8	S100+IL12+IL8/IL12+IL8	Down-P	[23]
	MDM cells (human COPD)	Dexa ^b + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
IL-6	Epithelial cells (human COPD)	S100 + polyinosinic: polycyti- dylic acid	S100 + I:P/I:P	Down-P	[23]
	Alveolar macrophages (AMs)	LPS + Dexa + BIRB-796	LPS + Dexa + BIRB-796/LPS	Down-P	[38]
	Macrophages (CS exposed rat)	MSC+CS	MSC+CS/CS	Down-G	[39]
	Guinea Pig AMs	SB 239063 + LPS	SB 239063 + LPS/LPS	Down-P	[40]
CXCL-8	AMs (human COPD)	BIRB-796 + LPS	BIRB-796 + LPS/LPS	Down-P	[41]
	Epithelial cells (human COPD)	S100+I:P	S100+l:P/l:P	Down-P	[23]
	PBMCs (COPD human)	LPS + Dexa + GW856553		Down-P	[42]
CCL-5	Epithelial cells (human COPD)	S100 + I:P	S100+l:P/l:P	Down-P	[23]
	PBMCs (human COPD IV)	SB203580 + LPS	SB203580+LPS/LPS	Down-P	[43]
TNFa	Blood neutrophils (human COPD)	S100+LPS	S100+LPS/basal	Down-P	[23]
	AM cells (human COPD)	BIRB-796 + LPS	BIRB-796 + LPS/LPS	Down-P	[41]
	Alveolar macrophages	LPS + Dexa + BIRB-796	LPS + Dexa + BIRB-796/LPS	Down-P	[38]
	Whole blood (COPD GOLD II)	SB-681323 + LPS	SB-681323 + LPS/Placebo + LPS	Down-P	[44]
	PBMCs (human COPD IV)	SB203580 + LPS	SB203580+LPS/LPS	Up-P	[43]
IL-10	Macrophages (CS exposed rat)	MSC+CS	MSC+CS/CS	Up-G	[39]
	PBMCs (human COPD IV)	SB203580 + LPS	SB203580+LPS/LPS	Up-P	[43]
	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
	Alveolar macrophages	LPS + Dexa + BIRB-796	LPS + Dexa + BIRB-796/LPS	Down-P	[38]
iNOS	Macrophages (CS exposed rat)	MSC+CS	MSC+CS/CS	Down-G	[39]
COX-2	Macrophages (CS exposed rat)	MSC+CS	MSC+CS/CS	Down-G	[39]
PGE-2	Macrophages (CS exposed rat)	MSC+CS	MSC+CS/CS	Down-G	[39]
CCL7	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
TNFSF-9	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
CSF-2	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
CCL-2	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
TNFSF-15	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
TNFSF-4	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
IL-12B	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
TNF	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
CXCL-10	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
IL-27	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
CLCF-1	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
OSM	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
CCL-8	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
IL-6	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
IL-1A	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
CXCL-9	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
CXCL-2	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
CXCL-3	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
CXCL-1	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
IL-8	MDM cells (human COPD)	Dexa + SB706504 + LPS	LPS/Dexa + SB706504 + LPS	Down-G	[37]
	Alveolar macrophages	LPS + Dexa + BIRB-796	LPS + Dexa + BIRB-796/LPS	Down-P	[38]
GM-CSF	Alveolar macrophages	LPS + Dexa + BIRB-796	LPS + Dexa + BIRB-796/LPS	Down-P	[38]
IL-1ra	Alveolar macrophages	LPS + Dexa + BIRB-796	LPS + Dexa + BIRB-796/LPS	Down-P	[38]
MDC	- Alveolar macrophages	LPS + Dexa + BIRB-796	LPS + Dexa + BIRB-796/LPS	Down-P	[38]

^a Down Downregulation, Up Upregulation, P Protein, G Gene

^b Dexa Dexamethasone



Fig. 3 Association of p38 Signaling with COPD Pathogenesis. At the top of the figure, p38 activators and inhibitors are depicted. Downstream of p38 activation, cytokines are highlighted in green, while transcription factors are colored red. Other downstream biomolecules are represented in blue. The figure also highlights cell-specific downstream targets of p38 activation in various immune cell subtypes. However, for other molecules, the target cell types remain unclear

endothelial growth factor (VEGF). p38-mediated mucin hypersecretion is accompanied by a decrease in the function of the tyrosine phosphatase SHP-1 in COPD patients [45]. p38 activation in bronchial epithelial cells can contribute to CXCR3-induced chemotaxis [46]. The expression of IL-8 and p38 was dramatically increased following exposure to oxidative and inflammatory agents such as H_2O_2 , cytomix, and LPS (lipopolysaccharides) in the human bronchial epithelial cell line 16HBEBEAS-2B [47-50]. Meanwhile, p38 inhibition has been shown to reduce the production of pro-inflammatory cytokines such as IL-6, CXCL8, and CCL5 (Chemokine (C-C motif) ligand 5) from isolated lung epithelial cells of COPD patients [23] (Table 1 and Fig. 3). Reactive oxygen species can disrupt the integrity of the endothelial cell barrier through the p38-HSP27 (Heat Shock Protein 27)-dependent pathway, demonstrating the critical role of p38 signaling in the remodeling process of the lung epithelium [51]. C-reactive protein (CRP) and fibrinogen are systemic inflammation biomarkers that are associated with poor prognosis, increased mortality rate, and exacerbation periods in COPD patients [52]. CRP has been shown to be synthesized by respiratory epithelial cells [53]. The p38 inhibitors PH-797804 and losmapimod are suggested to regulate CRP and fibrinogen by inhibiting inflammatory pathways such as IL-1 β downstream signaling [54–57].

Additionally, p38 is involved in the expression of TNF- α -induced VEGF in human airway smooth muscle cells (HASMC) [57], and balanced VEGF expression is critical to maintaining alveolar cell integrity [58]. Suppression of VEGF receptors leads to apoptosis and remodeling in alveolar cells and promotes emphysema-like pathology. Excessive VEGF expression may contribute to a COPDlike phenotype without emphysema [35, 59] (Table 1 and Fig. 3).

p38 signaling can also give rise to the development of secondary complications associated with COPD exacerbations, such as rhinovirus-induced VEFG production and influenza virus-induced interferon-stimulated gene expression of p38 in airway epithelial cells [60, 61]. The limited efficacy of corticosteroids in alleviating the chronic inflammation of human bronchial epithelial cells

in asthma and COPD after an oxidative stress challenge has raised more attention to developing effective p38 inhibitors [62].

Another crucial finding is that in model mice, the premature senescence of airway epithelial cells (Clara cells) was induced by repeated exposure to a toxicant. This led to impaired epithelial regeneration and was accompanied by airway inflammation that was dependent on the activation of p38 MAPK. Moreover, in COPD patients, Clara cell senescence was observed to be accelerated and exacerbated by p38 MAPK, indicating its potential role in the development of COPD. This finding highlights the significance of Clara cell senescence and its interaction with p38 MAPK in the pathogenesis of COPD [63]. Several studies have reported similar observations regarding the role of premature senescence in COPD [64-66]. The collective body of research in this area underscores the importance of understanding the mechanisms underlying airway epithelial cell senescence and its implications in COPD.

Overall, these studies suggest that the p38 MAPK signaling pathway plays a crucial role in the progression of COPD in epithelial cells. Activation of the p38 MAPK pathway is associated with oxidative stress, cellular senescence, and inflammation, leading to the secretion of inflammatory proteins and the accumulation of senescent cells in the lungs. The dysregulation of TLRs [66], NOX4, Hsp70 [67, 68], miR-494-3p [64], CD147 [69], and ANGPTL4 [70] in COPD also involves the activation of the p38 MAPK signaling pathway.

A thorough investigation is necessary to fully understand the intricate molecular mechanisms that underlie the crucial role of the p38 MAPK signaling pathway in the pathogenesis of COPD and its related complications. It is imperative to find specific inhibitors and assess their safety and efficacy in clinical trials to develop effective treatments for COPD. Future research should be geared toward elucidating the underlying molecular mechanisms of the p38 MAPK signaling pathway in COPD pathogenesis and designing effective p38 inhibitors to alleviate chronic inflammation in bronchial and alveolar epithelial cells.

Macrophages

Macrophages are important in COPD because they are involved in inflammatory and phagocytic processes, cytokine production and cellular interactions. Macrophages in COPD have been shown to have different phenotypes, functions and localizations, depending on the disease stage and severity [71]. Macrophages can contribute to the pathogenesis of COPD by causing oxidative stress, protease/antiprotease imbalance, apoptosis, antigen presentation and tissue destruction. There are different kinds of macrophages in the respiratory system that have different physiological characteristics and unique functions. Alveolar macrophages (AMs) and IMs in the lung. AMs can be further divided into tissue-resident alveolar macrophages (TR-AMs) and monocyte-derived alveolar macrophages (Mo-AMs), depending on their origin and function. Furthermore, AMs can be subdivided into M1-like (proinflammatory) and M2-like (anti-inflammatory) phenotypes, depending on the stimuli they encounter [73]. M1-like macrophages contribute to the initiation and amplification of inflammation, while M2-like macrophages promote the resolution of inflammation and tissue repair.

Different stimuli, such as cigarette smoke, hypoxia, oxidative stress, and microbial infections, can modulate the phenotypes and functions of macrophages in COPD toward activation and polarization in inflammation and the immune response. p38 signaling is a pathway that regulates the expression of proinflammatory cytokines and mediators in macrophages, is activated by cigarette smoke and has increased activity in COPD alveolar macrophages. p38 signaling may contribute to inflammation, tissue damage and corticosteroid resistance in COPD.

Lung macrophage populations are elevated in both heavy smokers and COPD patients. Elevated levels of chemokines such as CCL2 and CXCL1, which are found to increase in the sputum and bronchoalveolar fluid of COPD patients, induce the recruitment of circulating monocytes [74] (Table 1 and Fig. 3). Furthermore, the percentage of alveolar macrophages expressing phosphop38 is consistently higher in COPD patients [15, 23, 38]. A cell-type-specific analysis of the presence of phosphop38 MAPK in the lung tissue of COPD patients, smokers, and nonsmokers revealed elevated levels in bronchial epithelial cells, macrophages, and CD20+ and CD8+ lymphocytes in COPD lungs but not in sputum and lung tissue neutrophils. Notably, p38 MAPK inhibition reduced proinflammatory mediator release in COPD lung CD8 cells and airway epithelia [23].

Activation of p38 can be initiated by the upregulation of cell receptors found upstream of the p38 signaling pathway. Fibrinogen-like protein 2 (FGL2) expression is upregulated in lung macrophages of COPD patients as well as in THP-1 cells, a human monocyte-macrophage line (Fig. 4). FGL2 is a transmembrane-anchored protein expressed on macrophages and endothelial cells that positively regulates proinflammatory cytokines such as IFN- γ (interferon-gamma) and TNF- α [75]. Conversely,



Fig. 4 Clinical manifestations of the $p38\alpha/\beta$ signaling pathway. The activation of specific signaling pathways triggers the activation of the $p38\alpha/\beta$ signaling pathway. This pathway influences various cellular messengers, leading to the development of different phenotypes in diverse cell types

FGL2 is also involved in LPS-induced macrophage activation through the p38 pathway. This interaction may stimulate the release of proinflammatory cytokines and enhance macrophage migration in the lung [76] (Fig. 3).

Despite numerous reports suggesting enhanced p38 phosphorylation in animal models after cigarette smoke exposure [77], p38 phosphorylation has been proposed to be more closely associated with COPD than with smoking [15] (Table 1 and Fig. 3). Research has proven that p38 inhibition can reduce the release of various proinflammatory cytokines, including TNF- α and granulo-cyte-macrophage colony-stimulating factor (GM-CSF), from COPD macrophages. Nonetheless, alveolar macrophages have been shown to be resistant to glucocorticoid anti-inflammatory therapies, and this phenomenon has been observed in both COPD patients and controls [23, 38, 78].

An in vitro study cultured the murine alveolar macrophage line (MH-S) with serum from COPD patients (GOLD stages of III/IV), which resulted in increased expression of CCL5 and decreased expression of IL-10 compared to healthy controls. However, secondary treatment of cells with the p38 inhibitor SB203580 had limited success in restoring impaired lymphocyte function and COPD-induced inflammation suppression, indicating the importance of p38-independent inflammatory mechanisms in the pathogenesis of COPD [43] (Table 2). Combining p38 inhibitors BIRB-796/SB706504 with corticosteroids in COPD patients can synergistically enhance the secretion of anti-LPS cytokines of alveolar macrophages, including IL-8, IL-6, TNF, IL-10, RANTES (Regulated on Activation, Normal T-cell Expressed and Secreted; CCL5), GM-CSF, IL-1β, IL-1ra, and MDC [37, 38, 78] (Table 1 and Fig. 3). This synergistic effect can be partially attributed to glucocorticoid-induced upregulation of MAPK phosphatase expression [79]. However, it should be noted that IL-1, IL-18, and CCL5 are resistant to coadministration of dexamethasone and SB706504 and thus do not show a substantial increase in expression [37]. Therefore, the therapeutic application of p38 inhibition is highly compromised due to the significant risk of inhibiting essential immune functions and its inefficacy on alveolar macrophages. Nonetheless, inhibiting the p38 pathway using the structurally distinct inhibitors VX745

Ref

[50]

[50] [43]

[43]

[43]

Down-P

Down-G&P [81]

Gene	Source	Treatment	Case/control	Up/down ^a
IL-8	16HBE Cell line	$H_2O_2/TNF-\alpha + IL-1\beta + INF\gamma/LPS$	H2O2/TNF-α + IL-1β + INFγ/LPS vs. baseline	Down-G
NFkB p65	16HBE Cell line	$H2O_2/TNF-\alpha + IL-1\beta + INF\gamma/LPS$	H2O2/TNF- α + IL-1 β + INF γ / baseline with LPS	Down-G
IL-10	MH-S cell line	GOLD III/IV serum + LPS + SB203580	GOLD III/IV serum + LPS + SB203580/ GOLD III/IV serum + LPS	Up-P
CCI-5	MH-S cell line	GOLD IV COPD serum + LPS + SB203580	GOLD III and IV COPD serum + LPS + SB203580/ GOLD III	Down-P

Table 2 Genes and proteins that are affected downstream of P38 signaling within in vitro models of COPD

^a Down Downregulation, Up Upregulation, P Protein, G Gene

MH-S cell line

A549 cells

TNFa

GRP-78

and SCIO469 does not alter innate immune responses such as phagocytosis or efferocytosis by macrophages [80]. A study showed that although SB706504 could significantly reduce TNF- α expression in LPS-stimulated alveolar macrophages in COPD patients and smokers, the expression was further maximized by coadministration of SB706504 and dexamethasone [37]. In general, p38 activation is insensitive to corticosteroids, and combining a p38 inhibitor with a corticosteroid can synergistically enhance the anti-inflammatory effects [38].

SB203580 + CS

Smokers with a high MMP-9/TIMP-1 ratio have been found to have a positive correlation with the annual decline in forced expiratory volume in 1 s (FEV1%), forced vital capacity (FVC%), and MMEF%pred. Moreover, inhibition of p38 MAPK and ERK suppressed the production of MMP-9 in alveolar macrophages (AMs) from smokers. Furthermore, Guan et al. [82] reported that MMP-12 contributes to the proliferation of mouse macrophages by regulating the ERK/P38 MAPK signaling pathway. These findings suggest that the MMP-9/TIMP-1 ratio could be a useful predictor of future risk of COPD among smokers with AHR. Additionally, previous studies have investigated the role of MMP-9/TIMP-1 imbalance in various lung diseases, including COPD and asthma [83].

Alveolar macrophages (AMs) and monocyte-derived macrophages (MDMs) from COPD patients showed lower levels of bacterial phagocytosis and efferocytosis than those from healthy controls. It was found that p38 MAPK inhibition impaired bacterial phagocytosis by M1 macrophages from COPD patients but enhanced efferocytosis by M2 macrophages from all groups [80], while Vij et al. [84] found that cigarette smoke activated p38 MAP kinase and inhibited autophagy in macrophages, leading to increased inflammation and impaired phagocytosis. However, none of the inhibitors, including p38 MAPK inhibitors, altered bacterial internalization or early intracellular bacterial killing in AM or MDM [80]. Additionally, in an in vitro model of human alveolar macrophage (AM) smoke exposure, cigarette smoke activated p38 MAP kinase in GM-CSF-derived macrophages (GM-MOs) and enhanced IL8 production while inhibiting phagocytosis, similar to the phenotype of smokers' Ams [85].

Neutrophils

and IV COPD serum + LPS

and IV COPD serum + LPS

SB203580+CS/CS

GOLD III and IV COPD serum + LPS + SB203580 GOLD III and IV COPD serum + LPS + SB203580/ GOLD III

Neutrophils are involved in the lung response to various infections and diseases, such as tuberculosis, acute lung injury, and chronic pulmonary diseases. Depending on the context, neutrophils can have beneficial or detrimental effects on lung tissue and function. The role of neutrophils in the lung is complex and dynamic and depends on the type, duration, and severity of the infection or injury. Neutrophils can either protect or damage lung tissue by regulating the balance between inflammation and resolution, defense and repair, or clearance and fibrosis. Based on these functions, neutrophils in the lung can be classified into different kinds according to their phenotype, activation state, origin, or location. Lung-resident neutrophils, blood-derived neutrophils, senescent (aged) neutrophils, and N1 and N2 neutrophils are subsets of neutrophils that can be found in the respiratory system [86, 87].

Lung-resident neutrophils are a subset of neutrophils that reside in the lung under steady-state conditions. They have a unique immunosuppressive phenotype that protects the lungs against damage from overexaggerated responses to injury or infection. They express high levels of CXCR4 and CD11b but low levels of CD62L. They are maintained by high levels of interleukin-6 (IL-6) and prostaglandin E2 (PGE2) in the lung [88, 89].

Blood-derived neutrophils are neutrophils that migrate from the blood into the lung upon inflammatory stimuli. They have a proinflammatory phenotype that helps fight infection and clear debris. They express high levels of CD62L and CD16 but low levels of CXCR4. They are recruited by chemokines such as CXCL1, CXCL2, CXCL5, CXCL8, CCL2, CCL3, CCL4, CCL5, CCL7, and CCL8 [90]. Senescent neutrophils are also a subset of neutrophils that have a prolonged lifespan due to inflammatory signals or oxidative stress. Senescence refers to a state of irreversible growth arrest and altered cellular function. They have an altered phenotype that makes them more prone to apoptosis or NET formation. They express high levels of CD44 and CD64 but low levels of CD16 and CD62L. They are cleared by macrophages or efferocytosis [91, 92].

N1 and N2 neutrophils are two subsets of neutrophils that have different polarization states depending on the cytokine environment. N1 neutrophils are pro-inflammatory and produce high levels of TNF- α , interferon gamma (IFN-γ), IL-12, IL-23, IL-1β, IL-6, and IL-8. N2 neutrophils are anti-inflammatory and produce high levels of IL-10, TGF-β, arginase 1 (ARG1), VEGF, and IL-4. N1 neutrophils have antitumorigenic effects by killing tumor cells, stimulating cytotoxic T cells, and inhibiting angiogenesis. N2 neutrophils have protumorigenic effects by promoting tumor growth, metastasis, and immunosuppression. N1 neutrophils are involved in the pathogenesis of autoimmune diseases such as rheumatoid arthritis, systemic lupus erythematosus, and multiple sclerosis by producing inflammatory mediators, forming NETs, and activating autoreactive T cells. N2 neutrophils are involved in the resolution of inflammation and tissue repair by producing anti-inflammatory mediators, clearing apoptotic cells, and inducing regulatory T cells. N1 neutrophils are essential for the clearance of intracellular pathogens such as Mycobacterium tuberculosis, Leishmania donovani, and Salmonella typhimurium by enhancing phagocytosis, ROS production, and cytokine secretion. N2 neutrophils are detrimental to the host defense against these pathogens by impairing phagocytosis, ROS production, and cytokine secretion [93–95].

In COPD, neutrophils represent the predominant subtype of white blood cells involved in the inflammatory response. However, it is important to recognize that neutrophils can exhibit functional heterogeneity, indicating the presence of different subtypes or phenotypes based on their activation status and functions within the disease context [96]. One subtype identified in COPD is the "primed" or "activated" neutrophils, characterized by an enhanced ability to release inflammatory mediators and reactive oxygen species. These neutrophils display an altered phenotype with increased expression of specific surface markers, such as CD11b and CD66b [97]. Another subtype is the "senescent" neutrophils found in the airways of COPD patients, which are characterized by impaired apoptosis and reduced clearance by macrophages, leading to their accumulation in the lungs [98].

Different subtypes of neutrophils contribute to the inflammatory processes and tissue damage observed in

COPD. However, it is important to note that neutrophil counts alone cannot solely determine the severity or progression of COPD. A comprehensive evaluation of the disease requires clinical assessment, considering symptoms, exacerbation history, and imaging findings. Additionally, it is important to acknowledge that neutrophil counts can vary and fluctuate among individuals with COPD, influenced by factors such as disease stage, exacerbation status, and individual patient characteristics [1].

Neutrophil counts in COPD display heterogeneity across different respiratory system locations. Inflammation, immune response, and bacterial infection contribute to increased neutrophil counts in the airways, bronchi, bronchial epithelium [99], mucous glands, BAL fluid [98, 100], and sputum samples of COPD patients [98, 101–103]. Neutrophil counts in peripheral blood can be either increased or normal, depending on the disease phase [102]. The systemic nature of neutrophilic inflammation in COPD suggests involvement beyond localized regions. In the lung parenchyma and alveoli, neutrophil infiltration occurs in regions of tissue damage and inflammation, with variations in counts linked to disease severity and exacerbations [98].

Multiple studies have provided evidence of increased p38 signaling in alveolar macrophages, epithelial cells, and lymphocytes within the lungs of individuals with COPD, while neutrophils do not exhibit the same upregulation [15, 23]. This suggests that p38 signaling may have distinct roles and effects across different cell types. However, other research has demonstrated the involvement of p38 signaling in neutrophil activation, migration, and survival in COPD. Notably, p38 signaling is responsible for mediating the release of neutrophil elastase, a protease implicated in pulmonary damage and inflammation [16]. Furthermore, p38 signaling has been identified as a regulator of CXCR2 expression, a chemokine receptor that guides neutrophil recruitment to the airways in COPD [104].

COPD lung neutrophils are a significant source of CXCL8 (IL8) secretion, which is the primary cytokine involved in the recruitment neutrophils during COPD pathogenesis [105]. After localization, neutrophils contribute to alveolar destruction and airway remodeling by releasing various metalloproteases, such as MMP-8 and MMP-9, as well as several other proteinases, including elastase, cathepsin-G, and proteinase-3. Although p38 isoforms have been reported in several immune cells [25], the dual phosphorylation and activation of p38 have not been observed in resident sputum neutrophils in COPD patients. It seems that the activation of this kinase is not necessary for the proinflammatory activity of lung neutrophils in COPD [23]. The selective inhibitor of p38 α and p38 β , SB100, was not effective in suppressing the proinflammatory cytokines released from lung neutrophils [23]. Additionally, lung neutrophils exhibit low expression of the glucocorticoid receptor [106] (Table 1 and Fig. 4). This altered behavior of lung neutrophils suggests that specific anti-neutrophil therapies beyond broad anti-inflammatory agents are necessary to manage COPD complications.

Therefore, the role of p38 signaling in neutrophils in COPD is of paramount importance, contingent upon the specific context and activating stimuli. The inhibition of p38 signaling holds promise in attenuating inflammation and averting tissue damage in COPD. However, it is imperative to acknowledge the potential adverse effects, including compromised host defense mechanisms and hindered bacterial clearance, associated with p38 signaling inhibition. Consequently, further investigation is warranted to gain a comprehensive understanding of the precise mechanisms and consequences of p38 signaling in neutrophils within the context of COPD.

Lymphocytes

Recent studies have explored the relationship between inflammatory cell infiltration and COPD, with particular attention given to CD8+ (TC1) cells [107]. These studies have found that the increased numbers of these cells in COPD patients are the only significant difference in inflammatory cell infiltration between asymptomatic smokers and those with COPD [108, 109]. Furthermore, this infiltration is correlated with alveolar destruction levels and the intensity of airflow obstruction [107]. In addition to CD8+ (TC1) cells, upregulation of phosphop38 has also been associated with the severity of lung function decline and the number of infiltrating CD8+ T-lymphocytes in alveolar walls [15].

The therapeutic significance of p38 inhibitors has been emphasized, given the observation that selective inhibition of p38 α and β can suppress IL-2 production in lymphocytes, making these isoforms highly relevant to CD8+ T-cell function [110]. In addition, SB100-induced p38 inhibition reduces proinflammatory cytokine production in lung and blood CD8+ cells in COPD patients, while p38 inhibition significantly reduces IFN-y release from COPD lung-isolated CD8+ cells [23]. Nevertheless, phospho-p38 expression in lymphocytes found in the peripheral airway submucosa was comparable in COPD patients, smokers, and nonsmokers (Table 1 and Fig. 3). In COPD patients, the inhibitory effect of glucocorticoids on IFN-y release from bronchi-alveolar lymphocytes is reduced compared to controls [111]. Moreover, IFN-y induces alveolar macrophages to secrete cytokines in a glucocorticoid-insensitive manner through STAT1 (signal transducer and activator of transcription 1) activation in COPD patients [112]. Thus, the inhibition of p38 signaling in lung CD8+ cells could overcome glucocorticoid insensitivity mechanisms, such as IFN- γ production from CD8+ cells and subsequent STAT1-mediated cytokine production from macrophages. Interestingly, the combination of a low dose of a selective p38 inhibitor, losmapimod (GW856553), with dexamethasone has been shown to have a synergistic effect on the suppression of CXCL8 release from peripheral blood mononuclear cells (PBMCs) of COPD patients [42] (Table 1 and Fig. 3).

Smooth muscle cells

Smooth muscle cells (SMCs) are a type of involuntary muscle cells that are found in various organs and tissues, such as the blood vessels, the digestive tract, the urinary bladder, and the eye. Smooth muscle cells can contract and relax to regulate the flow of fluids and substances in the body.

In the lung, SMCs are mainly found in the walls of the airways, such as the trachea, bronchi, and bronchioles. They help control the diameter of the airways and the resistance to airflow. Smooth muscle cells in the lung can also respond to various stimuli, such as hormones, neurotransmitters, inflammatory mediators, and environmental factors. For example, smooth muscle cells can contract in response to histamine, a chemical released during allergic reactions, causing bronchoconstriction and asthma symptoms.

Smooth muscle cells in the lung are also involved in some diseases and disorders, such as COPD, pulmonary hypertension, and lung cancer. In COPD, smooth muscle cells in the airways become hypertrophied (enlarged) and hyperplastic (increased in number), contributing to airway remodeling and obstruction. In pulmonary hypertension, smooth muscle cells in the pulmonary arteries become proliferative and migratory, leading to vascular remodeling and increased blood pressure in the lungs [113].

The role of p38 signaling in SMCs in COPD is not fully understood, but some studies have suggested that it may be involved in the pathogenesis and progression of the disease. In SMCs, p38 MAPKs may have different roles depending on the location and stage of the disease. In airway SMCs, p38 MAPKs may contribute to airway inflammation and remodeling by inducing the production of proinflammatory cytokines, such as IL-8. In addition, p38 MAPKs may regulate the contractility and proliferation of airway SMCs, which may affect airway hyperresponsiveness and obstruction. In vascular SMCs, p38 MAPKs may be involved in pulmonary hypertension, a common complication of COPD. p38 MAPKs may promote the proliferation and migration of vascular SMCs, leading to vascular remodeling and increased pulmonary vascular resistance [114]. Therefore, p38 signaling in SMCs in COPD may have both beneficial and detrimental effects

on the disease outcome. Inhibiting p38 MAPKs may have anti-inflammatory and bronchodilatory effects, but it may also impair host defense and bacterial clearance.

SMCs are involved in the inflammation associated with COPD [115]. SMCs release macrophage activating factors and chemokines, such as IL-8 and GM-CSF [116, 117]. Furthermore, the release of inflammatory mediators from HASMCs can cause insensitivity to corticosteroids. In contrast to corticosteroids, resveratrol can effectively reduce IL-8 and GM-CSF release or moderately reduce VEGF release from HASMCs in COPD patients. Therefore, resveratrol may be a preferred therapy for COPD over corticosteroids [118]. A study demonstrated that resveratrol's p38MAPK blockage can be implemented in corticosteroid-insensitive SMCs in COPD [119].

Additionally, in COPD, SMCs in the airways become hypertrophied (enlarged) and hyperplastic (increased in number), contributing to airway remodeling and obstruction [120]. In addition, SMCs in COPD show altered metabolism and mitochondrial function, which may affect their growth, survival, and inflammation [121].

Endothelial cells

Endothelial cells play a crucial role in the lung by lining the inner surface of blood vessels and lymphatic vessels. They are involved in various important functions, including gas exchange, vascular permeability, angiogenesis, inflammation, and fibrosis. These cells form a thin barrier between the blood and the alveoli, enabling the exchange of oxygen and carbon dioxide. Additionally, endothelial cells produce nitric oxide, a vasodilator that regulates blood flow and pressure in the lung. One of the key functions of endothelial cells is controlling the movement of fluid and molecules across the blood vessel wall. They can adjust their permeability by altering their shape, junctions, and cytoskeleton. Moreover, endothelial cells secrete factors that influence the permeability of other cells, such as epithelial cells and fibroblasts [122]. Endothelial cells also play a vital role in angiogenesis, which is the formation of new blood vessels from preexisting ones. This process is essential for normal lung development, growth, and repair. In response to various environmental signals, such as hypoxia, growth factors, and cytokines, endothelial cells can sense and respond by proliferating, migrating, and differentiating to form new vessels. Furthermore, endothelial cells actively participate in the inflammatory response within the lung. They accomplish this by expressing adhesion molecules, chemokines, and cytokines that recruit and activate leukocytes. Additionally, endothelial cells interact with immune cells such as macrophages and dendritic cells to modulate their function [123]. In the context of pulmonary fibrosis, endothelial cells contribute to the pathological process through a phenomenon called endothelial-to-mesenchymal transition (EndMT). This process involves endothelial cells losing their endothelial identity and acquiring mesenchymal characteristics. EndMT can lead to the loss of capillaries and the accumulation of myofibroblasts and extracellular matrix in the lung, ultimately contributing to fibrosis.

In individuals with COPD, there is evidence of shortened telomerase activity and reduced telomere length in pulmonary endothelial cells. These findings suggest the presence and increased accumulation of senescent endothelial cells in COPD patients [124]. The senescence observed in the pulmonary endothelium of COPD patients can be attributed to chronic exposure to oxidative stress and the persistence of inflammation. These pathological conditions worsen the release of inflammatory cytokines from endothelial cells, contributing to the senescence of endothelial function in the lungs of individuals with COPD [124, 125]. Furthermore, COPD patients demonstrate reduced levels of endothelial PAS domain-containing protein 1 (EPAS1) messenger RNA (mRNA) and an increase in EPAS1 promoter methylation, which correlates with elevated TLR4 expression. Through in vitro experimentation, it was observed that TLR4 inhibits EPAS1 mRNA expression while concurrently promoting promoter methylation in endothelial cells. These findings substantiate the notion that TLR4 overexpression within endothelial cells contributes to the progression of COPD by downregulating EPAS1 expression [126].

Exposure to cigarette smoke, a prominent risk factor for COPD, disrupts multiple pathways in endothelial cells, leading to endothelial cell dysfunction and inflammation. Notably, the p38 MAPK pathway plays a significant role in mediating endothelial cell dysfunction and inflammation in the context of COPD. The involvement of p38 MAPK in driving endothelial cell dysfunction and inflammation underscores its critical role in the pathogenesis of COPD. Further investigations are warranted to unravel the underlying mechanisms by which p38 MAPK contributes to these processes in endothelial cells. This knowledge will pave the way for the development of targeted therapies that can modulate this pathway and potentially alleviate endothelial dysfunction and inflammation in COPD.

Fibroblasts

Fibroblasts represent a diverse population of cells that play a crucial role in maintaining the structural integrity of the lung under both normal and pathological

conditions. Their primary function involves the synthesis and remodeling of the extracellular matrix (ECM), which provides support to the lung tissue. Additionally, they actively participate in processes such as wound healing, inflammation, and fibrosis. Various types of cells closely associated with fibroblasts include resident fibroblasts, myofibroblasts, pericytes, mesenchymal stem cells (MSCs), and fibrocytes. Among these, resident fibroblasts constitute the predominant cell type within the lung interstitium and can be categorized further based on their location, morphology, function, and specific cell surface markers, such as Thy-1 [127, 128]. Myofibroblasts, on the other hand, are activated fibroblasts that exhibit heightened contractility and secrete substantial amounts of ECM proteins such as collagen and fibronectin. While they play a critical role in wound closure and tissue repair, dysregulation of myofibroblasts can lead to fibrosis [129]. Pericytes, which are contractile cells surrounding capillaries and microvessels in the lung, regulate vascular tone, permeability, and angiogenesis. In response to certain stimuli, pericytes can differentiate into myofibroblasts or other types of mesenchymal cells [128]. Mesenchymal stem cells represent a multipotent cell population capable of self-renewal and differentiation into various lineages, including fibroblasts, myofibroblasts, adipocytes, chondrocytes, and osteoblasts. These cells possess immunomodulatory and anti-inflammatory properties, and they can migrate to sites of injury or inflammation. [130]. Last, fibrocytes are circulating cells derived from the bone marrow that express both hematopoietic and mesenchymal markers. In response to inflammatory signals, fibrocytes can migrate to the lung and differentiate into fibroblasts or myofibroblasts. Their role in lung fibrosis involves enhancing ECM deposition and promoting fibroblast proliferation [130].

In COPD, fibroblasts play a significant role in various pathological mechanisms that contribute to disease progression and severity. These mechanisms include EMT, senescence, protease-antiprotease imbalance, and cellcell interactions. Additionally, the p38 signaling pathway plays a crucial role in regulating the function and phenotype of fibroblasts in COPD.

EMT is particularly implicated in smoking-related COPD, as it can lead to the loss of epithelial barrier function, increased inflammation, and enhanced deposition of the ECM [131, 132]. The p38 signaling pathway is one of the key pathways involved in mediating EMT in response to cigarette smoke and other stimuli [133].

Senescent fibroblasts, characterized by reduced proliferation, increased proinflammatory and profibrotic mediators, and impaired ECM remodeling, also contribute to COPD pathology [134, 135]. The p38

signaling pathway is involved in inducing and maintaining senescence in fibroblasts through various mechanisms, including the DNA damage response, oxidative stress, and cytokine signaling [136, 137].

Protease-antiprotease imbalance refers to an alteration in the balance between proteases (enzymes that breakdown proteins) and antiproteases (proteins that inhibit proteases), leading to excessive degradation of the ECM and tissue damage. Fibroblasts can produce both proteases (such as matrix metalloproteinases, or MMPs) and antiproteases (such as plasminogen activator inhibitors, or PAIs) and can modulate their activity in response to various stimuli. In COPD, fibroblasts may exhibit an altered protease-antiprotease balance that favors ECM destruction. The p38 signaling pathway is involved in regulating the expression and activity of proteases and antiproteases in fibroblasts through transcriptional and posttranslational mechanisms [138, 139].

In severe COPD, the uncoupling of pericytes from the microvasculature, as well as their migration toward the larger pulmonary arteries, are significant pathogenic factors contributing to emphysema and pulmonary hypertension (PH) [140]. Furthermore, cigarette smoke extract (CSE) has been found to activate the proliferation of pulmonary fibroblasts through p38 MAPK signaling, resulting in airway remodeling in COPD [141].

Treatment of lung fibroblasts obtained from former and current smokers with CSE revealed that CSE exposure affects the cell cycle by reducing the S phase and increasing the G1 and G2/M phases, increasing the expression of p53 and p21, activating the p38 and ERK 1/2 pathways, and augmenting IL-8 release. Inhibitors of p38 and ERK 1/2 reversed the effects of CSE on the cell cycle and IL-8 release, suggesting that cigarette smoke exposure may hamper the reparative potential of lung fibroblasts by altering the expression of p53 and p21 and the progression of the cell cycle, with a potential role for p38 and ERK 1/2 signaling [142].

In summary, the findings from these studies suggest that p38 MAPK signaling plays a critical role in regulating lung fibroblast function and may be involved in the pathogenesis of lung diseases such as COPD. Inhibition of p38 signaling has the potential to be a therapeutic strategy in reducing fibroblast-mediated inflammation and airway remodeling in these diseases.

Targeting p38 as a therapeutic strategy for COPD treatment

Several p38 inhibitors have been developed and tested in cellular and animal models of COPD, as well as in clinical trials with COPD patients. These inhibitors have shown potent anti-inflammatory effects in vitro and in vivo, often greater than those of corticosteroids or other kinase inhibitors. In the subsequent sections, we will provide an in-depth analysis of the findings derived from in vitro, ex vivo, and in vivo investigations, as well as insights obtained from clinical trials.

Insights from in vitro, ex vivo, and in vivo animal models

There has been a growing interest in the clinical efficacy of p38 inhibition for COPD complications. P38 MAPK inhibitors may be a promising treatment choice for COPD. The majority of p38 inhibitors target the alpha and beta isoforms, which have been implicated in the inflammatory processes associated with COPD. The inhibitors include PH-797804, SB-681323 (dilmapimod), SB-239063, RV568, and GW856553 [15, 31, 143, 144]. Isoform-selective p38 MAPK inhibitors exert their effects by binding to the ATP-binding site and selectively inhibiting the alpha and beta isoforms [145].

In an LPS-challenged mouse model, RV568 affected both α and γ isoforms of p38, significantly reducing chemotactic mediators of Th1 and Th17 cytokines, airway remodeling markers, and malondialdehyde, an oxidative stress biomarker [146]. However, as RV568 has a considerable impact on the SRC kinase family (steroid receptor coactivator) and HCKs (hematopoietic cell kinase) in particular, these results cannot be solely attributed to p38 inhibition [146] (Table 3 and Fig. 3). Additionally, human mesenchymal stem cells were used to ameliorate airway inflammation and emphysema by downregulating cyclooxygenase-2 expression (COX-2) and prostaglandin E2 (PGE2) production through the inhibition of p38 and ERK [39] (Fig. 3 and Table 1). Studies have shown that inhibitors of P38 MAPK can effectively reduce airway inflammation, with the p8 MAPK inhibitor SB706504 demonstrating a decreased LPS-induced release of TNF- α from macrophages in vitro. A study demonstrated that simultaneous exposure of PBMCs from COPD patients to both dexamethasone and GW856553 resulted in an increased reduction in CXCL8 levels, suggesting restoration of COPD sensitivity to corticosteroids [42]. Therefore, despite the inadequate response of COPD patients to conventional anti-inflammatory treatments such as corticosteroids, coadministration of p38 inhibitors and corticosteroids may show more promising therapeutic effects.

The coadministration of dexamethasone and the p38 MAPK inhibitor BIRB-796 resulted in greater cytokine suppression in alveolar macrophages than each drug alone. This pharmacological synergism may allow for the use of smaller doses of each drug, aiming to optimize efficacy and minimize adverse effects. Thus, this therapeutic approach may provide insight into the

potential of p38 MAPK inhibition as a real-life treatment for COPD, an area that is currently surrounded by uncertainties [150].

In vivo experiments in mice have revealed that the p38 MAPK inhibitor SD-282 inhibits smoke-induced increases in the number of bronchoalveolar lavage (BAL) neutrophils and macrophages, while steroids have negligible effects. An investigation on the effect of CSE in mice showed that the expression of IL-33 in bronchial endothelial cells and peripheral blood mononuclear cells (PBMCs) of COPD mice is upregulated, leading to enhanced production of IL-6 and interleukin-8 (IL-8). Importantly, the study demonstrated that p38 MAPK inhibitors effectively reduced the secretion of IL-6 and IL-8 in PBMCs and controlled systemic inflammation in COPD [151].

In another study, the association between oxidative stress and structural alterations in muscles in elastaseinduced emphysema mouse models was investigated, focusing on the effect of p38 inhibitors. Activation of the p38 MAPK signaling pathway in the soleus muscles led to the activation of the ubiquitin–proteasome system and autophagy, contributing to muscle atrophy. However, treatment with astaxanthin or p38 inhibitors mitigated the muscle structural changes by deactivating the p38 MAPK signaling pathway. These findings provide insight into the role of oxidative stress in muscle alterations in COPD and suggest a potential target for sarcopenia in this context [33].

The effects of cigarette smoke extract (CSE) on macrophage apoptosis and the underlying mechanisms showed that CSE induced apoptosis in macrophages and increased the expression of cleaved caspase 3. Additionally, CSE activated the endoplasmic reticulum stress (ERS) pathway and increased the phosphorylation levels of p38, JNK, and ERK1/2. Importantly, the inhibition of p38 significantly reduced CSE-induced apoptosis, while the inhibition of ERK1/2 promoted apoptosis. Furthermore, CSE increased intracellular calcium levels, and a calcium chelator partially attenuated the apoptosis and phosphorylation of P38 and STAT1 induced by CSE. These findings suggest that CSE induces caspase 3-dependent apoptosis in macrophages through ERS and the intracellular calcium/P38/STAT1 pathway, highlighting the potential role of P38 inhibitors as a therapeutic target in COPD [152].

Additionally, studying the exacerbation in a COPD mouse model by mimicking viral infection (TLR3 ligand) and cigarette smoke (CS) exposure revealed that the TLR3 ligand enhanced the expression of inflammatory cytokines and remodeling factors in pulmonary epithelial cells. However, treatment with p38 inhibitors effectively reduced the expression of inflammatory cytokines and Table 3 Genes and proteins that are affected downstream of p38 signaling within in vivo models of COPD

Gene	Source	Treatment	Case/control	Up/down ^a	Ref
MMP-12	COPD model in Mice (BALF sample)	CS+LPS	CS+LPS/PBS	Up-P	[147]
	Emphysema mice model (BALF sample)	CS+LPS	Emphysema mice/WT	Up-P	[147]
	Lung tissue (emphysema mice model)	CS+SB203580	CS+SB203580/CS	Down-G	[148]
IL-16	COPD model in Mice (BALF sample)	CS+LPS	CS+LPS/PBS	Up-P	[147]
	Emphysema mice model (BALF sample)	CS+LPS	Emphysema mice/WT	Up-P	[147]
IL-27	COPD model in Mice (BALF sample)	CS+LPS	CS+LPS/PBS	Up-P	[147]
	Emphysema mice model (BALF sample)	CS+LPS	Emphysema mice/WT	Up-P	[147]
Leptin	COPD model in Mice (BALF sample)	CS+LPS	CS+LPS/PBS	Up-P	[147]
	Emphysema mice model (BALF sample)	CS+LPS	Emphysema mice/WT	Up-P	[147]
TCA-3	COPD model in Mice (BALF sample)	CS+LPS	CS+LPS/PBS	Up-P	[147]
	Emphysema mice model (BALF sample)	CS+LPS	Emphysema mice/WT	Up-P	[147]
KC (CXCL-1)	COPD model in Mice (BALF sample)	CS+LPS	CS+LPS/PBS	Up-P	[147]
	Emphysema mice model (BALF sample)	CS+LPS	Emphysema mice/WT	Up-P	[147]
	Lung tissue (emphysema mice model)	CS+SB203580	CS+SB203580/CS	Down-G	[55]
CRP	Emphysema mice model (BALF sample)	CS+LPS	Emphysema mice/WT	Up-P	[147]
	Circulating chemokine (COPD GOLD II)	GW856553	GW856553/Plasebo	Down-P	[149]
	Human COPD (GOLD II/III)	PH-797804	PH-797804/Placebo	Down-P	[55]
ITAC	Emphysema mice model (BALF sample)	CS+LPS	Emphysema mice/WT	Up-P	[147]
Adiponectin	Emphysema mice model (BALF sample)	CS+LPS	Emphysema mice/WT	Up-P	[147]
Prolactin	Emphysema mice model (BALF sample)	CS+LPS	Emphysema mice/WT	Up-P	[147]
uPAR	Emphysema mice model (BALF sample)	CS+LPS	Emphysema mice/WT	Up-P	[147]
FAM-174A	Whole blood (Human COPD)	Dilmapimod	Dilmapimod/placebo	Down-G	[54]
ADM	Whole blood (Human COPD)	Dilmapimod	Dilmapimod/placebo	Down-G	[54]
CCI-8	Whole blood (Human COPD)	Dilmapimod	Dilmapimod/placebo	Down-G	[54]
CXCI-1	Whole blood (Human COPD)	Dilmapimod	Dilmapimod/placebo	Down-G	[54]
II-1B	Whole blood (Human COPD)	Dilmapimod	Dilmapimod/placebo	Down-G	[54]
· - · •	l ung tissue (emphysema mice model)	CS + SB203580	CS + SB203580/CS	Down-G	[55]
TI R-1	Whole blood (Human COPD)	Dilmapimod	Dilmapimod/placebo	Down-G	[54]
7NF-519	Whole blood (Human COPD)	Dilmanimod	Dilmapimod/placebo	Down-G	[54]
MMP-9	Whole blood (Human COPD)	Dilmapimod	Dilmapimod/placebo	Down-G	[54]
	Guinea Pig BAL fluid	SB 239063 + LPS	SB 239063 + 1 PS/1 PS	Down-P	[40]
COX-2	CS exposed rat	MSC+CS	MSC + CS/CS	Down-P	[39]
PGF-2	CS exposed rat	MSC+CS	MSC+CS/CS	Down-P	[39]
CXCL-8	Septum (human COPD)	-	COPD/healthy	Up-G&P	[143]
MMP-2	l ung tissue (emphysema mice model)	CS+SB203580	CS + SB203580/CS	Down-G	[55]
TNFa	l ung tissue (emphysema mice model)	CS + SB203580	CS + SB203580/CS	Down-G	[55]
	alveolar macrophages (COPD GOLD IV)	LPS+AZD7624	COPD/healthy	Down-P	[144]
	Rat whole blood	SB-239063+1PS	SB 239063 ± 1 PS/1 PS	Down-P	[40]
MIP-1a	lung tissue (emphysema mice model)	CS + SB203580	CS+SB203580/CS	Down-G	[55]
II-6	Lung tissue (emphysema mice model)	CS + SB203580	CS + SB203580/CS	Down-G	[55]
			COPD/bealthy	Down-P	[33]
	Circulating champling (COPD GOLD II)	CW856553	CVI D/ Healthy	Down-P	[144]
	Guinea Pig BAL fluid	SB-230063 ±1 PC	SB 239063 ± 1 PS/1 PS	Down-P	[/ /]
Fibringgen	Circulating chemokine (COPD GOLD II)	GW856553	GW/856553/Placabo	Down-P	[+0] [1/0]
11-8		GW050555	CW/856552 /Discobo		[140]
		CR_601272	CR-681323/Placebo		[149] [77]
I IJF=27	Circulating chemokine (COPD GOLD II)	20-00-023	30-001323/FIdCEDU	DOWIFF	[44]

^a Down Downregulation, Up Upregulation, P Protein, G Gene

remodeling factors while increasing the expression of E-cadherin. Additionally, p38 inhibitors attenuated airway hyperreactivity induced by the TLR3 ligand. These findings suggest that targeting the TLR3/p38 pathway could be a potential therapeutic approach for managing airway inflammation and remodeling in COPD [153].

To investigate the interaction of inflammation and oxidative stress with protease-antiprotease imbalance, a mouse model was exposed to CSE, which showed an enhanced inflammatory response induced by neutrophil elastase (NE) in bronchial epithelial cells. This enhancement was found to be dependent on the activation of proteinase-activated receptor 2 (PAR2) and the extracellular signal-regulated kinase (ERK) pathway. Interestingly, inhibition of p38 MAPK attenuated the upregulation of PAR2 and the subsequent increase in NE-induced interleukin-8 (IL-8) production [154].

Insights from clinical trials

Due to the predominant expression of p38 α and β proteins in the lungs of COPD patients, therapeutic strategies and clinical trials have primarily focused on these p38 isoforms [15, 31]. There is a negligible chance of cross-interaction for α and β p38 isoforms with their specific inhibitors [155]. Although initial efforts and clinical trials of orally administered p38 inhibitors were made, p38 oral compounds were discontinued for further trials with the emergence of inhaled formulations, except Novartis' Acumapimod, an oral inhibitor of p38α (IC50 < 1 µM) [156, 157]. A study investigated acumapimod as a part of an ongoing clinical trial (NCT02700919) for the treatment of AECOPD patients (acute exacerbations of COPD). The study showed that short-term therapy with acumapimod could improve lung function indices and decrease inflammation biomarkers, clinical outcomes, and length of hospital stay. Thus, acumapimod was suggested as a potential drug for the treatment of AECOPD, which can reduce the healthcare burden and improve patients' quality of life [158].

In contrast, losmapimod and dilmapimod, which are oral medications, were withdrawn from phase II clinical trials due to their disappointing results [56, 156, 159]. Similarly, the oral application of losmapimod failed to reduce the frequency of AECOPD [160]. However, a study showed that losmapimod significantly reduced the plasma level of fibrinogen in these patients [149] (Table 4). In contrast, inhaled p38 inhibitors have been suggested to reduce the systemic side effects of COPD medications [161, 162]. Table 4 summarizes the p38 inhibitors that have undergone phase II clinical trials, along with their clinical outcomes. For example, a clinical trial on the effectiveness and safety of inhaled PH-797804 in patients with COPD GOLD stage II/III [55] showed

an improvement in prebronchodilator FEV1 and the transitional dyspnea index (TDI) [55] (Table 4).

In an investigation of AZD7624, an inhaled $p38\alpha/\beta$ inhibitor, for treating COPD patients, it was discovered that while the inhibitor significantly reduced TNF- α and neutrophils in sputum, there was no statistically significant disparity between AZD7624 and placebo among 213 COPD patients concerning the number of days to the first moderate or severe exacerbation or early dropout [144]. Another randomized controlled trial of AZD7624 revealed that although the administration of this inhibitor was safe and well tolerated, it did not offer any clinical advantages in terms of COPD exacerbation management and general lung functions in infrequently exacerbated patients [166] (Table 4).

During a phase II clinical trial, RV-568, also known as JNJ-49095397, a narrow-spectrum inhaled p38 inhibitor, was found to successfully inhibit proinflammatory cytokines in macrophages, airway epithelial cells, and smooth muscle cells [167, 168]. The administration of 100 µg/day RV-568 was well tolerated in COPD patients for seven days [169]. Studies have shown that inhaled RV-568 significantly improves the FEV1 index after 14 days and reduces sputum malondialdehyde and myeloperoxidase levels [170]. Furthermore, another study indicated that RV-568 inhibits the expression of IL-1 β and CXCL-8 cytokines by 90% and 73%, respectively, at the highest administered doses [171]. Moreover, other studies have shown the anti-inflammatory properties and lung function benefits of RV-568, which are synergistically augmented by corticosteroid administration [146] (Table 4).

In a clinical study of patients with GOLD II/III COPD, the oral p38 inhibitor PH-797804 was found to reduce the levels of CRP, a systemic inflammatory biomarker, and significantly improve lung function after 6 weeks of administration. However, there was no significant difference in the levels of CC16, IL-6, surfactant protein D (SPD), or fibrinogen compared to placebo controls [55] (Table 3 and Fig. 3). Another oral p38 inhibitor, SB-681323, has been shown to decrease TNF- α levels in LPS-stimulated blood samples of COPD patients, while prednisolone had no effect on TNF- α levels in these patients [44] (Table 3 and Fig. 3). A transcriptomic study of blood inflammatory factors in COPD patients found that a single dose of oral SB-681323 downregulated several inflammatory factors associated with p38 inhibition by mediating the involvement of IL-1 β and STAT1 in [54].

Clinical trials of p38 MAPK inhibitors in humans have shown that these inhibitors decrease serum levels of both TNF- α and IL-6 after LPS administration, as well as acute phase reactants associated with inflammation, such as serum erythrocyte sedimentation rate (ESR) and c-reactive protein (CRP) [145].

Table 4 List of clinical trials assessing the efficacy of different p38 inhibitors in COPD

NCT Number ^a	Status	last update	Study Results	Interventions	Dosing	Phases	Enrollment	Publication
NCT02815488	Terminated	March 2019	NA ^b	CHF6297	Single and repeated doses	Phase 2	118	[150]
NCT00642148	Completed	July 27, 2009	NA	GW856553 (Losmapi- mod)	12-weeks of treatment with GW856553 7.5 mg twice daily	Phase 2	306	[149]
NCT00144859	Completed	July 2005	NA	SB681323 (Dilmapi- mod)	7.5 mg Per Day	Phase 2	82	NA
NCT02366637	Terminated	May 2015	With Results	PF-03715455 (aka compound 1ab)	Orally inhaled placebo twice a day (BID) for 4 weeks	Phase 2	13	NA
NCT01541852	Completed	February 2014	NA	GW856553 (Losmapi- mod)	7.5 mg tablet twice daily	Phase 2	72	[163]
NCT01332097	Completed	May 2013	NA	BCT197 (Acumapi- mod)	single 75 mg oral dose of BCT197 capsules	Phase 2	183	[164]
NCT01218126	Completed	December 21, 2011	With Results	GW856553 (Losmapi- mod)	three doses of los- mapimod (2.5 mg, 7.5 mg and 15 mg) twice daily	Phase 2	604	[56, 159]
NCT02299375	Completed	December 21, 2011	With Results	GW856553 (Losmapi- mod)	15 mg (mg) twice daily	Phase 2	184	[160]
NCT00559910	Completed	June 30, 2016	NA	PH-797804	0.5, 3, 6 or 10 mg PH-797804 once daily and treated for 6 weeks follow- ing a 2-week run-in	Phase 2	230	[55]
NCT01937338	Completed	April 2014	NA	AZD7624	nebulizer solution; 20 mg/mL for inhala- tion	Phase 1	60	[144]
NCT02238483	Completed	April 4, 2016	With Results	AZD7624	Inhaled AZD7624 solution, 11 mg/mL	Phase 2	213	[144]
NCT01475292	Completed	April 2012	NA	RV568	RV568 50 and 100 ug administered via a nebulizer once daily for 14 days	Phase 2	30	[146]
NCT01703052	Completed	May 2012	NA	CHF6001	a single-dose dry-powder inhaler including 20, 100, 200, 400, 800, 1,600, and 2,000 µg and 2,400, 4,000, and 4,800 µg	Phase 1	74	[165]
NCT02386761	Completed	May 2015	NA	CHF6001	a single-dose dry-powder inhaler including 2,400, 4,000, and 4,800 µg	Phase 1	48	[165]
NCT00439881	Completed	June 2012	NA	SB-681323	single intravenous dose	Phase 1	16	[21]

^a ClinicalTrials.gov Identifier

^b NA Not available

A recently published systematic review and metaanalysis evaluated the safety and efficacy of p38 MAPKIs as an alternative to corticosteroids in the treatment of COPD in ten randomized controlled trials (RCTs) involving a total of 1,751 participants [44, 55, 56, 144, 146, 149, 158, 160, 172, 173]. The analysis focused on various outcome measures, including lung function, inflammatory biomarkers, and quality of life [174]. The results indicated that while p38 MAPKIs were found to be safe, they did not demonstrate significant effects compared to placebo

in terms of efficacy. In summary, while the present systematic review and meta-analysis suggest that p38 MAP-KIs do not demonstrate significant efficacy compared to placebo in COPD treatment, it is imperative to exercise caution when interpreting these findings due to the limited number of included studies and the heterogeneity resulting from combining different p38 MAPKIs. Further research is warranted to provide a more comprehensive understanding of the role and potential benefits of p38 MAPKIs in the management of COPD.

Conclusion

In the context of COPD, p38 inhibition has been observed to effectively mitigate inflammation and oxidative stress in the pulmonary and muscular tissues of affected individuals, which are closely associated with airflow restriction, respiratory distress, and reduced physical tolerance [55, 174]. Moreover, the p38 inhibitors have demonstrated the capability to improve key lung function parameters and alleviate dyspnea in COPD patients, as assessed through measurements such as forced expiratory volume in 1 s (FEV1), forced vital capacity (FVC), and transition dyspnea index (TDI) [175]. Notably, p38 inhibitors appear to offer superior therapeutic outcomes compared to steroids for specific subsets of COPD patients, given that steroids possess limited antiinflammatory efficacy and are not recommended as standalone treatments for COPD [55, 174]. However, it is important to acknowledge that p38 inhibitors may entail undesirable adverse effects, including liver toxicity and cutaneous reactions, which impose restrictions on their clinical usage and raise concerns regarding their safety [16]. Additionally, the transient efficacy of p38 inhibitors, which tends to dissipate within a matter of weeks, can be attributed to the intricate and redundant nature of the inflammatory network. Inhibition of one pathway may inadvertently activate alternative pathways that perpetuate inflammation. Furthermore, the effects of p38 inhibitors can vary among different subgroups of COPD patients, contingent upon their specific inflammatory phenotypes and the severity of their disease. The longterm implications of p38 inhibitors on COPD progression and mortality remain uncertain since most clinical trials have focused primarily on short-term assessments of lung function and symptomatology [16, 174].

Additionally, activation of p38 in distinct cell types, including macrophages and neutrophils, can induce inflammatory responses and tissue damage, thereby contributing to the pathogenesis of COPD. For example, p38 activation in these inflammatory cells can elicit the release of pro-inflammatory cytokines and chemokines, perpetuating the chronic inflammation observed in COPD. In such cases, inhibiting p38 signaling holds potential for reducing excessive inflammation and attenuating disease progression. However, it is imperative to acknowledge that the p38 signaling pathway also assumes indispensable roles in cellular processes that are fundamental for maintaining tissue homeostasis and facilitating repair mechanisms. For instance, activation of p38 in epithelial cells has been shown to enhance cell survival, preserve the integrity of the epithelial barrier, and facilitate tissue repair mechanisms. Consequently, inhibiting p38 signaling in these cells has the potential to disrupt these advantageous activities and compromise the regenerative capacity of lung tissue. Therefore, while the inhibition of p38 signaling may manifest anti-inflammatory effects, it is crucial to consider the potential ramifications on other cell functions and the delicate equilibrium between inflammation and tissue repair in the context of COPD. The challenge lies in selectively targeting the detrimental aspects of p38 signaling without compromising the beneficial functions, necessitating further research and a more comprehensive understanding of the intricate interplay between diverse cell types and signaling pathways implicated in COPD pathogenesis [113].

Finally, it is crucial to recognize that COPD is a multifaceted and intricate disease with a plethora of underlying mechanisms that contribute to its onset and progression. While inflammation, oxidative stress, and imbalances in tissue repair are pivotal factors in the pathogenesis of COPD, it is imperative to acknowledge the involvement of other elements as well, including genetic predisposition, environmental exposures, and immune dysregulation. COPD should not be regarded solely as an inflammatory condition; however, inflammation undeniably plays a significant role in its development and advancement. Persistent inflammation in the airways and lung tissue is a defining characteristic of COPD and contributes to the manifestation of symptoms and complications. Nonetheless, it is vital to note that COPD encompasses a complex interplay of multiple underlying mechanisms beyond inflammation. In addition to inflammation, COPD involves oxidative stress, imbalances in protease-antiprotease activity, impaired tissue repair processes, and structural alterations in the lungs, among other factors. Consequently, focusing solely on inhibiting inflammation may not comprehensively address all facets of the disease and could potentially lead to a misleading perspective.

As stated, the impaired clearance of senescent cells in the lungs of COPD patients, including senescent epithelial cells and inflammatory cells such as macrophages and neutrophils, can lead to their accumulation. This accumulation poses a challenge in COPD, as these senescent cells contribute to chronic inflammation and tissue damage. Dysregulated signaling pathways, including the activation of p38, play a crucial role in driving inflammation and worsening the disease. Therefore, targeting p38 signaling has emerged as a potential therapeutic approach for COPD. Inhibiting p38 activation can help reduce the release of inflammatory mediators and mitigate the inflammatory cascade, leading to a reduction in tissue damage and improvement in lung function. However, it is important to exercise caution when inhibiting p38, as it also plays critical roles in essential cellular processes required for tissue homeostasis and repair. Striking a balance between suppressing inflammation and preserving beneficial cellular functions is crucial when considering p38 as a therapeutic target in COPD management.

By unraveling the intricate mechanisms at play, we can develop more targeted and effective therapies that selectively modulate p38 activity, striking a balance between suppressing inflammation and preserving beneficial cellular functions for optimal COPD management. To effectively manage COPD, treatment strategies should be aimed at restoring the equilibrium of tissue repair mechanisms, alleviating oxidative stress, and targeting specific molecular pathways implicated in COPD progression. The utilization of combinatorial therapies, involving the concurrent use of multiple therapeutic agents targeting distinct aspects of the disease, is gaining prominence in COPD management. This approach recognizes the multifaceted nature of COPD and strives to address the diverse mechanisms contributing to disease progression. By concurrently targeting repair imbalances, reducing inflammation, and addressing other factors, combinatorial therapies have the potential to offer more comprehensive and effective treatment outcomes. It is important to note that the development and implementation of combinatorial therapies necessitate extensive research, clinical trials, and personalized approaches to tailor treatment strategies to individual patients. The ultimate objective is to achieve a delicate balance between suppressing inflammation and promoting tissue repair and homeostasis to optimize outcomes for individuals with COPD.

Abbreviations

Ams	Alveolar macrophages
ARG1	Arginase 1
ASK1	Apoptosis signal-regulating kinase 1
BAL	Bronchoalveolar lavage
CCL5	Chemokine (C-C motif) ligand 5
COPD	Chronic obstructive pulmonary disease
COX-2	Cyclooxygenase-2 expression
CSE	Cigarette smoke extract
DUSPs	Dual-specificity phosphatases
ECM	Extracellular matrix
EndMT	Endothelial-to-mesenchymal transition
EPAS1	Domain-containing protein 1
ERS	Endoplasmic reticulum stress

FEV1	Forced expiratory volume in 1 s
FVC	Forced vital capacity
GM-CSF	Granulocyte-macrophage colony-stimulating factor
HASMC	Human airway smooth muscle cells
HCKs	Hematopoietic cell kinase
HSP27	Heat Shock Protein 27
IFN-γ	Interferon gamma
IL-1β	Interleukin 1 beta
IL-6	Interleukin 6
IL-8	Interleukin 8
IMs	Interstitial macrophages
LABAs	Long-acting β2-agonists
LAMAs	Long-acting muscarinic antagonists
LPS	Lipopolysaccharide
MAP3K1	Mitogen-activated protein kinase kinase kinase 1
MKPs	MAP kinase phosphatases
MLK3	Mixed lineage kinase 3
Mo-AMs	Monocyte-derived alveolar macrophages
mRNA	Messenger RNA
MSCs	Mesenchymal stem cells
PBMCs	Peripheral blood mononuclear cells
PGE2	Prostaglandin E2
PP2A/C	Protein phosphatase 2A/C
RANTES	Regulated on activation, normal T-cell expressed and secreted
ROS	Reactive oxygen species
SMCs	Smooth muscle cells
SPD	Surfactant protein D
SRC	Steroid Receptor Coactivator
TAK1	TGF-β-activated kinase 1
TDI	Transition dyspnea index
TGF-β	Transforming growth factor β
TLR4	Toll-like receptor 4
TNF-α	Tumor necrosis factor-alpha
TR-AMs	Tissue-resident alveolar macrophages
VEGF	Vascular endothelial growth factor
WHO	World Health Organization
Wip1	Wild-type p53-induced phosphatase 1

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Authors' contributions

A.A., S.A., J.S. Z.S., and M.K. designed the study. S.A., A.A, and S.A. wrote the manuscript. M.K., Z.S. and A.E critically commented and structured the clinical data of the manuscript. All authors reviewed the manuscript.

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Competing interests

The authors declare no competing interests.

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References

- 1. Venkatesan P. GOLD COPD report: 2023 update. Lancet Respir Med. 2023;11(1):18.
- World Health Organisation (WHO). Chronic obstructive pulmonary disease (COPD). 2023. Available from: https://www.who.int/news-room/ fact-sheets/detail/chronic-obstructive-pulmonary-disease-(copd).
- 3. Raoof S, Shah M, Make B, Allaqaband H, Bowler R, Fernando S, et al. Lung imaging in COPD part 1: clinical usefulness. Chest. 2023;164(1):69–84.
- Agustí A, Melén E, DeMeo DL, Breyer-Kohansal R, Faner R. Pathogenesis of chronic obstructive pulmonary disease: understanding the contributions of gene–environment interactions across the lifespan. Lancet Respir Med. 2022;10(5):512–24.
- Lange P, Celli B, Agustí A, Boje Jensen G, Divo M, Faner R, et al. Lungfunction trajectories leading to chronic obstructive pulmonary disease. N Engl J Med. 2015;373(2):111–22.
- Pellegrino D, Casas-Recasens S, Faner R, Palange P, Agusti A. When GETomics meets aging and exercise in COPD. Respir Med. 2023;216:107294.
- Agustí A, Celli BR, Criner GJ, Halpin D, Anzueto A, Barnes P, et al. Global initiative for chronic obstructive lung disease 2023 report: GOLD executive summary. Am J Respir Crit Care Med. 2023;207(7):819–37.
- Gamble E, Grootendorst DC, Hattotuwa K, O'Shaughnessy T, Ram FS, Qiu Y, et al. Airway mucosal inflammation in COPD is similar in smokers and ex-smokers: a pooled analysis. Eur Respir J. 2007;30(3):467–71.
- 9. Zhao X, Mehrabi R, Xu J-R. Mitogen-activated protein kinase pathways and fungal pathogenesis. Eukaryot Cell. 2007;6(10):1701–14.
- Cuenda A, Rousseau S. p38 MAP-kinases pathway regulation, function and role in human diseases. Biochim Biophys Acta Mol Cell Res. 2007;1773(8):1358–75.
- Pelaia C, Vatrella A, Gallelli L, Lombardo N, Sciacqua A, Savino R, et al. Role of p38 mitogen-activated protein kinase in asthma and COPD: pathogenic aspects and potential targeted therapies. Drug Des Dev Ther. 2021;15:1275–84.
- Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. BMJ. 2021;372:n71.
- Yang Y, Kim SC, Yu T, Yi YS, Rhee MH, Sung GH, et al. Functional roles of p38 mitogen-activated protein kinase in macrophage-mediated inflammatory responses. Mediators Inflamm. 2014;2014:352371.
- Asih PR, Prikas E, Stefanoska K, Tan ARP, Ahel HI, Ittner A. Functions of p38 MAP kinases in the central nervous system. Front Mol Neurosci. 2020;13:570586.
- 15. Renda T, Baraldo S, Pelaia G, Bazzan E, Turato G, Papi A, et al. Increased activation of p38 MAPK in COPD. Eur Respir J. 2008;31(1):62–9.
- 16. Singh D. P38 inhibition in COPD; cautious optimism. Thorax. 2013;68(8):705–6.
- 17. Barnes PJ. Targeting cytokines to treat asthma and chronic obstructive pulmonary disease. Nat Rev Immunol. 2018;18(7):454–66.
- 18. Kirkham PA, Barnes PJ. Oxidative stress in COPD. Chest. 2013;144(1):266–73.

- Lim TG, Kim JE, Jung SK, Li Y, Bode AM, Park JS, et al. MLK3 is a direct target of biochanin A, which plays a role in solar UV-induced COX-2 expression in human keratinocytes. Biochem Pharmacol. 2013;86(7):896–903.
- 20. Cuadrado A, Nebreda AR. Mechanisms and functions of p38 MAPK signalling. Biochemical journal. 2010;429(3):403–17.
- Fisk M, Gajendragadkar PR, Maki-Petaja KM, Wilkinson IB, Cheriyan J. Therapeutic potential of p38 MAP kinase inhibition in the management of cardiovascular disease. Am J Cardiovasc Drugs. 2014;14(3):155–65.
- Martinez-Limon A, Joaquin M, Caballero M, Posas F, de Nadal E. The p38 pathway: from biology to cancer therapy. Int J Mol Sci. 2020;21(6):1913.
- Gaffey K, Reynolds S, Plumb J, Kaur M, Singh D. Increased phosphorylated p38 mitogen-activated protein kinase in COPD lungs. Eur Respir J. 2013;42(1):28–41.
- Brennan CM, Emerson CP Jr, Owens J, Christoforou N. p38 MAPKs roles in skeletal muscle physiology, disease mechanisms, and as potential therapeutic targets. JCI Insight. 2021;6(12):e149915.
- Zarubin T, Han J. Activation and signaling of the p38 MAP kinase pathway. Cell Res. 2005;15(1):11–8.
- 26. Ashwell JD. The many paths to p38 mitogen-activated protein kinase activation in the immune system. Nat Rev Immunol. 2006;6(7):532.
- Salvador JM, Mittelstadt PR, Guszczynski T, Copeland TD, Yamaguchi H, Appella E, et al. Alternative p38 activation pathway mediated by T cell receptor–proximal tyrosine kinases. Nat Immunol. 2005;6(4):390.
- Pramanik R, Qi X, Borowicz S, Choubey D, Schultz RM, Han J, et al. p38 lsoforms have opposite effects on AP-1-dependent transcription through regulation of c-Jun the determinant role of the isoforms in the p38 MAPK signal specificity. J Biol Chem. 2003;278(7):4831–9.
- Qi X, Pohl NM, Loesch M, Hou S, Li R, Qin J-Z, et al. p38α antagonizes p38γ activity through c-Jun-dependent ubiquitin-proteasome pathways in regulating Ras transformation and stress response. J Biol Chem. 2007;282(43):31398–408.
- Beardmore VA, Hinton HJ, Eftychi C, Apostolaki M, Armaka M, Darragh J, et al. Generation and characterization of p38β (MAPK11) gene-targeted mice. Mol Cell Biol. 2005;25(23):10454–64.
- 31. Chung KF. p38 mitogen-activated protein kinase pathways in asthma and COPD. Chest. 2011;139(6):1470–9.
- 32. Wydra VR, Ditzinger RB, Seidler NJ, Hacker FW, Laufer SA. A patent review of MAPK inhibitors (2018 present). Expert Opin Ther Pat. 2023;33(6):421–44.
- Mano Y, Tsukamoto M, Wang KY, Nabeshima T, Kosugi K, Tajima T, et al. Oxidative stress causes muscle structural alterations via p38 MAPK signaling in COPD mouse model. J Bone Miner Metab. 2022;40(6):927–39.
- Gao W, Li L, Wang Y, Zhang S, Adcock IM, Barnes PJ, et al. Bronchial epithelial cells: the key effector cells in the pathogenesis of chronic obstructive pulmonary disease? Respirology. 2015;20(5):722–9.
- Pérez-Padilla R, Schilmann A, Riojas-Rodriguez H. Respiratory health effects of indoor air pollution. Int J Tuberc Lung Dis. 2010;14(9):1079–86.
- 36. Barnes PJ. The cytokine network in chronic obstructive pulmonary disease. Am J Respir Cell Mol Biol. 2009;41(6):631–8.
- Kent LM, Smyth LJ, Plumb J, Clayton CL, Fox SM, Ray DW, et al. Inhibition of lipopolysaccharide-stimulated chronic obstructive pulmonary disease macrophage inflammatory gene expression by dexamethasone and the p38 mitogen-activated protein kinase inhibitor N-cyano-N'-(2-[[8-(2, 6-difluorophenyl)-4-(4-fluoro-2-methylphenyl)-7-oxo-7, 8-dihydropyrido [2, 3-d] pyrimidin-2-yl] amino] ethyl) guanidine (SB706504). J Pharmacol Exp Ther. 2009;328(2):458–68.
- Armstrong J, Harbron C, Lea S, Booth G, Cadden P, Wreggett K, et al. Synergistic effects of p38 mitogen-activated protein kinase inhibition with a corticosteroid in alveolar macrophages from patients with chronic obstructive pulmonary disease. J Pharmacol Exp Ther. 2011;338(3):732–40.
- Gu W, Song L, Li X-M, Wang D, Guo X-J, Xu W-G. Mesenchymal stem cells alleviate airway inflammation and emphysema in COPD through down-regulation of cyclooxygenase-2 via p38 and ERK MAPK pathways. Sci Rep. 2015;5:8733.
- Underwood DC, et al. SB 239063, a p38 MAPK inhibitor, reduces neutrophilia, inflammatory cytokines, MMP-9, and fibrosis in lung. Am J Physiol Lung Cell Mol Physiol. 2000;279(5):L895–902.
- Ratcliffe MJ, Dougall IG. Comparison of the anti-inflammatory effects of Cilomilast, Budesonide and a p38 Mitogen activated protein kinase

inhibitor in COPD lung tissue macrophages. BMC Pharmacol Toxicol. 2012;13(1):15.

- Khorasani N, Baker J, Johnson M, Chung KF, Bhavsar PK. Reversal of corticosteroid insensitivity by p38 MAPK inhibition in peripheral blood mononuclear cells from COPD. Int J Chron Obstruct Pulmon Dis. 2015;10:283.
- Meng A, Zhang X, Wu S, Wu M, Li J, Yan X, et al. In vitro modeling of COPD inflammation and limitation of p38 inhibitor–sB203580. Int J Chron Obstruct Pulmon Dis. 2016;11:909.
- 44. Singh D, Smyth L, Borrill Z, Sweeney L, Tal-Singer R. A randomized, placebo-controlled study of the effects of the p38 MAPK inhibitor SB-681323 on blood biomarkers of inflammation in COPD patients. J Clin Pharmacol. 2010;50(1):94–100.
- Jang MK, Kim S-H, Lee K-Y, Kim T-B, Moon KA, Park CS, et al. The tyrosine phosphatase, SHP-1, is involved in bronchial mucin production during oxidative stress. Biochem Biophys Res Commun. 2010;393(1):137–43.
- 46. Shahabuddin S, Ji R, Wang P, Brailoiu E, Dun N, Yang Y, et al. CXCR3 chemokine receptor-induced chemotaxis in human airway epithelial cells: role of p38 MAPK and PI3K signaling pathways. Am J Physiol Cell Physiol. 2006;291(1):C34–9.
- Yang J-j, Wang D-d, Sun T-y. Flagellin of Pseudomonas aeruginosa induces transforming growth factor beta 1 expression in normal bronchial epithelial cells through mitogen activated protein kinase cascades. Chin Med J. 2011;124(4):599–605.
- Roos-Engstrand E, Wallin A, Bucht A, Pourazar J, Sandström T, Blomberg A. Increased expression of p38 MAPK in human bronchial epithelium after lipopolysaccharide exposure. Eur Respir J. 2005;25(5):797–803.
- Durham AL, McLaren A, Hayes BP, Caramori G, Clayton CL, Barnes PJ, et al. Regulation of Wnt4 in chronic obstructive pulmonary disease. FASEB J. 2013;27(6):2367–81.
- Vallese D, Ricciardolo FL, Gnemmi I, Casolari P, Brun P, Sorbello V, et al. Phospho-p38 MAPK expression in COPD patients and asthmatics and in challenged bronchial epithelium. Respiration. 2015;89(4):329–42.
- Alfaro-Moreno E, Martínez L, García-Cuellar C, Bonner JC, Murray JC, Rosas I, et al. Biologic effects induced in vitro by PM10 from three different zones of Mexico City. Environ Health Perspect. 2002;110(7):715.
- 52. Faner R, Tal-Singer R, Riley JH, Celli B, Vestbo J, MacNee W, et al. Lessons from ECLIPSE: a review of COPD biomarkers. Thorax. 2014;69(7):666–72. https://thorax.bmj.com/content/69/7/666.
- 53. Agassandian M, Shurin GV, Ma Y, Shurin MR. C-reactive protein and lung diseases. Int J Biochem Cell Biol. 2014;53:77–88.
- Betts JC, Mayer RJ, Tal-Singer R, Warnock L, Clayton C, Bates S, et al. Gene expression changes caused by the p38 MAPK inhibitor dilmapimod in COPD patients: analysis of blood and sputum samples from a randomized, placebo-controlled clinical trial. Pharmacol Res Perspect. 2015;3(1):e00094.
- MacNee W, Allan RJ, Jones I, De Salvo MC, Tan LF. Efficacy and safety of the oral p38 inhibitor PH-797804 in chronic obstructive pulmonary disease: a randomised clinical trial. Thorax. 2013;68(8):738–45. https:// thorax.bmj.com/content/68/8/738.short.
- Watz H, Barnacle H, Hartley BF, Chan R. Efficacy and safety of the p38 MAPK inhibitor losmapimod for patients with chronic obstructive pulmonary disease: a randomised, double-blind, placebo-controlled trial. Lancet Respir Med. 2014;2(1):63–72.
- Raidl M, Sibbing B, Strauch J, Müller K, Nemat A, Schneider PM, et al. Impaired TNFα-induced VEGF expression in human airway smooth muscle cells from smokers with COPD: role of MAPkinases and histone acetylation—effect of dexamethasone. Cell Biochem Biophys. 2007;49(2):98–110.
- Sakao S, Tatsumi K, Hashimoto T, Igari H, Shino Y, Shirasawa H, et al. Vascular endothelial growth factor and the risk of smoking-related COPD. Chest. 2003;124(1):323–7.
- 59. Elias JA, Kang MJ, Crouthers K, Homer R, Lee CG. State of the art. Mechanistic heterogeneity in chronic obstructive pulmonary disease: insights from transgenic mice. Proc Am Thorac Soc. 2006;3(6):494–8.
- Leigh R, Oyelusi W, Wiehler S, Koetzler R, Zaheer RS, Newton R, et al. Human rhinovirus infection enhances airway epithelial cell production of growth factors involved in airway remodeling. J Allergy Clin Immunol. 2008;121(5):1238-45.e4.
- 61. Hayashi S, Jibiki I, Asai Y, Gon Y, Kobayashi T, Ichiwata T, et al. Analysis of gene expression in human bronchial epithelial cells upon influenza

virus infection and regulation by p38 mitogen-activated protein kinase and c-Jun-N-terminal kinase. Respirology. 2008;13(2):203–14.

- 62. Heijink I, van Oosterhout A, Kliphuis N, Jonker M, Hoffmann R, Telenga E, et al. Oxidant-induced corticosteroid unresponsiveness in human bronchial epithelial cells. Thorax. 2014;69(1):5–13.
- Zhou F, Onizawa S, Nagai A, Aoshiba K. Epithelial cell senescence impairs repair process and exacerbates inflammation after airway injury. Respir Res. 2011;12(1):78.
- 64. Zeng Q, Zeng J. Inhibition of miR-494-3p alleviates oxidative stressinduced cell senescence and inflammation in the primary epithelial cells of COPD patients. Int Immunopharmacol. 2021;92:107044.
- 65. Tao Y, Sun Y, Wu B, Xu D, Yang J, Gu L, et al. Overexpression of FOXA2 attenuates cigarette smoke-induced cellular senescence and lung inflammation through inhibition of the p38 and Erk1/2 MAPK pathways. Int Immunopharmacol. 2021;94:107427.
- Baker JR, Vuppusetty C, Colley T, Hassibi S, Fenwick PS, Donnelly LE, et al. MicroRNA-570 is a novel regulator of cellular senescence and inflammaging. FASEB J. 2019;33(2):1605–16.
- 67. Hao B, Sun R, Guo X, Zhang L, Cui J, Zhou Y, et al. NOX4-derived ROS promotes collagen I deposition in bronchial smooth muscle cells by activating noncanonical p38MAPK/Akt-mediated TGF-beta signaling. Oxid Med Cell Longev. 2021;2021:6668971.
- Hulina-Tomaskovic A, Heijink IH, Jonker MR, Somborac-Bacura A, Grdic Rajkovic M, Rumora L. Pro-inflammatory effects of extracellular Hsp70 and cigarette smoke in primary airway epithelial cells from COPD patients. Biochimie. 2019;156:47–58.
- 69. Yu Q, Yang D, Chen X, Chen Q. CD147 increases mucus secretion induced by cigarette smoke in COPD. BMC Pulm Med. 2019;19(1):29.
- Liu H, Wang XX, Chen P. Angiopoietin-like 4 knockdown attenuates cigarette smoke extract-induced oxidative stress and apoptosis in lung bronchial epithelial cells by inhibiting NADPH oxidase. Allergol Immunopathol (Madr). 2022;50(5):47–56.
- Byrne AJ, Mathie SA, Gregory LG, Lloyd CM. Pulmonary macrophages: key players in the innate defence of the airways. Thorax. 2015;70(12):1189–96.
- 72. Soriano JB, Polverino F, Cosio BG. What is early COPD and why is it important? Eur Respir J. 2018;52(6):1801448.
- Hou F, Xiao K, Tang L, Xie L. Diversity of macrophages in lung homeostasis and diseases. Front Immunol. 2021;12:753940.
- Traves S, Culpitt S, Russell R, Barnes P, Donnelly L. Increased levels of the chemokines GROα and MCP-1 in sputum samples from patients with COPD. Thorax. 2002;57(7):590–5.
- Liu M, Mendicino M, Ning Q, Ghanekar A, He W, McGilvray I, et al. Cytokine-induced hepatic apoptosis is dependent on FGL2/fibroleukin: the role of Sp1/Sp3 and STAT1/PU. 1 composite cis elements. J Immunol. 2006;176(11):7028–38.
- Liu Y, Xu S, Xiao F, Xiong Y, Wang X, Gao S, et al. The FGL2/fibroleukin prothrombinase is involved in alveolar macrophage activation in COPD through the MAPK pathway. Biochem Biophys Res Commun. 2010;396(2):555–61.
- Marwick JA, Kirkham PA, Stevenson CS, Danahay H, Giddings J, Butler K, et al. Cigarette smoke alters chromatin remodeling and induces proinflammatory genes in rat lungs. Am J Respir Cell Mol Biol. 2004;31(6):633–42.
- Smith S, Fenwick P, Nicholson A, Kirschenbaum F, Finney-Hayward T, Higgins L, et al. Inhibitory effect of p38 mitogen-activated protein kinase inhibitors on cytokine release from human macrophages. Br J Pharmacol. 2006;149(4):393–404.
- Abraham SM, Lawrence T, Kleiman A, Warden P, Medghalchi M, Tuckermann J, et al. Antiinflammatory effects of dexamethasone are partly dependent on induction of dual specificity phosphatase 1. J Exp Med. 2006;203(8):1883–9.
- Bewley MA, Belchamber KB, Chana KK, Budd RC, Donaldson G, Wedzicha JA, et al. Differential effects of p38, MAPK, PI3K or rho kinase inhibitors on bacterial phagocytosis and efferocytosis by macrophages in COPD. PLoS One. 2016;11(9):e0163139.
- He B, et al. Cigarette smoke extract induces the expression of GRP78 in A549 cells via the p38/MAPK pathway. Mol Med Rep. 2013;8(6):1683–8.
- Guan C, Xiao Y, Li K, Wang T, Liang Y, Liao G. MMP-12 regulates proliferation of mouse macrophages via the ERK/P38 MAPK pathways during inflammation. Exp Cell Res. 2019;378(2):182–90.

- Lo CY, Huang HY, He JR, Huang TT, Heh CC, Sheng TF, et al. Increased matrix metalloproteinase-9 to tissue inhibitor of metalloproteinase-1 ratio in smokers with airway hyperresponsiveness and accelerated lung function decline. Int J Chron Obstruct Pulmon Dis. 2018;13:1135–44.
- Vij N, Chandramani-Shivalingappa P, Van Westphal C, Hole R, Bodas M. Cigarette smoke-induced autophagy impairment accelerates lung aging, COPD-emphysema exacerbations and pathogenesis. Am J Physiol Cell Physiol. 2018;314(1):C73–87.
- Winkler AR, Nocka KH, Sulahian TH, Kobzik L, Williams CM. In vitro modeling of human alveolar macrophage smoke exposure: enhanced inflammation and impaired function. Exp Lung Res. 2008;34(9):599–629.
- Fridlender ZG, Sun J, Kim S, Kapoor V, Cheng G, Ling L, et al. Polarization of tumor-associated neutrophil phenotype by TGF-beta: "N1" versus "N2" TAN. Cancer Cell. 2009;16(3):183–94.
- Wculek SK, Malanchi I. Neutrophils support lung colonization of metastasis-initiating breast cancer cells. Nature. 2015;528(7582):413–7.
- Juzenaite G, Secklehner J, Vuononvirta J, Helbawi Y, Mackey JBG, Dean C, et al. Lung marginated and splenic murine resident neutrophils constitute pioneers in tissue-defense during systemic E. coli challenge. Front Immunol. 2021;12:597595.
- Bae GH, Kim YS, Park JY, Lee M, Lee SK, Kim JC, et al. Unique characteristics of lung-resident neutrophils are maintained by PGE2/PKA/Tgm2mediated signaling. Blood. 2022;140(8):889–99.
- Summers C, Rankin SM, Condliffe AM, Singh N, Peters AM, Chilvers ER. Neutrophil kinetics in health and disease. Trends Immunol. 2010;31(8):318–24.
- 91. Kolaczkowska E. The older the faster: aged neutrophils in inflammation. Blood. 2016;128(19):2280–2.
- Uhl B, Vadlau Y, Zuchtriegel G, Nekolla K, Sharaf K, Gaertner F, et al. Aged neutrophils contribute to the first line of defense in the acute inflammatory response. Blood. 2016;128(19):2327–37.
- Mihaila AC, Ciortan L, Macarie RD, Vadana M, Cecoltan S, Preda MB, et al. Transcriptional profiling and functional analysis of N1/N2 neutrophils reveal an immunomodulatory effect of S100A9-blockade on the proinflammatory N1 subpopulation. Front Immunol. 2021;12:708770.
- 94. Ohms M, Moller S, Laskay T. An attempt to polarize human neutrophils toward N1 and N2 phenotypes in vitro. Front Immunol. 2020;11:532.
- Masucci MT, Minopoli M, Carriero MV. Tumor associated neutrophils. Their role in tumorigenesis, metastasis, prognosis and therapy. Front Oncol. 2019;9:1146.
- Sapey E, Stockley JA, Greenwood H, Ahmad A, Bayley D, Lord JM, et al. Behavioral and structural differences in migrating peripheral neutrophils from patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 2011;183(9):1176–86.
- 97. Papayannopoulos V. Neutrophil extracellular traps in immunity and disease. Nat Rev Immunol. 2018;18(2):134–47.
- Barnes PJ, Burney PG, Silverman EK, Celli BR, Vestbo J, Wedzicha JA, et al. Chronic obstructive pulmonary disease. Nat Rev Dis Primers. 2015;1:15076.
- Saetta M, Turato G, Facchini FM, Corbino L, Lucchini RE, Casoni G, et al. Inflammatory cells in the bronchial glands of smokers with chronic bronchitis. Am J Respir Crit Care Med. 1997;156(5):1633–9.
- Ding L, Yang J, Zhang C, Zhang X, Gao P. Neutrophils modulate fibrogenesis in chronic pulmonary diseases. Front Med (Lausanne). 2021;8:616200.
- 101. Maestrelli P, Richeldi L, Moretti M, Fabbri LM. Analysis of sputum in COPD. Thorax. 2001;56(6):420–2.
- 102. Ferrari M, Pizzini M, Cazzoletti L, Ermon V, Spelta F, De Marchi S, et al. Circulating eosinophil levels and lung function decline in stable chronic obstructive pulmonary disease: a retrospective longitudinal study. J Bras Pneumol. 2022;48(6):e20220183.
- Singh D, Edwards L, Tal-Singer R, Rennard S. Sputum neutrophils as a biomarker in COPD: findings from the ECLIPSE study. Respir Res. 2010;11(1):77.
- Barnes PJ. Therapeutic approaches to asthma-chronic obstructive pulmonary disease overlap syndromes. J Allergy Clin Immunol. 2015;136(3):531–45.
- Mukaida N. Pathophysiological roles of interleukin-8/CXCL8 in pulmonary diseases. Am J Physiol Lung Cell Mol Physiol. 2003;284(4):L566–77.

- Plumb J, Gaffey K, Kane B, Malia-Milanes B, Shah R, Bentley A, et al. Reduced glucocorticoid receptor expression and function in airway neutrophils. Int Immunopharmacol. 2012;12(1):26–33.
- Koch A, Gaczkowski M, Sturton G, Staib P, Schinköthe T, Klein E, et al. Modification of surface antigens in blood CD8+ T-lymphocytes in COPD: effects of smoking. Eur Respir J. 2007;29(1):42–50.
- Hogg JC, Chu F, Utokaparch S, Woods R, Elliott WM, Buzatu L, et al. The nature of small-airway obstruction in chronic obstructive pulmonary disease. N Engl J Med. 2004;350(26):2645–53.
- Grumelli S, Corry DB, Song L-Z, Song L, Green L, Huh J, et al. An immune basis for lung parenchymal destruction in chronic obstructive pulmonary disease and emphysema. PLoS Med. 2004;1(1):e8.
- 110. Tham EL, Mescher MF. Signaling alterations in activation-induced nonresponsive CD8 T cells. J Immunol. 2001;167(4):2040–8.
- 111. Kaur M, Smyth LJ, Cadden P, Grundy S, Ray D, Plumb J, et al. T lymphocyte insensitivity to corticosteroids in chronic obstructive pulmonary disease. Respir Res. 2012;13(1):20.
- 112. Southworth T, Metryka A, Lea S, Farrow S, Plumb J, Singh D. IFN-γ synergistically enhances LPS signalling in alveolar macrophages from COPD patients and controls by corticosteroid-resistant STAT1 activation. Br J Pharmacol. 2012;166(7):2070–83.
- Tesfaigzi Y, Curtis JL, Petrache I, Polverino F, Kheradmand F, Adcock IM, et al. Does COPD originate from different cell types? Am J Respir Cell Mol Biol. 2023. https://doi.org/10.1165/rcmb.2023-0175PS.
- 114. Knobloch J, Jungck D, Kronsbein J, Stoelben E, Ito K, Koch A. LABAs and p38MAPK inhibitors reverse the corticosteroid-insensitivity of IL-8 in airway smooth muscle cells of COPD. J Clin Med. 2019;8(12):2058.
- Adcock IM, Barnes PJ. Molecular mechanisms of corticosteroid resistance. Chest. 2008;134(2):394–401.
- Mullan CS, Riley M, Clarke D, Tatler A, Sutcliffe A, Knox AJ, et al. β-Tryptase regulates IL-8 expression in airway smooth muscle cells by a PAR-2–independent mechanism. Am J Respir Cell Mol Biol. 2008;38(5):600–8.
- Knobloch J, Peters H, Jungck D, Mueller K, Strauch J, Koch A. TNFαinduced GM-CSF release from human airway smooth muscle cells depends on activation of an ET-1 autoregulatory positive feedback mechanism. Thorax. 2009;64(12):1044–52.
- 118. Knobloch J, Sibbing B, Jungck D, Lin Y, Urban K, Stoelben E, et al. Resveratrol impairs the release of steroid-resistant inflammatory cytokines from human airway smooth muscle cells in chronic obstructive pulmonary disease. J Pharmacol Exp Ther. 2010;335(3):788–98.
- 119. Rahman I. Antioxidant therapeutic advances in COPD. Ther Adv Respir Dis. 2008;2(6):351–74.
- Fang L, Zhang M, Li J, Zhou L, Tamm M, Roth M. Airway smooth muscle cell mitochondria damage and mitophagy in COPD via ERK1/2 MAPK. Int J Mol Sci. 2022;23(22):13987.
- Michaeloudes C, Kuo CH, Haji G, Finch DK, Halayko AJ, Kirkham P, et al. Metabolic re-patterning in COPD airway smooth muscle cells. Eur Respir J. 2017;50(5):1700202.
- 122. Claesson-Welsh L, Dejana E, McDonald DM. Permeability of the endothelial barrier: identifying and reconciling controversies. Trends Mol Med. 2021;27(4):314–31.
- Amersfoort J, Eelen G, Carmeliet P. Immunomodulation by endothelial cells - partnering up with the immune system? Nat Rev Immunol. 2022;22(9):576–88.
- Amsellem V, Gary-Bobo G, Marcos E, Maitre B, Chaar V, Validire P, et al. Telomere dysfunction causes sustained inflammation in chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 2011;184(12):1358–66.
- 125. Houben JM, Mercken EM, Ketelslegers HB, Bast A, Wouters EF, Hageman GJ, et al. Telomere shortening in chronic obstructive pulmonary disease. Respir Med. 2009;103(2):230–6.
- Li H, Yang T, Li FY, Ning Q, Sun ZM. TLR4 overexpression inhibits endothelial PAS domain-containing protein 1 expression in the lower respiratory tract of patients with chronic COPD. Cell Physiol Biochem. 2016;39(2):685–92.
- Baglole CJ, Reddy SY, Pollock SJ, Feldon SE, Sime PJ, Smith TJ, et al. Isolation and phenotypic characterization of lung fibroblasts. Methods Mol Med. 2005;117:115–27.
- Barron L, Gharib SA, Duffield JS. Lung pericytes and resident fibroblasts: busy multitaskers. Am J Pathol. 2016;186(10):2519–31.

- 129. Willis BC, duBois RM, Borok Z. Epithelial origin of myofibroblasts during fibrosis in the lung. Proc Am Thorac Soc. 2006;3(4):377–82.
- Becerril C, Montano M, Cisneros J, Mendoza-Milla C, Pardo A, Ortiz-Quintero B, et al. Mesenchymal-epithelial transition in fibroblasts of human normal lungs and interstitial lung diseases. Biomolecules. 2021;11(3):378.
- 131. Spagnolo P, Semenzato U. Revealing the pathogenic and ageingrelated mechanisms of the enigmatic idiopathic pulmonary fibrosis (and chronic obstructive pulmonary disease). Curr Opin Pulm Med. 2022;28(4):296–302.
- Sohal SS, Reid D, Soltani A, Ward C, Weston S, Muller HK, et al. Evaluation of epithelial mesenchymal transition in patients with chronic obstructive pulmonary disease. Respir Res. 2011;12(1):130.
- 133. Zhu L, Xu F, Kang X, Zhou J, Yao Q, Lin Y, et al. The antioxidant N-acetylcysteine promotes immune response and inhibits epithelialmesenchymal transition to alleviate pulmonary fibrosis in chronic obstructive pulmonary disease by suppressing the VWF/p38 MAPK axis. Mol Med. 2021;27(1):97.
- 134. Pelaia C, Vatrella A, Gallelli L, Lombardo N, Sciacqua A, Savino R, et al. Role of p38 mitogen-activated protein kinase in asthma and COPD: pathogenic aspects and potential targeted therapies. Drug Des Devel Ther. 2021;15:1275–84.
- Barnes PJ, Baker J, Donnelly LE. Cellular senescence as a mechanism and target in chronic lung diseases. Am J Respir Crit Care Med. 2019;200(5):556–64.
- 136. Herath D, Even B, Oranger M, Foresti R, Papy D, Boyer L, et al. Secreted phospholipase A2 XIIA triggers a mitochondrial damage-induced senescence in chronic obstructive pulmonary disease fibroblasts. Free Radic Biol Med. 2023;205:129–40.
- Chilosi M, Carloni A, Rossi A, Poletti V. Premature lung aging and cellular senescence in the pathogenesis of idiopathic pulmonary fibrosis and COPD/emphysema. Transl Res. 2013;162(3):156–73.
- Karakiulakis G, Papakonstantinou E, Aletras AJ, Tamm M, Roth M. Cell type-specific effect of hypoxia and platelet-derived growth factor-BB on extracellular matrix turnover and its consequences for lung remodeling. J Biol Chem. 2007;282(2):908–15.
- Ning W, Dong Y, Sun J, Li C, Matthay MA, Feghali-Bostwick CA, et al. Cigarette smoke stimulates matrix metalloproteinase-2 activity via EGR-1 in human lung fibroblasts. Am J Respir Cell Mol Biol. 2007;36(4):480–90.
- 140. Birbrair A. Pericyte biology: development, homeostasis, and disease. Adv Exp Med Biol. 2018;1109:1–3.
- 141. Yao H, Edirisinghe I, Rajendrasozhan S, Yang SR, Caito S, Adenuga D, et al. Cigarette smoke-mediated inflammatory and oxidative responses are strain-dependent in mice. Am J Physiol Lung Cell Mol Physiol. 2008;294(6):L1174–86.
- Dong S, Zhang X, He Y, Xu F, Li D, Xu W, et al. Synergy of IL-27 and TNF-alpha in regulating CXCL10 expression in lung fibroblasts. Am J Respir Cell Mol Biol. 2013;48(4):518–30.
- 143. Huang C, Xie M, He X, Gao H. Activity of sputum p38 MAPK is correlated with airway inflammation and reduced FEV1 in COPD patients. Med Sci Monit. 2013;19:1229.
- 144. Patel NR, Cunoosamy DM, Fagerås M, Taib Z, Asimus S, Hegelund-Myrbäck T, et al. The development of AZD7624 for prevention of exacerbations in COPD: a randomized controlled trial. Int J Chron Obstruct Pulmon Dis. 2018;13:1009.
- Banerjee A, Koziol-White C, Panettieri R Jr. p38 MAPK inhibitors, IKK2 inhibitors, and TNFa inhibitors in COPD. Curr Opin Pharmacol. 2012;12(3):287–92.
- 146. Charron CE, Russell P, Ito K, Lea S, Kizawa Y, Brindley C, et al. RV568, a narrow-spectrum kinase inhibitor with p38 MAPK-α and-γ selectivity, suppresses COPD inflammation. Eur Respir J. 2017;50(4):1700188.
- Amano H, et al. p38 Mitogen-activated protein kinase accelerates emphysema in mouse model of chronic obstructive pulmonary disease. J Recept Signal Transduct Res. 2014;34(4):299–306.
- Marumo S, et al. p38 mitogen-activated protein kinase determines the susceptibility to cigarette smoke-induced emphysema in mice. BMC Pulm Med. 2014;14(1):79.
- 149. Lomas DA, Lipson DA, Miller BE, Willits L, Keene O, Barnacle H, et al. An oral inhibitor of p38 MAP kinase reduces plasma fibrinogen in

patients with chronic obstructive pulmonary disease. J Clin Pharmacol. 2012;52(3):416–24.

- Pelaia C, Vatrella A, Sciacqua A, Terracciano R, Pelaia G. Role of p38-mitogen-activated protein kinase in COPD: pathobiological implications and therapeutic perspectives. Expert Rev Respir Med. 2020;14(5):485–91.
- 151. Wu H, Yang S, Wu X, Zhao J, Zhao J, Ning Q, et al. Interleukin-33/ ST2 signaling promotes production of interleukin-6 and interleukin-8 in systemic inflammation in cigarette smoke-induced chronic obstructive pulmonary disease mice. Biochem Biophys Res Commun. 2014;450(1):110–6.
- 152. Feng H, Li M, Altawil A, Yin Y, Zheng R, Kang J. Cigarette smoke extracts induce apoptosis in Raw264.7 cells via endoplasmic reticulum stress and the intracellular Ca(2+)/P38/STAT1 pathway. Toxicol In Vitro. 2021;77:105249.
- 153. Jiang JJ, Chen SM, Li HY, Xie QM, Yang YM. TLR3 inhibitor and tyrosine kinase inhibitor attenuate cigarette smoke/poly I:C-induced airway inflammation and remodeling by the EGFR/TLR3/MAPK signaling pathway. Eur J Pharmacol. 2021;890:173654.
- Lee KH, Lee J, Jeong J, Woo J, Lee CH, Yoo CG. Cigarette smoke extract enhances neutrophil elastase-induced IL-8 production via proteinaseactivated receptor-2 upregulation in human bronchial epithelial cells. Exp Mol Med. 2018;50(7):1–9.
- 155. Fischer S, Koeberle SC, Laufer SA. p38α mitogen-activated protein kinase inhibitors, a patent review (2005–2011). Expert Opin Ther Pat. 2011;21(12):1843–66.
- Norman P. Investigational p38 inhibitors for the treatment of chronic obstructive pulmonary disease. Expert Opin Investig Drugs. 2015;24(3):383–92.
- 157. Novartis A. 5-membered heterocyclebased P38 kinase inhibitors. US20120264801. 2012.
- Strâmbu IR, Kobalava ZD, Magnusson BP, MacKinnon A, Parkin JM. Phase II study of single/repeated doses of acumapimod (BCT197) to treat acute exacerbations of COPD. COPD. 2019;16(5–6):344–53.
- 159. Fisk M, Mohan D, Cheriyan J, Yang L, Fuld J, McEniery C, et al. Evaluation of losmapimod in patients with chronic obstructive pulmonary disease (COPD) with systemic inflammation stratified using fibrinogen ('EVOLU-TION'): rationale and protocol. Artery Res. 2014;8(1):24–34.
- Pascoe S, Costa M, Marks-Konczalik J, McKie E, Yang S, Scherbovsky PS. Biological effects of p38 MAPK inhibitor losmapimod does not translate to clinical benefits in COPD. Respir Med. 2017;130:20–6.
- Millan DS. What is the potential for inhaled p38 inhibitors in the treatment of chronic obstructive pulmonary disease? Future Med Chem. 2011;3(13):1635–45.
- 162. Chopra P, Kanoje V, Semwal A, Ray A. Therapeutic potential of inhaled p38 mitogen-activated protein kinase inhibitors for inflammatory pulmonary diseases. Expert Opin Investig Drugs. 2008;17(10):1411–25.
- Coello C, Fisk M, Mohan D, Brown A, Murphy P, Wikinson I, et al. Quantitative kinetic modelling of FDG in lung imaging of COPD. J Nucl Med. 2015;56(supplement 3):378. https://jnm.snmjournals.org/content/56/ supplement_3/378.short.
- 164. Strâmbu I, Parkin JM, Magnusson B, MacKinnon A. Treatment of acute exacerbations in COPD: an exploratory Phase II study of single and repeated doses of acumapimod (BCT197), an oral p38 inhibitor. InA33. CLINICAL STUDIES IN OBSTRUCTIVE LUNG DISEASE 2017 May (pp. A1333-A1333). American Thoracic Society. https://www.atsjournals. org/doi/abs/10.1164/ajrccm-conference.2017.195.1_MeetingAbstracts. A1333.
- 165. Mariotti F, et al. Safety, tolerability, and pharmacokinetics of single and repeat ascending doses of CHF6001, a novel inhaled phosphodiesterase-4 inhibitor: two randomized trials in healthy volunteers. Int J Chron Obstruct Pulmon Dis. 2018;13:3399–410.
- 166. Make BJ, Patel N, Cunoosamy D, Brown MN, Fageras M, Lundin S, Asimus S, Palmer R, Taib Z, Jansson P, Rennard SI. A randomized controlled trial of AZD7624, an inhaled p38 MAP kinase inhibitor, in COPD patients on ICS/LABA with a history of frequent exacerbations. InC95. COPD ADVANCES IN THERAPY 2017 May (pp. A6741-A6741). American Thoracic Society. https://www.atsjournals.org/doi/abs/10.1164/ajrccmconference.2017.195.1_MeetingAbstracts.A6741.
- Charron C, Coates M, Ito K, Vuppusetty C, To M, Murray J, Strong P, Rapeport G. Superior Effects Of RV568, A Novel Narrow Spectrum Kinase Inhibitor, To BIRB796, A P38MAPK Inhibitor, On Pro-Inflammatory

Cytokine Roduction In Macrophages. InB46. INHALED AND SYSTEMIC ANTI-INFLAMMATORY THERAPY IN COPD 2011 May (pp. A3088-A3088). American Thoracic Society. https://www.atsjournals.org/doi/abs/10. 1164/ajrccm-conference.2011.183.1_MeetingAbstracts.A3088.

- 168. Knobloch J, Urban K, Jungck D, Stoelben E, Ito K, Koch A. Effects Of RV1088, A Narrow Spectrum Kinase Inhibitor, On Pro-Inflammatory Cytokine Production In Smooth Muscle Cells Obtained From COPD Patients. InB69. NOVEL INSIGHTS INTO AIRWAY INFLAMMATION AND REMODELING IN ASTHMA AND COPD 2011 May (pp. A3616-A3616). American Thoracic Society. https://www.atsjournals.org/doi/abs/10. 1164/ajrccm-conference.2011.183.1_MeetingAbstracts.A3616.
- 169. Woodward K, Brindley C, Warrington S, Rapeport G, Cass L. Tolerability and pharmacokinetic profile of Rv568, a narrow spectrum kinase inhibitor, following single and repeat intranasal dosing in healthy volunteers. Am J Respir Crit Care Med. 2013;187:A4118.
- 170. Russell P, Cass L, Ito K, Charron C, Strong P, Rapeport G, et al. Safety, pharmacokinetic and pharmacodynamic profile of RV568, a narrow spectrum kinase inhibitor, following repeat inhaled dosing in COPD patients. Eur Respir J. 2013;42(Suppl 57):1971. https://erj.ersjournals. com/content/42/Suppl_57/1971.short.
- Charron C, Cass L, Dine G, Strong P, Rapeport G, Ito K. Pharmacodynamic analysis of Rv568, a narrow spectrum kinase inhibitor, following single and repeat intranasal dosing in healthy volunteers. Am J Respir Crit Care Med. 2013;187:A1494.
- ClinicalTrials.gov [Internet]. Pfizer: Identifier NCT02366637, An evaluation of PF-03715455 in moderate to severe chronic obstructive pulmonary disease. 2016. Available from: https://clinicaltrials.gov/study/ NCT02366637.
- 173. Fisk M, Cheriyan J, Mohan D, Forman J, Maki-Petaja KM, McEniery CM, et al. The p38 mitogen activated protein kinase inhibitor losmapimod in chronic obstructive pulmonary disease patients with systemic inflammation, stratified by fibrinogen: a randomised double-blind placebocontrolled trial. PLoS One. 2018;13(3):e0194197.
- Yu H, Su X, Lei T, Zhang L, Feng Z, Zhang C, et al. Safety and efficacy of p38 mitogen-activated protein kinase inhibitors (MAPKIs) in COPD. Front Pharmacol. 2022;13:950035.
- 175. Higham A, Karur P, Jackson N, Cunoosamy DM, Jansson P, Singh D. Differential anti-inflammatory effects of budesonide and a p38 MAPK inhibitor AZD7624 on COPD pulmonary cells. Int J Chron Obstruct Pulmon Dis. 2018;13:1279–88.

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